









Draft Genome Sequence of the Griseofulvin-Producing Fungus *Xylaria flabelliformis* Strain G536

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ABSTRACT The draft genome of the ascomycete fungus *Xylaria flabelliformis* (previously known as *Xylaria cubensis*) was sequenced using Illumina paired-end technology. The assembled genome is 41.2 Mb long and contains 11,404 genes. This genome will contribute to our understanding of *X. flabelliformis* secondary metabolism and the organism's ability to live as a decomposer as well as an endosymbiont.

The genomes of fungi in the order Xylariales (Sordariomycetes, Ascomycota) contain some of the highest numbers of genes involved in secondary metabolism among fungi (1). *Xylaria flabelliformis* (previously known as *Xylaria cubensis*) (2, 3) is a filamentous fungus in the order Xylariales that lives both as a decomposer of organic matter (4) and as an endosymbiont of plants and lichens (5). *Xylaria flabelliformis* is known to produce the fungistatic compound griseofulvin, an FDA-approved drug that is also considered an “essential medicine” by the World Health Organization (6, 7). To better understand the secondary metabolism and evolution of *X. flabelliformis*, we sequenced the genome of a representative strain.

Xylaria flabelliformis strain G536 (8) was grown on liquid yeast extract soy peptone dextrose (YESD) medium. After 7 days, the mycelium was filtered through a sterile filter, retrieving the mycelial mass, which was then ground to a fine powder with a sterile mortar and pestle by using liquid nitrogen. The fine powder was then transferred to a bashing bead tube with DNA lysis buffer from the Zymo Quick-DNA fungal/bacterial miniprep kit (catalog number D6005). The powder in the bashing bead tube was further disrupted and homogenized in a Qiagen TissueLyser LT bead mill for 5 min. Genomic DNA was extracted using procedures outlined in the Zymo Quick-DNA fungal/bacterial miniprep kit and sonicated to a size of ~550 bp. A sequencing library was constructed using the Illumina TruSeq library preparation method. Paired-end sequencing (300 bp from each end) was performed on an Illumina MiSeq version 3 instrument run at HudsonAlpha Discovery (Huntsville, AL), producing a total of 23,234,771 paired-end reads.

The raw reads were trimmed of adapter and low-quality sequences with Trimmomatic version 0.36 (9) by using a custom list of adapter sequences (see the Figshare document at <https://www.doi.org/10.6084/m9.figshare.8986505>) and the parameters “ILLUMINACLIP:2:30:10 LEADING:3 TRAILING:10 SLIDINGWINDOW:4:15 MINLEN:50.” The reads were then *de novo* assembled with SPAdes version 3.13.1 (10) using the “-careful” and “-cov-cutoff auto” options. The final genome assembly consisted of 41,150,291 bp spread over 155 scaffolds (164 contigs), with an N_{50} value of 488,275 bp and a GC content of 47.44%. Gene prediction was performed using AUGUSTUS version 3.3.2 (11), with *Histoplasma capsulatum* as the training species and the settings “-minexonintronprob=0.1,” “-minmeanexonintronprob=0.4,” and “-noInFrameStop=True.” The genome contained 11,404 predicted protein-coding genes. Analyses of the predicted

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protein product sequences with BUSCO version 3.1.0 (12) and the sodariomycete_odb9 database showed that the proteome contained 93.6% of the BUSCOs as complete proteins. The annotation was converted to an SQN file by using the NCBI-provided script tbl2asn and submitted to GenBank.

To gain insights into *X. flabelliformis* secondary metabolism, we predicted biosynthetic gene clusters with antiSMASH version 4.1.0 (13) using the “–taxon fungi” option. A total of 86 putative biosynthetic gene clusters were predicted, including clusters likely to produce both griseofulvin and cytochalasin, two metabolites known to be produced by *X. flabelliformis* G536 (8, 14). Tables summarizing the antiSMASH results can be found in the Figshare document at <https://www.doi.org/10.6084/m9.figshare.8986505>. Our work represents a major step toward understanding the evolution and biochemical output of this fungus.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [VFLP0000000](https://www.doi.org/10.6084/m9.figshare.8986505). The version described in this paper is VFLP01000000. The Illumina raw reads have been deposited at the Sequence Read Archive under accession number [SRX5939388](https://www.doi.org/10.6084/m9.figshare.8986505).

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REFERENCES

- Helaly SE, Thongbai B, Stadler M. 2018. Diversity of biologically active secondary metabolites from endophytic and saprotrophic fungi of the ascomycete order Xylariales. *Nat Prod Rep* 35:992–1014. <https://doi.org/10.1039/c8np00010g>.
- Ju Y-M, Hsieh H-M, Dominick S. 2016. The *Xylaria* names proposed by C. G. Lloyd. *N Am Fungi* 11:1–31.
- Réblová M, Miller AN, Rossman AY, Seifert KA, Crous PW, Hawksworth DL, Abdel-Wahab MA, Cannon PF, Daranagama DA, De Beer ZW, Huang S-K, Hyde KD, Jayawardena R, Jaklitsch W, Jones EBG, Ju Y-M, Judith C, Maharachchikumbura SSN, Pang K-L, Petrini LE, Raja HA, Romero AI, Shearer C, Senanayake IC, Voglmayr H, Weir BS, Wijayawardena NN. 2016. Recommendations for competing sexual-asexually typified generic names in Sordariomycetes (except Diaporthales, Hypocreales, and Magnaporthales). *IMA Fungus* 7:131–153. <https://doi.org/10.5598/ima fungus.2016.07.01.08>.
- Rogers JD. 1984. *Xylaria cubensis* and its anamorph *Xylocoremium flabelliforme*, *Xylaria allantoides*, and *Xylaria poitei* in continental United States. *Mycologia* 76:912–923. <https://doi.org/10.2307/3793147>.
- U’Ren JM, Miadlikowska J, Zimmerman NB, Lutzoni F, Stajich JE, Arnold AE. 2016. Contributions of North American endophytes to the phylogeny, ecology, and taxonomy of Xylariaceae (Sordariomycetes, Ascomycota). *Mol Phylogenet Evol* 98:210–232. <https://doi.org/10.1016/j.ympev.2016.02.010>.
- Petersen AB, Rønneest MH, Larsen TO, Clausen MH. 2014. The chemistry of griseofulvin. *Chem Rev* 114:12088–12107. <https://doi.org/10.1021/cr400368e>.
- World Health Organization. 2017. The selection and use of essential medicines. World Health Organization, Geneva, Switzerland.
- Sica VP, Rees ER, Tchegnon E, Bardsley RH, Raja HA, Oberlies NH. 2016. Spatial and temporal profiling of griseofulvin production in *Xylaria cubensis* using mass spectrometry mapping. *Front Microbiol* 7:544. <https://doi.org/10.3389/fmicb.2016.00544>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Stanke M, Schöffmann O, Morgenstern B, Waack S. 2006. Gene prediction in eukaryotes with a generalized hidden Markov model that uses hints from external sources. *BMC Bioinformatics* 7:62. <https://doi.org/10.1186/1471-2105-7-62>.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>.
- Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. *Nucleic Acids Res* 45:W36–W41. <https://doi.org/10.1093/nar/gkx319>.
- Knowles SL, Raja HA, Wright AJ, Lee AML, Caesar LK, Cech NB, Mead ME, Steenwyk JL, Ries LNA, Goldman GH, Rokas A, Oberlies NH. 2019. Mapping the fungal battlefield: using *in situ* chemistry and deletion mutants to monitor interspecific chemical interactions between fungi. *Front Microbiol* 10:285. <https://doi.org/10.3389/fmicb.2019.00285>.