

# Is a Picture Worth 1000 Calories: The Neuroimaging of Obesity

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## Abstract

In healthy weight individuals, complex brain circuits interact with peripheral feeding signals to control feeding behavior, and it is thought that the dysregulation of these circuits can lead to excessive food intake and obesity. Human neuroimaging studies have shown BMI-dependent deficits in dopamine neurotransmission encoding reward, suggesting a “hypodopaminergic reward deficiency” whereby obese individuals overeat to compensate for a hypofunctioning reward circuitry. However, other imaging studies demonstrate hyperactivation of dopamine networks that positively correlate with BMI in obese individuals. Animal studies link these seemingly opposing theories, revealing that insulin promotes the intracellular trafficking and surface expression of the dopamine transporter (DAT) while inhibiting that of the norepinephrine transporter (NET). Together these transporters control dopamine levels in the striatum and cortex respectively, areas critically involved in reward, habits, and cognitive control. The purpose of this review is to integrate the molecular aspects of food overconsumption and obesity with human neuroimaging data, focusing on the role and dysregulation of dopamine in the neural circuits subserving food intake.

## An Introduction to the Obesity Epidemic

The fundamental neurocircuitry of the homeostatic feeding system and its interactions with peripheral feeding signals to modulate appetitive behavior and energy expenditure around a physiologic set point has maintained a relatively stable human body composition until only recently, when the prevalence of obesity has increased dramatically<sup>1</sup>. The rapid elevation in obesity over the past generation, with nearly seventy percent of the United States population meeting criteria for being overweight<sup>2</sup>, suggests environmental factors play a key role. Current research indicates the presence and dysfunction of expanded neural circuits controlling reward, habits, and decision-making may mediate feeding behavior and subsequent overconsumption, contributing to the obesity epidemic<sup>3-7</sup>.

Animal research has been critical for elucidating molecular aspects of obesity, with studies showing that food overconsumption is both driven and paralleled by broad changes in dopaminergic circuitry. Indeed, an overarching question in the field is how the physiologic response to food consumption augments brain dopaminergic circuits that enable the progression and maintenance of obesity. Neuroimaging is an important and novel tool for non-invasively examining the structural, molecular, and functional correlates of obesity<sup>8</sup>. The purpose of this review is to integrate the molecular aspects of food overconsumption and obesity

with human neuroimaging data, focusing on the role and dysregulation of dopamine in the neural circuits subserving food intake.

## Molecular Aspects of Dopamine in Obesity

### *Homeostatic Feeding, Dopamine, Reward*

The hypothalamus regulates homeostatic feeding (i.e. food consumption for the purpose of maintaining energy balance; for review, see<sup>1, 9, 10</sup>), by responding to peripheral hormonal signals relaying information about the body's energy state<sup>11, 12</sup>. The anorexigenic gut peptides leptin and insulin, negative feedback adiposity signals circulating in proportion to body fat mass, indicate a positive energy balance while the orexigenic gut peptide ghrelin, whose levels inversely correlate with adiposity, signals a negative energy balance. In addition to their homeostatic action in the hypothalamus to regulate future feeding behavior, these peripheral hormonal signals also act on the mesolimbic dopamine system. Activity in mesolimbic reward circuitry (for review, see<sup>13</sup>), an area that is acutely activated with all drugs of abuse<sup>14</sup>, implies that feeding signals operate outside of brain circuits subserving homeostatic feeding and that feeding itself may have rewarding properties.

Current evidence suggests that gut peptides signaling a positive energy balance function to negatively modulate midbrain dopamine (DA) neurotransmission and food

## Keywords

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## CANDIDATE REVIEWS

reward while those signaling a negative energy balance are positive DA modulators. For example, as determined by both electrophysiology and receptor knockout studies, leptin acts directly on the DA neurons of the ventral tegmental area (VTA) to inhibit action potential firing<sup>15, 16</sup> and reduce food intake<sup>15</sup> and reward-seeking behaviors<sup>17, 18</sup>. In contrast, ghrelin activates VTA DA neurons, triggering feeding<sup>19</sup>. New research points to a critical role for insulin in the regulation of reward circuitry. Insulin promotes the intracellular trafficking and surface expression of the dopamine transporter (DAT) via the PI3K/Akt signaling pathway, regulating the high-affinity uptake of dopamine from the mesolimbic synapse<sup>20-23</sup> while reducing food-intake<sup>17</sup>. Further, dopamine receptor (D2R) expression<sup>24</sup> is impaired in insulin-depleted states, suggesting a hypofunctioning of the dopamine reward system with insulin resistance. These findings together demonstrate the role of peripheral feeding signals, particularly insulin, in fine-tuning extracellular synaptic dopamine in the reward circuitry and subsequently influencing feeding behavior.

An understanding of mesolimbic dopamine's function and behavioral correlates is critical for discerning the role of dopamine dysregulation in obesity. In the mesolimbic circuitry, dopamine encodes the expectation of, motivation for, and approach behaviors seeking reward<sup>13, 25, 26</sup>, all processes which are "hijacked" in the early stages of addiction<sup>14, 27</sup>. Consistent with dopamine's role in reward, dopamine levels are elevated during food seeking<sup>28, 29</sup>, exposure to and consumption of novel food stimuli<sup>30, 31</sup>, and daily intermittent consumption of both sugar<sup>32-34</sup> and fat<sup>35, 36</sup>. Further, it is the phasic firing of these dopamine neurons that encodes this food reward<sup>26, 37-39</sup>. In contrast, evoked dopamine release, basal dopamine levels<sup>35, 40, 41</sup>, and D2R availability<sup>24, 42</sup> are blunted in chronic obesity. One study links these two states, demonstrating increased basal DA and DA efflux in obesity-prone young, insulin-sensitive rats in the mesolimbic reward system but decreased basal DA and DA efflux in obesity-prone, adult, insulin-resistant rats<sup>43</sup>. These results, combined with evidence that short-term elevations in insulin or glucose increase basal DA<sup>44</sup> while decreasing D2R<sup>42, 45</sup>, provides evidence for the progressive nature of dopamine dysregulation in obesity.

According to the dopamine reward hypothesis, dopamine signaling in the mesolimbic system encodes reward and promotes reward-seeking behavior; consequently, impaired dopamine signaling will focus and drive behaviors aimed at restoring dopamine tone<sup>6, 46</sup>. It is hypothesized that the blunted dopamine signaling in obesity may attenuate the rewarding aspects of food, a hypodopaminergic reward

deficiency syndrome (HRDS), leading obese individuals to consume increasing quantities of palatable food to achieve the same level of reward<sup>41, 47</sup>. A problem with this "reward deficiency" hypothesis, however, is that decreased perceived reward might be expected to suppress rather than promote excessive feeding. An alternative view is that reduced dopamine receptor availability may be a consequence, rather than a cause, of obesity due to elevated dopamine levels from excessive food intake and/or abnormal food seeking<sup>4, 5, 28-31, 37</sup>. Several studies have, in fact, demonstrated a hyperresponsiveness to reward in the mesolimbic circuitry in obesity<sup>5, 48, 49</sup> corroborating this hypothesis. Indeed, dysregulation of dopamine circuitry is a clear component of obesity, but the exact nature of the dysregulation remains undefined.

### *Food-Seeking, Habits, and Addiction*

Despite mesolimbic dopamine having a clear role in reward and feeding behavior, studies in dopamine deficient mice (a severely hypoactive phenotype which will die of starvation without supplemented dopamine) show that viral restoration of dopamine to the nucleus accumbens does not restore feeding behavior<sup>6, 50</sup>. However, restoration of dopamine to the dorsal striatum, specifically the dorsolateral striatum, rescues the dopamine-deficient phenotype and induces feeding<sup>51-53</sup>. These results suggest a role for dopamine action outside the mesolimbic reward system in feeding behavior. In fact, it is the dorsal striatum that mediates goal-directed behaviors and habit formation such as the repeated seeking of reward-conditioned, highly salient, food stimuli<sup>54-56</sup>.

Habits are "sequential, repetitive, motor, or cognitive behaviors elicited by external or internal triggers that, once released, can go to completion without conscious oversight"<sup>55</sup>. Habits begin as goal-directed behaviors, where a salient<sup>57</sup> stimulus is achieved through a specific action sequence, but progress to cue-mediated behaviors with repeated reward training that persist even with reward devaluation<sup>58, 59</sup>. This progression involves an underlying ventral-to-dorsal striatal shift<sup>14, 55, 60</sup> as dopamine-directed reward behaviors of the ventral striatum are replaced by dorsal striatal cue-initiated action sequences<sup>61, 62</sup> mediated by multiple neurotransmitters that do not appear to be under the regulatory influence of insulin. Indeed, this shift is well defined with food reward, indicating that salient foods and their cues are sufficient to initiate reward-seeking and the subsequent habitual behaviors characteristic of addiction<sup>14, 63</sup>.

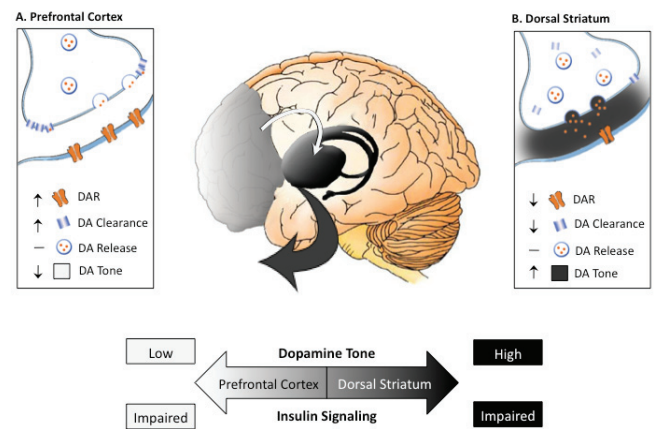
### *Decision-Making and Disinhibition*

The progression from reward learning to habit formation relies on active oversight by the prefrontal cortex (PFC)<sup>64</sup>, a region responsible for ‘top-down’ regulation of subcortical function to promote situation-appropriate and task-relevant behaviors<sup>65</sup>. While the complexities of PFC function are beyond the scope of this article (for review, see <sup>66-68</sup>), there is strong evidence for the specific role of dopamine in regulating PFC activity<sup>69, 70</sup> through volume transmission maintaining extrasynaptic dopamine tone<sup>70</sup>. Dopamine appears to improve prefrontal cortical cognitive function<sup>71-73</sup> by enhancing glutamatergic signaling through D1 receptor binding<sup>74, 75</sup>, however this effect is non-linear where either too much<sup>76</sup> or too little<sup>77, 78</sup> dopamine actually impairs proper PFC function. This non-linear impairment is readily seen in measures of response inhibition, where both deficits<sup>79-82</sup> and elevations<sup>83</sup> in central dopamine produce a faster cue-driven response and/or a decreased ability to rapidly inhibit unwanted responses.

Dopamine tone is maintained in the prefrontal cortex by the norepinephrine transporter (NET)<sup>84, 85</sup> whose intracellular trafficking and surface expression, in contrast to the striatal dopamine transporter, is inhibited by insulin<sup>86</sup>. Insulin further inhibits dopamine release in the PFC<sup>87</sup>, thus providing multiple mechanisms that would both serve to diminish cortical dopamine levels. As dopamine acts through an inverted-U response<sup>88</sup>, even minor deviations from optimal tone can alter PFC function<sup>70</sup>. In the setting of impaired insulin signaling, such dysregulation may set the stage for the emergence of the habitual, cue-driven behaviors. Indeed, given that the increased availability of highly palatable food provided by the modern environment requires the continuous inhibition of cue-mediated feeding behaviors, it is easy to see how dopamine-mediated prefrontal disinhibition could unmask the established subcortical salience attributions and response patterns leading to obesity<sup>89</sup>.

### The Progression to Obesity

Here we propose a plausible molecular mechanism by which the physiologic response to food consumption promotes progressive neuroadaptations in brain dopaminergic circuits subserving reward, habits, and decision-making that further bias towards the maintenance of obesity. Insulin maintains dopamine homeostasis in reward circuitry, supporting a synaptic environment ideal for the perception of food reward. The onset of mild insulin resistance with repeated consumption of highly palatable food drives a striatal synaptic hyperdopaminergia from increased dopamine release, decreased dopamine clearance, and allostatic



**Figure 1.** Model for Dopamine Neurotransmission in the context of impaired insulin signaling in A) prefrontal cortex and B) dorsal striatum

downregulation of dopamine receptor function, effectively blunting the impact of dopamine reward signaling and facilitating the emergence of cue-driven food seeking behavior. Further, concomitant insulin-mediated cortical neuroadaptations promote a prefrontal hypodopaminergic tone serving to unmask response patterns directed at palatable food acquisition and consumption (see Figure 1). In the next section, we review how available neuroimaging evidence supports this model and identify important next steps for neuroimaging in unraveling obesity pathogenesis and implications for treatment.

### Translating Molecules to Systems

#### Neuroimaging in Obesity

Exploring feeding behavior with molecular (PET) and functional (MR) neuroimaging facilitates the systems-level study of feeding behavior and translation of molecular obesity research to humans. Such methods have in fact convincingly uncovered strong evidence for widespread neural dysregulation in obesity. The multisensory elements of food are reflected as its unified flavor<sup>90</sup> which, combined with individual and environmental factors, contributes to the pleasure derived from food consumption and its hedonic value<sup>91</sup>. In healthy individuals, flavor perception robustly activates the gustatory network, including the thalamus, insula, frontal operculum, inferior frontal gyrus, and orbitofrontal cortex<sup>92-96</sup>. Hunger elicits heightened activity in these areas and further activation in the striatum, midbrain, and prefrontal cortex<sup>95, 96</sup>. This activation correlates with perceived stimulus pleasantness<sup>92, 97</sup> and reward value<sup>95</sup>, providing evidence for the representation of hedonic ‘liking’ in these regions.

Yet feeding behavior extends beyond the mere liking of food, and whether food is sought for consumption depends on how much it is “wanted”<sup>4</sup>. This incentive value is influenced by the sight and smell of food; these sensory experiences reflect food availability and the anticipation of food, which can then be contrasted with the effort to achieve them<sup>91</sup>. Indeed, the presentation of a salient food cue elicits reward associated with food anticipation and subsequent food seeking<sup>91, 98, 99</sup>. Visual food stimuli increase activity in areas including the midbrain, amygdala, dorsal striatum, cingulate, insula, and orbitofrontal cortex<sup>94, 96, 98, 100, 101</sup>, where activity is amplified by hunger<sup>98, 100, 102, 103</sup>. The fasting state also elicits activation of hedonic brain areas including the ventral striatum<sup>102, 104, 105</sup> which implies that fasting enhances the rewarding properties of food<sup>106</sup>, consistent with behavioral studies.

Overweight and obese individuals in the fasting state also demonstrate activation in circuits subserving food reward<sup>107-110</sup>. In contrast to healthy individuals, obese individuals exhibit BMI-dependent potentiation of activation by salient food cues in gustatory areas such as the orbitofrontal cortex and insula, and in brain regions receiving dopaminergic inputs, including dorsal and ventral striatum<sup>107, 108, 110, 111</sup>. Hyperactivity in these areas could represent enhanced expected food reward promoting dopamine release, driving the motivation and behaviors aimed at food consumption<sup>112</sup>. Evidence for greater reward sensitivity in obesity<sup>5, 113</sup> is consistent with this hypothesis. Alternatively, if the obese state is characterized by prefrontal dysregulation of inhibitory circuits under dopaminergic modulation, the observed hyperactivity results from the unmasking of habitual circuits normally under cortical regulation. One fMRI study examining response inhibition in adolescent girls demonstrated a BMI-dependent loss of activity in the prefrontal areas subserving inhibitory control<sup>114</sup>, however the relationship of this attenuation to subcortical activity was not assessed. An important next step will be to determine how prefrontal functional brain activity changes with subcortical activity in light of differential dopamine clearance in these regions.

While fMRI studies indicate neural dysfunction in obesity, they do not assess the mechanism by which it occurs. Molecular imaging evidence directly demonstrating dopamine dysregulation comes from a small number of PET studies finding BMI-dependent decreases in striatal dopamine D2 receptor availability<sup>47, 115</sup>. This reduction depends on the magnitude and duration of overfeeding<sup>42</sup>, supporting the hypothesis of an allostatic downregulation of D2 receptors with chronic overeating. However, these PET studies assessed D2 receptor availability using the ra-

dioligand [<sup>11</sup>C]-raclopride which competes with synaptic dopamine for receptor binding, and therefore the decreased binding explained as a reduction in D2 receptor availability could also reflect increases in synaptic dopamine. This interpretation would suggest an elevated basal dopamine tone in obesity. Among adults viewing food cues who received an acute methamphetamine dose (stimulating presynaptic dopamine release), normal-weight individuals demonstrated increases in dopamine<sup>116</sup> while the dopamine levels of obese individuals remained constant<sup>117</sup>, providing evidence for blunted dopamine signaling in obesity.

### *Implications for Treatment*

While the precise etiologic nature of dopamine dysregulation in obesity remains unclear, several imaging studies support plasticity in the brain circuits underlying obesity and thus opportunities for treatment. Initial clinical observations of mild weight loss in patients receiving treatment with dopamine agonists<sup>118</sup> have been replicated in animal studies<sup>119</sup>, however the cognitive/psychiatric side effects render these drugs problematic. Further, dopamine administration will be ineffective if post-synaptic dopamine signaling is impaired in obesity. Promising observations come from studies demonstrating that bariatric surgery<sup>120</sup> and weight loss<sup>42</sup> increase D2 receptor levels and decrease functional activity in dopamine reward circuitry<sup>121</sup> while increasing activity in the prefrontal cortex<sup>122</sup>. One explanation for these effects on dopamine circuits is the drastic changes in insulin levels following bariatric surgery; however, there have been no longitudinal controlled clinical trials to examine the direct effect of insulin on normalizing dopamine neurotransmission in obesity. Such research will be critical in understanding the pathogenesis of obesity, potential therapeutic targets in insulin signaling pathways, and future opportunities for treatment.

### **Conclusion**

Recent scientific evidence demonstrating that central nervous system dopamine is under the regulatory influence of insulin offers a plausible mechanism for understanding obesity as a dysregulation of neural systems controlling reward, habits, and decision-making. Here we have reviewed the molecular processes underlying insulin's effect on dopamine circuitry and feeding behavior, and how these findings link the opposing theories of a hypodopaminergic reward deficiency versus a hyperresponsiveness to reward in the obese state. We further extend the interpretations of this research by proposing a novel model for obesity as a progressive disruption of subcortical and prefrontal brain

circuitry initiated and perpetuated by insulin resistance and subsequent dysregulation of extracellular dopamine. While future research is necessary, the hypothesis of insulin's ability to reset central dopamine tone and subsequently reshape feeding behavior offers exciting new opportunities for the clinical management of obesity.

## References

- Niswender KD, Baskin DG and Schwartz MW (2004). Insulin and its evolving partnership with leptin in the hypothalamic control of energy homeostasis. *Trends in endocrinology and metabolism: TEM*. 15 (8): 362-369.
- Flegal KM, Carroll MD, Ogden CL and Curtin LR (2010). Prevalence and trends in obesity among US adults, 1999-2008. *JAMA*. 303 (3): 235-241.
- Adam TC and Epel ES (2007). Stress, eating and the reward system. *Physiology & behavior*. 91 (4): 449-458.
- Berridge KC, Ho CY, Richard JM and DiFeliceantonio AG (2010). The tempted brain eats: pleasure and desire circuits in obesity and eating disorders. *Brain research*. 1350: 43-64.
- Davis C, Strachan S and Berkson M (2004). Sensitivity to reward: implications for overeating and overweight. *Appetite*. 42 (2): 131-138.
- Palmiter RD (2007). Is dopamine a physiologically relevant mediator of feeding behavior? *Trends in neurosciences*. 30 (8): 375-381.
- Volkow ND, Wang GJ and Baler RD (2011). Reward, dopamine and the control of food intake: implications for obesity. *Trends in cognitive sciences*. 15 (1): 37-46.
- Gibson CD, Carnell S, Ochner CN and Geliebter A (2010). Neuroimaging, gut peptides and obesity: novel studies of the neurobiology of appetite. *Journal of neuroendocrinology*. 22 (8): 833-845.
- Schwartz MW, Woods SC, Porte Jr. D, Seeley RJ and Baskin DG (2000). Central nervous system control of food intake. *Nature*. 404: 661-671.
- Morton GJ, Cummings DE, Baskin DG, Barsh GS and Schwartz MW (2006). Central nervous system control of food intake and body weight. *Nature*. 443 (7109): 289-295.
- Moran TH (2006). Gut peptide signaling in the controls of food intake. *Obesity*. 14 Suppl 5: 250S-253S.
- Saper CB, Chou TC and Elmquist JK (2002). The need to feed: Homeostatic and hedonic control of eating. *Neuron*. 36: 199-211.
- Berridge KC and Robinson TE (1998). What is the role of dopamine in reward: hedonic impact, reward learning, or incentive salience? *Brain Research Reviews*. 28 (3): 309-369.
- Koob GF and Volkow ND (2010). Neurocircuitry of addiction. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 35 (1): 217-238.
- Hommel JD, Trinko R, Sears RM, Georgescu D, Liu ZW, Gao XB, Thurmon JJ, Marinelli M and DiLeone RJ (2006). Leptin receptor signaling in midbrain dopamine neurons regulates feeding. *Neuron*. 51 (6): 801-810.
- Fulton S, Pissios P, Manchon RP, Stiles L, Frank L, Pothos EN, Maratos-Flier E and Flier JS (2006). Leptin regulation of the mesoaccumbens dopamine pathway. *Neuron*. 51 (6): 811-822.
- Figlewicz DP, Bennett JL, Naleid AM, Davis C and Grimm JW (2006). Intraventricular insulin and leptin decrease sucrose self-administration in rats. *Physiology & behavior*. 89 (4): 611-616.
- Davis JF, Choi DL, Schurdak JD, Fitzgerald MF, Clegg DJ, Lipton JW, Figlewicz DP and Benoit SC (2011). Leptin regulates energy balance and motivation through action at distinct neural circuits. *Biological psychiatry*. 69 (7): 668-674.
- Abizaid A, Liu ZW, Andrews ZB, Shanabrough M, Borok E, Elsworth JD, Roth RH, Sleeman MW, Picciotto MR, Tschoop MH, Gao XB and Horvath TL (2006). Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *The Journal of clinical investigation*. 116 (12): 3229-3239.
- Carvelli L, Moron JA, Kahlig KM, Ferrer JV, Sen N, Lechleiter JD, Leeb-Lundberg LM, Merrill G, Lafer EM, Ballou LM, Shippenberg TS, Javitch JA, Lin RZ and Galli A (2002). PI 3-kinase regulation of dopamine uptake. *J Neurochem*. 81 (4): 859-869. Seminal paper defining the molecular mechanism for insulin regulation of the dopamine transporter.**
- Daws LC, Avison MJ, Robertson SD, Niswender K, Galli A and Saunders C (2011). Insulin signalling and addiction. *Neuropharmacology*. Ahead of Print.
- Figlewicz DP, Evans SB, Murphy J, Hoen M and Baskin DG (2003). Expression of receptors for insulin and leptin in the ventral tegmental area/substantia nigra (VTA/SN) of the rat. *Brain research*. 964 (1): 107-115.
- Garcia BG, Wei Y, Moron JA, Lin RZ, Javitch JA and Galli A (2005). Akt is essential for insulin modulation of amphetamine-induced human dopamine transporter cell-surface redistribution. *Molecular pharmacology*. 68 (1): 102-109.
- Johnson PM and Kenny PJ (2010). Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nature neuroscience*. 13 (5): 635-641.
- Schultz W (2007). Multiple dopamine functions at different time courses. *Annual review of neuroscience*. 30: 259-288.
- Schultz W, Dayan P and Montague PR (1997). A neural substrate of prediction and reward. *Science*. 275 (5306): 1593-1599.
- Schultz W (2011). Potential vulnerabilities of neuronal reward, risk, and decision mechanisms to addictive drugs. *Neuron*. 69 (4): 603-617.
- Hernandez L and Hoebel BG (1988). Food reward and cocaine increase extracellular dopamine in the nucleus accumbens as measured by microdialysis. *Life sciences*. 42 (18): 1705-1712.
- Salamone JD, Steinpreis RE, McCullough LD, Smith P, Grebel D and Mahan K (1991). Haloperidol and nucleus accumbens dopamine depletion suppress lever pressing for food but increase free food consumption in a novel food choice procedure. *Psychopharmacology*. 104 (4): 515-521.
- Bassareo V and Di Chiara G (1997). Differential influence of associative and nonassociative learning mechanisms on the responsiveness of prefrontal and accumbal dopamine transmission to food stimuli in rats fed ad libitum. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 17 (2): 851-861.
- Bassareo V and Di Chiara G (1999). Modulation of feeding-induced activation of mesolimbic dopamine transmission by appetitive stimuli and its relation to motivational state. *The European journal of neuroscience*. 11 (12): 4389-4397.
- Avena NM, Rada P and Hoebel BG (2008). Evidence for sugar addiction: behavioral and neurochemical effects of intermittent, excessive sugar intake. *Neuroscience and biobehavioral reviews*. 32 (1): 20-39.

## CANDIDATE REVIEWS

33. Avena NM, Rada P, Moise N and Hoebel BG (2006). Sucrose sham feeding on a binge schedule releases accumbens dopamine repeatedly and eliminates the acetylcholine satiety response. *Neuroscience*. 139 (3): 813-820.
34. Rada P, Avena NM and Hoebel BG (2005). Daily bingeing on sugar repeatedly releases dopamine in the accumbens shell. *Neuroscience*. 134 (3): 737-744.
35. Rada P, Bocarsly ME, Barson JR, Hoebel BG and Leibowitz SF (2010). Reduced accumbens dopamine in Sprague-Dawley rats prone to overeating a fat-rich diet. *Physiology & behavior*. 101 (3): 394-400.
36. Liang NC, Hajnal A and Norgren R (2006). Sham feeding corn oil increases accumbens dopamine in the rat. *American journal of physiology Regulatory, integrative and comparative physiology*. 291 (5): R1236-1239.
37. Roitman MF, Stuber GD, Phillips PE, Wightman RM and Carelli RM (2004). Dopamine operates as a subsecond modulator of food seeking. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 24 (6): 1265-1271.
38. Roitman MF, Wheeler RA and Carelli RM (2005). Nucleus accumbens neurons are innately tuned for rewarding and aversive taste stimuli, encode their predictors, and are linked to motor output. *Neuron*. 45 (4): 587-597.
39. Roitman MF, Wheeler RA, Wightman RM and Carelli RM (2008). Real-time chemical responses in the nucleus accumbens differentiate rewarding and aversive stimuli. *Nature neuroscience*. 11 (12): 1376-1377.
40. Geiger BM, Behr GG, Frank LE, Caldera-Siu AD, Beinfeld MC, Kokkotou EG and Pothos EN (2008). Evidence for defective mesolimbic dopamine exocytosis in obesity-prone rats. *The FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 22 (8): 2740-2746.
41. Geiger BM, Haburcak M, Avena NM, Moyer MC, Hoebel BG and Pothos EN (2009). Deficits of mesolimbic dopamine neurotransmission in rat dietary obesity. *Neuroscience*. 159 (4): 1193-1199.
42. Thanos PK, Michaelides M, Piyis YK, Wang GJ and Volkow ND (2008). Food restriction markedly increases dopamine D2 receptor (D2R) in a rat model of obesity as assessed with in-vivo muPET imaging ([11C] raclopride) and in-vitro ([3H] spiperone) autoradiography. *Synapse*. 62 (1): 50-61.
43. Anderzhanova E, Covasa M and Hajnal A (2007). Altered basal and stimulated accumbens dopamine release in obese OLETF rats as a function of age and diabetic status. *American journal of physiology Regulatory, integrative and comparative physiology*. 293 (2): R603-611.
44. Bello NT and Hajnal A (2006). Alterations in blood glucose levels under hyperinsulinemia affect accumbens dopamine. *Physiology & behavior*. 88 (1-2): 138-145.
45. Bello NT, Lucas LR and Hajnal A (2002). Repeated sucrose access influences dopamine D2 receptor density in the striatum. *Neuroreport*. 13 (12): 1575-1578.
46. Blum K, Braverman ER, Holder JM, Lubar JF, Monastra VJ, Miller D, Lubar JO, Chen TJ and Comings DE (2000). Reward deficiency syndrome: a biogenetic model for the diagnosis and treatment of impulsive, addictive, and compulsive behaviors. *Journal of psychoactive drugs*. 32 Suppl: i-iv, 1-112.
47. **Wang GJ, Volkow ND, Logan J, Pappas NR, Wong CT, Zhu W, Netusil N and Fowler JS (2001). Brain dopamine and obesity. *Lancet*. 357 (9253): 354-357. First human PET study of obesity, showing deficits in midbrain dopamine neurotransmission in obese individuals.**
48. Dawe S and Loxton NJ (2004). The role of impulsivity in the development of substance use and eating disorders. *Neuroscience and biobehavioral reviews*. 28 (3): 343-351.
49. Mathes WF, Nehrenberg DL, Gordon R, Hua K, Garland T, Jr. and Pomp D (2010). Dopaminergic dysregulation in mice selectively bred for excessive exercise or obesity. *Behavioural brain research*. 210 (2): 155-163.
50. Heusner CL, Hnasko TS, Szczypka MS, Liu Y, During MJ and Palmiter RD (2003). Viral restoration of dopamine to the nucleus accumbens is sufficient to induce a locomotor response to amphetamine. *Brain research*. 980 (2): 266-274.
51. Darvas M and Palmiter RD (2010). Restricting dopaminergic signaling to either dorsolateral or medial striatum facilitates cognition. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (3): 1158-1165.
52. Hnasko TS, Perez FA, Scouras AD, Stoll EA, Gale SD, Luquet S, Phillips PE, Kremer EJ and Palmiter RD (2006). Cre recombinase-mediated restoration of nigrostriatal dopamine in dopamine-deficient mice reverses hypophagia and bradykinesia. *Proceedings of the National Academy of Sciences of the United States of America*. 103 (23): 8858-8863.
53. Szczypka MS, Kwok K, Brot MD, Marck BT, Matsumoto AM, Donahue BA and Palmiter RD (2001). Dopamine production in the caudate putamen restores feeding in dopamine-deficient mice. *Neuron*. 30 (3): 819-828.
54. Faure A, Haberland U, Conde F and El Massioui N (2005). Lesion to the nigrostriatal dopamine system disrupts stimulus-response habit formation. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 25 (11): 2771-2780.
55. Graybiel AM (2008). Habits, rituals, and the evaluative brain. *Annual review of neuroscience*. 31: 359-387.
56. Yin HH, Knowlton BJ and Balleine BW (2004). Lesions of dorsolateral striatum preserve outcome expectancy but disrupt habit formation in instrumental learning. *The European journal of neuroscience*. 19 (1): 181-189.
57. Berridge KC, Robinson TE and Aldridge JW (2009). Dissecting components of reward: 'liking', 'wanting', and learning. *Current opinion in pharmacology*. 9 (1): 65-73.
58. Balleine BW and Dickinson A (1998). Goal-directed instrumental action: contingency and incentive learning and their cortical substrates. *Neuropharmacology*. 37 (4-5): 407-419.
59. Dickinson A, Nicholas DJ and Adams CD (1983). The Effect of the Instrumental Training Contingency on Susceptibility to Reinforcer Devaluation. *Quarterly Journal of Experimental Psychology Section B-Comparative and Physiological Psychology*. 35 (Feb): 35-51.
60. Hyman SE, Malenka RC and Nestler EJ (2006). Neural mechanisms of addiction: the role of reward-related learning and memory. *Annual review of neuroscience*. 29: 565-598.
61. Yin HH (2010). The sensorimotor striatum is necessary for serial order learning. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (44): 14719-14723.
62. Yin HH, Knowlton BJ and Balleine BW (2005). Blockade of NMDA receptors in the dorsomedial striatum prevents action-outcome learning in instrumental conditioning. *The European journal of neuroscience*. 22 (2): 505-512.
63. Kalivas PW (2009). The glutamate homeostasis hypothesis

of addiction. *Nature reviews Neuroscience*. 10 (8): 561-572.

64. Berke JD (2003). Learning and memory mechanisms involved in compulsive drug use and relapse. *Methods Mol Med*. 79: 75-101.

**65. Li CS, Huang C, Constable RT and Sinha R (2006). Imaging response inhibition in a stop-signal task: neural correlates independent of signal monitoring and post-response processing. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 26 (1): 186-192. First fMRI study using the stop signal paradigm to assess the neural correlates of response inhibition.**

66. Arnsten AF (2009). Stress signalling pathways that impair prefrontal cortex structure and function. *Nature reviews Neuroscience*. 10 (6): 410-422.

67. Miller EK and Cohen JD (2001). An integrative theory of prefrontal cortex function. *Annual review of neuroscience*. 24: 167-202.

68. Robbins TW and Arnsten AF (2009). The neuropsychopharmacology of fronto-executive function: monoaminergic modulation. *Annual review of neuroscience*. 32: 267-287.

69. Goldman-Rakic PS (1998). The cortical dopamine system: role in memory and cognition. *Advances in pharmacology*. 42: 707-711.

70. Seamans JK and Yang CR (2004). The principal features and mechanisms of dopamine modulation in the prefrontal cortex. *Prog Neurobiol*. 74 (1): 1-58.

71. Phillips AG, Ahn S and Floresco SB (2004). Magnitude of dopamine release in medial prefrontal cortex predicts accuracy of memory on a delayed response task. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 24 (2): 547-553.

72. Mehta MA and Riedel WJ (2006). Dopaminergic enhancement of cognitive function. *Curr Pharm Des*. 12 (20): 2487-2500.

73. Chudasama Y and Robbins TW (2004). Dopaminergic modulation of visual attention and working memory in the rodent prefrontal cortex. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 29 (9): 1628-1636.

74. Kruse MS, Premont J, Krebs MO and Jay TM (2009). Interaction of dopamine D1 with NMDA NR1 receptors in rat prefrontal cortex. *European neuropsychopharmacology : the journal of the European College of Neuropsychopharmacology*. 19 (4): 296-304.

75. Sarantis K, Matsokis N and Angelatou F (2009). Synergistic interactions of dopamine D1 and glutamate NMDA receptors in rat hippocampus and prefrontal cortex: involvement of ERK1/2 signaling. *Neuroscience*. 163 (4): 1135-1145.

76. Zahrt J, Taylor JR, Mathew RG and Arnsten AF (1997). Supranormal stimulation of D1 dopamine receptors in the rodent prefrontal cortex impairs spatial working memory performance. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 17 (21): 8528-8535.

77. Crofts HS, Dalley JW, Collins P, Van Denderen JC, Everitt BJ, Robbins TW and Roberts AC (2001). Differential effects of 6-OHDA lesions of the frontal cortex and caudate nucleus on the ability to acquire an attentional set. *Cereb Cortex*. 11 (11): 1015-1026.

78. Robbins TW and Roberts AC (2007). Differential regulation of fronto-executive function by the monoamines and acetylcholine. *Cereb Cortex*. 17 Suppl 1: i151-160.

79. Langley K, Marshall L, van den Bree M, Thomas H, Owen M, O'Donovan M and Thapar A (2004). Association of the dopamine D4 receptor gene 7-repeat allele with neuropsychological test perfor-

mance of children with ADHD. *Am J Psychiatry*. 161 (1): 133-138.

80. Eagle DM, Tufft MR, Goodchild HL and Robbins TW (2007). Differential effects of modafinil and methylphenidate on stop-signal reaction time task performance in the rat, and interactions with the dopamine receptor antagonist cis-flupenthixol. *Psychopharmacology*. 192 (2): 193-206.

81. Bari A, Mar AC, Theobald DE, Elands SA, Oganya KC, Eagle DM and Robbins TW (2011). Prefrontal and monoaminergic contributions to stop-signal task performance in rats. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 31 (25): 9254-9263.

82. Congdon E, Lesch KP and Canli T (2008). Analysis of DRD4 and DAT polymorphisms and behavioral inhibition in healthy adults: implications for impulsivity. *Am J Med Genet B Neuropsychiatr Genet*. 147B (1): 27-32.

83. Colzato LS, van den Wildenberg WP, van Wouwe NC, Pannebakker MM and Hommel B (2009). Dopamine and inhibitory action control: evidence from spontaneous eye blink rates. *Experimental brain research Experimentelle Hirnforschung Experimentation cerebrale*. 196 (3): 467-474.

84. Moron JA, Brockington A, Wise RA, Rocha BA and Hope BT (2002). Dopamine uptake through the norepinephrine transporter in brain regions with low levels of the dopamine transporter: evidence from knock-out mouse lines. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 22 (2): 389-395.

85. Waymunt HK, Schenk JO and Sorg BA (2001). Characterization of extracellular dopamine clearance in the medial prefrontal cortex: role of monoamine uptake and monoamine oxidase inhibition. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 21 (1): 35-44.

86. Robertson SD, Matthies HJ, Owens WA, Sathanathan V, Christianson NS, Kennedy JP, Lindsley CW, Daws LC and Galli A (2010). Insulin reveals Akt signaling as a novel regulator of norepinephrine transporter trafficking and norepinephrine homeostasis. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (34): 11305-11316.

87. Schoffeleers AN, Drukarch B, De Vries TJ, Hogenboom F, Schetters D and Pattij T (2011). Insulin modulates cocaine-sensitive monoamine transporter function and impulsive behavior. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 31 (4): 1284-1291.

88. Vijayraghavan S, Wang M, Birnbaum SG, Williams GV and Arnsten AF (2007). Inverted-U dopamine D1 receptor actions on prefrontal neurons engaged in working memory. *Nature neuroscience*. 10 (3): 376-384.

89. George O and Koob GF (2010). Individual differences in prefrontal cortex function and the transition from drug use to drug dependence. *Neuroscience and biobehavioral reviews*. 35 (2): 232-247.

90. Small DM (2008). Flavor and the formation of category-specific processing in olfaction. *Chemosensory Perception*. 1 (2): 136-146.

91. Small DM (2009). Individual differences in the neurophysiology of reward and the obesity epidemic. *International journal of obesity*. 33 Suppl 2: S44-48.

92. Bender G, Veldhuizen MG, Meltzer JA, Gitelman DR and Small DM (2009). Neural correlates of evaluative compared with passive tasting. *The European journal of neuroscience*. 30 (2): 327-338.

93. Felsted JA, Ren X, Chouinard-Decorte F and Small DM (2010). Genetically determined differences in brain response to a pri-

## CANDIDATE REVIEWS

mary food reward. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (7): 2428-2432.

94. O'Doherty JP, Deichmann R, Critchley HD and Dolan RJ (2002). Neural responses during anticipation of a primary taste reward. *Neuron*. 33 (5): 815-826.

95. Small DM, Zatorre RJ, Dagher A, Evans AC and Jones-Gotman M (2001). Changes in brain activity related to eating chocolate: from pleasure to aversion. *Brain : a journal of neurology*. 124 (Pt 9): 1720-1733.

96. Uher R, Treasure J, Heining M, Brammer MJ and Campbell IC (2006). Cerebral processing of food-related stimuli: effects of fasting and gender. *Behavioural brain research*. 169 (1): 111-119.

97. Stice E, Yokum S, Blum K and Bohon C (2010). Weight gain is associated with reduced striatal response to palatable food. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (39): 13105-13109.

98. Pelchat ML, Johnson A, Chan R, Valdez J and Ragland JD (2004). Images of desire: food-craving activation during fMRI. *NeuroImage*. 23 (4): 1486-1493.

99. Roefs A, Herman CP, Macleod CM, Smulders FT and Jansen A (2005). At first sight: how do restrained eaters evaluate high-fat palatable foods? *Appetite*. 44 (1): 103-114.

100. LaBar KS, Gitelman DR, Parrish TB, Kim Y-H, Nobre AC and Mesulam MM (2001). Hunger selectively modulates corticolimbic activation to food stimuli in humans. *Behavioral Neuroscience*. 115 (2): 493-500.

101. Porubska K, Veit R, Preissl H, Fritsche A and Birbaumer N (2006). Subjective feeling of appetite modulates brain activity: an fMRI study. *NeuroImage*. 32 (3): 1273-1280.

102. Cornier MA, Salzberg AK, Endly DC, Bessesen DH, Rojas DC and Tregellas JR (2009). The effects of overfeeding on the neuronal response to visual food cues in thin and reduced-obese individuals. *PLoS One*. 4 (7): e6310.

103. Del Parigi A, Gautier JF, Chen K, Salbe AD, Ravussin E, Reiman E and Tataranni PA (2002). Neuroimaging and obesity: mapping the brain responses to hunger and satiation in humans using positron emission tomography. *Annals of the New York Academy of Sciences*. 967: 389-397.

104. Beaver JD, Lawrence AD, van Ditzhuijzen J, Davis MH, Woods A and Calder AJ (2006). Individual differences in reward drive predict neural responses to images of food. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 26 (19): 5160-5166.

105. Goldstone AP, Prechtl de Hernandez CG, Beaver JD, Muhammed K, Croese C, Bell G, Durighel G, Hughes E, Waldman AD, Frost G and Bell JD (2009). Fasting biases brain reward systems towards high-calorie foods. *The European journal of neuroscience*. 30 (8): 1625-1635.

106. Stoeckel LE, Cox JE, Cook EW, 3rd and Weller RE (2007). Motivational state modulates the hedonic value of food images differently in men and women. *Appetite*. 48 (2): 139-144.

107. Fletcher PC, Napolitano A, Skeggs A, Miller SR, Delafont B, Cambridge VC, de Wit S, Nathan PJ, Brooke A, O'Rahilly S, Farooqi IS and Bullmore ET (2010). Distinct modulatory effects of satiety and sibutramine on brain responses to food images in humans: a double dissociation across hypothalamus, amygdala, and ventral striatum. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 30 (43): 14346-14355.

108. Rothmund Y, Preuschhof C, Bohner G, Bauknecht HC,

Klingebiel R, Flor H and Klapp BF (2007). Differential activation of the dorsal striatum by high-calorie visual food stimuli in obese individuals. *NeuroImage*. 37 (2): 410-421.

109. Stice E, Spoor S, Bohon C and Small DM (2008). Relation between obesity and blunted striatal response to food is moderated by TaqIA A1 allele. *Science*. 322 (5900): 449-452.

110. Stoeckel LE, Weller RE, Cook EW, 3rd, Twieg DB, Knowlton RC and Cox JE (2008). Widespread reward-system activation in obese women in response to pictures of high-calorie foods. *NeuroImage*. 41 (2): 636-647. fMRI study examining the neural correlates of viewing food cues, stratified by nutritional content, in obese and healthy-weight individuals.

111. Yokum S, Ng J and Stice E (2011). Attentional Bias to Food Images Associated With Elevated Weight and Future Weight Gain: An fMRI Study. *Obesity*.

112. Wang GJ, Volkow ND, Thanos PK and Fowler JS (2009). Imaging of brain dopamine pathways: implications for understanding obesity. *J Addict Med*. 3 (1): 8-18.

113. Davis C, Levitan RD, Kaplan AS, Carter J, Reid C, Curtis C, Patte K, Hwang R and Kennedy JL (2008). Reward sensitivity and the D2 dopamine receptor gene: A case-control study of binge eating disorder. *Progress in neuro-psychopharmacology & biological psychiatry*. 32 (3): 620-628.

114. Batterink L, Yokum S and Stice E (2010). Body mass correlates inversely with inhibitory control in response to food among adolescent girls: an fMRI study. *NeuroImage*. 52 (4): 1696-1703.

115. Volkow ND, Wang GJ, Telang F, Fowler JS, Thanos PK, Logan J, Alexoff D, Ding YS, Wong C, Ma Y and Pradhan K (2008). Low dopamine striatal D2 receptors are associated with prefrontal metabolism in obese subjects: possible contributing factors. *NeuroImage*. 42 (4): 1537-1543.

116. Volkow ND, Wang GJ, Fowler JS, Logan J, Jayne M, Franceschi D, Wong C, Gatley SJ, Gifford AN, Ding YS and Pappas N (2002). "Nonhedonic" food motivation in humans involves dopamine in the dorsal striatum and methylphenidate amplifies this effect. *Synapse*. 44 (3): 175-180.

117. Wang GJ, Geliebter A, Volkow ND, Telang FW, Logan J, Jayne MC, Galanti K, Selig PA, Han H, Zhu W, Wong CT and Fowler JS (2011). Enhanced Striatal Dopamine Release During Food Stimulation in Binge Eating Disorder. *Obesity*.

118. Bina KG and Cincotta AH (2000). Dopaminergic agonists normalize elevated hypothalamic neuro peptide Y and corticotropin-releasing hormone, body weight gain, and hyperglycemia in ob/ob mice. *Neuroendocrinology*. 71 (1): 68-78.

119. Davis LM, Michaelides M, Cheskin LJ, Moran TH, Aja S, Watkins PA, Pei Z, Contoreggi C, McCullough K, Hope B, Wang GJ, Volkow ND and Thanos PK (2009). Bromocriptine administration reduces hyperphagia and adiposity and differentially affects dopamine D2 receptor and transporter binding in leptin-receptor-deficient Zucker rats and rats with diet-induced obesity. *Neuroendocrinology*. 89 (2): 152-162.

120. Steele KE, Prokopowicz GP, Schweitzer MA, Magunsoo TH, Lidor AO, Kuwabawa H, Kumar A, Brasic J and Wong DF (2010). Alterations of central dopamine receptors before and after gastric bypass surgery. *Obes Surg*. 20 (3): 369-374.

121. Ochner CN, Kwok Y, Conceicao E, Pantazatos SP, Puma LM, Carnell S, Teixeira J, Hirsch J and Geliebter A (2011). Selective reduction in neural responses to high calorie foods following gastric bypass surgery. *Ann Surg*. 253 (3): 502-507.



122. McCaffery JM, Haley AP, Sweet LH, Phelan S, Raynor HA, Del Parigi A, Cohen R and Wing RR (2009). Differential functional magnetic resonance imaging response to food pictures in successful weight-loss maintainers relative to normal-weight and obese controls. *The American journal of clinical nutrition*. 90 (4): 928-934.