The Role of the Principal Sensory Nucleus in Discriminative Touch Eva Sawyer

Specialized facial somatosensory organs have evolved in diverse groups of animals, and the sense of touch that these organs transduce is important for normal behavior. The principal sensory nucleus of the spinal trigeminal complex is the first relay for facial discriminative touch in the central nervous system. Much of the work done on this nucleus is done in rodents, where the ability to trace the central representations of whiskers follicle innervation has been a useful tool for experimenters. Questions remain about the role of the nucleus, from uncertainties about the basic anatomy to its role in forming the disproportionate representation of the body seen in the cortical somatosensory maps. Comparative neurobiology points out that some non-rodent animals with specialized trigeminal somatosensory organs, such as the star-nosed mole, have a much larger principal sensory nucleus than one would expect for a mammal of their size. Complementing rodent work with studies on these species has the potential to help resolve puzzles about the entire spinal trigeminal complex, and the principal sensory nucleus in particular.

Keywords: Somatosensory, trigeminal, segmentation, principal sensory nucleus, Barrels, star-nosed mole

Introduction

Facial somatosensory specializations help animals navigate their world. Examples include whiskers on the face rodents and seals^{1, 2}, corpuscles of Herdst and Grandry on the beak of ground-probing birds^{3, 4}, push-rod receptors on the bills of montremes⁵, integumentary sense organs on the jaws of crocodilians⁶, and Eimer's organs on the noses of talpid moles⁷. These adaptations are associated with exploration, foraging and feeding^{8, 9}. The co-evolution of sense organs and central processing centers is a theme in neurobiology^{10, ¹¹. Accordingly, when researchers have looked at the first relay of the trigeminal somatosensory stream, these specialized trigeminal touch organs tend to be paired with central specializations¹²⁻¹⁵.}

The spinal trigeminal complex (STC) is the main target for the primary somatosensory receptors innervating the scalp, face and oral structures. The complex consists of the principal sensory nucleus (PrV) at the most rostral position and the spinal trigeminal nucleus (STN) more caudally. The latter consists of three subnuclei with subnucleus pars oralis (SpVo), subnucleus pars interpolaris (SpVi), and subnucleus pars caudalis (SpVc) found at progressively more caudal positions, respectively^{16, 17}. SpVc merges with the dorsal horn of the spinal cord at its most caudal extent. The complex receives most of its sensory input from the somatosensory components of the trigeminal nerve (but also from the somatosensory components of the facial, glossopharyngeal and vagus nerves)¹⁶. Upon entering the brainstem, the trigeminal branch splits into an ascending branch that projects to the PrV and a descending branch that projects to the subdivisions of the STN.

Traditionally, there is a perception that in the somatosensory brainstem there is a division of labor so that the PrV mediates light touch sensation and the STN mediates pain¹⁸. In this view, the PrV is analogous to the dorsal column nuclei and the STN to the dorsal horn^{19, 20}. Broadly, there is truth in the vital importance of the PrV to discriminative touch and the STN, especially the more caudal regions, to pain, but a strict view of non-converging labeled lines has weak support. This article will focus on the PrV, but it would be misleading to present the nucleus as if it were completely independent from the STN. Therefore, the STN will be mentioned where appropriate.

Form and Function

Ramón y Cajal illustrated trigeminal afferent axons branching to form an ascending path to the PrV and a descending path to the STN²¹. He writes that he cannot be sure that all of these axons bifurcate because he cannot rely on the



silver stain as an unbiased technique, but many, if not all, of the fibers he saw bifurcated. In a reinvestigation with silver stains Anstrom states he found the bifurcating fibers Ramón y Cajal reported, as well as descending non-bifurcating fibers projecting to only the STN²². He wrote that he did not observe, but could not rule out the possibility of, ascending non-bifurcating fibers projecting only to the PrV. Presumably, if these fibers exist, they would be mechanosensory axons that project exclusively to the PrV.

Another anatomist found such fibers. Windle, like Ramón y Cajal and Anstrom, used silver stains, but his studies were on fetal pigs instead of fetal and young mice²³. He found three sub-populations of trigeminal afferents: 52% of axons bifurcated, 42% (mostly thin fibers) descended without bifurcating, and 6% (mostly thick fibers) ascended without bifurcating. He struggled to explain why other investigators did not observe large diameter non-bifurcating ascending fibers. One possibility he does not mention is that mice, with vibrissa used for whisking, and pigs, with a large glabrous nose used for rooting in soil, differ in their facial somatosensory specializations. Since the mechanoreceptors in these specializations differ, the animals may have different proportions of bifurcating and non-bifurcating trigeminal afferents.

Subsequent literature on ascending non-bifurcating axons is sparse, though the perception that this pathway exists is maintained in modern reviews and texts²⁴⁻²⁷. One method for studying projection patterns of sensory neurons in the brainstem is interaxonal injections of tracer combined with reconstruction of the labeled axons. This has been used in the STC with interaxonal injections of the neuronal tracers horseradish peroxidase or neurobiotin into the spinal trigeminal tract. Unfortunately, the injections are almost always at the level of SpVo or SpVi²⁸⁻³³, a method which could not isolate an ascending non-bifurcating population. A less biased technique would use interaxonal injections Figure 1: Schematic of main connections of the principal sensory nucleus (PrV) and the regulatory input directed through spinal trigeminal nucleus pars interpolaris (SpVi). The dotted line represents an unconfirmed class of sensory neurons projecting solely to the PrV. Thick lines represent the main pathway for low-threshold mechanoreception to the cortex. DR, dorsal raphe nucleus; LC, locus coeruleus; PN, Pontine nucleus; Pom, medial posterior nucleus; SC, superior colliculus; SpVc, spinal trigeminal nucleus pars caudalis; SpVo, spinal trigeminal nucleus pars oralis; ZI, zona incerta.

upstream of the bifurcation, as done by Shigenaga et al.³⁴, or to inject a far-reaching tracer into single ganglion cells, as Jaquin et al.³⁵ piloted in a methods paper. Neither study found ascending non-bifurcating axons. The absence of this class could be because such neurons are not present in rodents. However, given that neurons with the ascending non-bifurcating branching pattern made up only 6% of trigeminal afferents in pigs, a combined sample size of 12 axons in rodents is unlikely to represent sufficient sampling to warrant a strong conclusion that these cells are absent in rodents.

Within the PrV, SpVo and SpVi the somatotopic map of the face is inverted so that afferents from the mandibular branch project dorsally, the maxillary branch intermediately and the ophthalmic branch ventrally^{28, 31}. The anterior receptors are represented medially and the more posterior receptors, laterally. The arrangement in the SpVc is less well understood. Some studies find that the dorsal-ventral representation is unchanged but the medial-lateral representation is flipped, so that the anterior receptors are represented laterally and the more posterior receptors, medially^{17,} ³⁶⁻³⁸. They also find that in SpVc there is rostral-caudal skew that results in the more rostral afferents being represented more rostrally in the nucleus. This arrangement is reminiscent of the rostral-caudal mapping of dermatomes found in the dorsal horn for the rest of the body. Despite this work, reviews sometimes depict the SpVc as organized like the other subdivisions²⁶, and recently the somatotopy of SpVc has reemerged as an issue³⁷. It is noteworthy that there is still confusion about basic anatomy of the STC.

In addition to the main sensory input from primary sensory neurons, the PrV receives modulatory input from the STN and the cortex. Inhibitory GABAergic interneurons from the SpVi and excitatory glutamatergic interneurons from the SpVc project to the PrV⁴⁰. These connections let the STN influence the sensitivity of the PrV⁴¹. Projections

from the primary and secondary somatosensory areas (S1 and S2) to the STC could also facilitate top-down reduction of PrV sensitivity⁴²⁻⁴³. In rodents, the cortex-STN-PrV pathway is thought to be particularly important during active whisking, when the somatosensory signals induced by body movement, and not by the characteristics of a substrate, are irrelevant. Such a circuit could explain how the sensitivity of the PrV is reduced during active whisking⁴⁴. Other inputs are from the pontine tegmental nucleus⁴⁵⁻⁴⁶, the raphe nucleus⁴⁷ and the locus coeruleus⁴⁸. These likely reflect modulation of sensitivity based on the animal's level of alertness.

From the PrV, neurons project mainly to the contralateral ventral posterior medial nucleus (VPM) of the thalamus, which sends strong projections to S1^{12,49-51}. This trigeminal lemniscal pathway is particularly notable in rodents because every station on this pathway (the PrV, the VPM and S1) has a pattern of cell-dense patches that correspond in a one-to-one manner with the whiskers on the animal's snout, termed barrelettes¹⁷, barreloids⁵¹, and barrels¹² in each location, respectively. We will return to barrels later in the essay. Other important direct projections are to zona incerta⁵², the posterior medial nucleus of the thalamus⁴⁹ and the superior colliculus⁵³. These projections likely contribute to the regulation of movement. The main connections of the PrV are summarized in **Figure 1**.

Electrophysiological work in a variety of species complements the above anatomical findings. As would be expected from the termination patterns of large diameter bifurcating axons branching to every STC subdivision, the PrV and the STN contain mechanosensory neurons^{14,54-58}. Likewise, the anatomy shows that many small diameter fibers are non-bifurcating descending axons. If these are nociceptive c-fibers, then electrophysiological studies should find the STN enriched with nociceptive neurons. Indeed, electrophysiological studies that test for it fail to find nociceptive neurons in the PRV, but isolate them in STN^{54,56,59-61}.

Another promising area for animal studies is to use the power of genetic manipulations in model species to dissect the pathways of mechanoreceptors. For example, Li et al.⁶² drove expression of reporter proteins in different classes of low-threshold mechanoreceptors in order to follow sensory neurons from the receptors in the skin to their projections in the dorsal horn. The same techniques are yet to be applied to neurons projecting to the dorsal column nuclei or to the STC. Both studies would be valuable. In the whisker pathway, it would be interesting to see if the different classes of low-threshold mechanoreceptors have unique projection patterns to the subdivisions of the STC, which has so far been undetected with electrophysiology and tract tracing.

Human case studies provide strong results that support the view of parallel pain and touch pathways. Lesions in the PrV cause deficits in touch sensation with sparing of temperature sense and nociception in the face63, while lesions in the SpVc spare touch sensation but usually lead to the loss of nociception and temperature perception¹⁸. Thus, surgical damage to this area is a treatment for intractable orofacial pathogenic pain. With case studies such as these providing most of the background for the understanding of the human trigeminal system, a recent fMRI study was surprising. The study looked for changes in the blood oxygen level in humans who were experiencing noxious and nonnoxious cutaneous and muscle stimulation to their face. As expected, noxious cutaneous and noxious muscle stimulation elicited changes in regions of the STN, but unexpectedly, the noxious muscle stimulation also elicited activation in PrV⁶⁴.

These unexpected results showing integration of pain and sensory information in the brainstem fit with a history of confusion about trying to connect the anatomical data, the electrophysiological data and now the fMRI data — which all show some integration of low-threshold mechanoreception and nociception — with human case studies, which show a strong division of touch and pain sensation between the nuclei^{18,22,64}. One problem is that lesion studies rely on eliminating an entire node of a network, which provides strong but crude results. Another problem is that there is a push to make labeled lines for pain and touch explain the anatomical divisions of the STC, despite evidence that the divisions will not fit well into those categories. That debate, however, is beyond the scope of this article (see ⁶⁵ for review).

Comparative neurobiology

Rodents have been seen as well suited for studies of the trigeminal touch pathway because their barrel system is more amenable to experimentation than non-patterned areas. The cortical barrels are impressive. For example, in rats, barrels cover 20% of S1, a total area of 9 mm² ⁶⁶. The development of this pattern is dependent on an intact PrV^{67} . As othes have pointed out, it is odd that all the input for such a large cortical representation is funneled through a small PrV^{54} , 0.56mm³ in the case of a rat¹³. Part of the resolution is that the cortex also receives connections from the



Figure 2: Cytochrome oxidase stained sections of a monse, rat and star-nosed mole brainstem cut in the horizontal plane. Chosen sections maximized the volume of the principal sensory nucleus (PrV). Compared to the mouse and the rat, the star-nosed mole has a large PrV. SpVc, spinal trigeminal pars caudalis; SpVi, spinal trigeminal pars interpolaris; SpVo, spinal trigeminal pars oralis.

STN. Specifically, the caudal portions of SpVi project to the ventrolateral portion of the VPM in the thalamus. The VPM projects to the inter-barrel space, termed septa, in the cortex⁶⁸.

In rats, the volume of the SpVi is 1.66 mm³, almost three times larger than the PrV, and has more distinct barrelettes than the PrV^{17,69}. Earlier we saw that the SpVi has a role in modulating sensory sensitivity based on directed movements of the whiskers. A large part of rodent exploratory behavior is active whisking — coordinated movements of the six muscles innervating each whisker pad to move the whiskers against the surface being examined^{2,70-72}. All this suggests that rats and other rodents are good models for studying the SpVi, particularly because the subnucleus relates to active sensory behavior. But other animals may be better suited for investigating the PrV.

Comparative studies point out that in some species with elaborate somatosensory trigeminal sensory organs, the PrV is hypertrophied^{9,14}. The case of the star-nosed mole is particularly informative because of the amount already known about its nervous system. The star consists of 22 fleshy appendages covered with Eimer's organs. Eimer's organs are composed of regular geometric arrangements of Merkel cell-neurite complexes, laminated corpuscles and free nerve-endings⁷. The star can be moved forward as a whole, and groups of appendages can be extended to bring the organ surface into contact with a substrate⁷³. When moles forage, they rapidly move the star, touching it to the surfaces of the damp soil of their habitat to locate small food items in the mud⁷⁴. There is a behavioral preference to use the two medial ventral rays when inspecting potential food items75. The neuroanatomical correlate of that preference is a larger representation of that ray in S1, and smaller receptive field size in the representation of the medial ventral rays than in the other rays^{73,76}. The behavioral preference and the increased resolution suggest those rays as a somatosensory analog of the retina's fovea⁷⁵.

To put the size of the star-nosed mole PrV into perspective, the absolute volume of the PrV of a 55g star-nosed mole is larger than the PrV of a 274g rat — it is about 630% the size expected based on the proportions of a rat¹³⁻¹⁴. In comparison, the SpVi subdivision of the star-nosed mole is only about 50% larger than expected⁹ (unpublished result). Acknowledging that the comparisons are crude and cover a wide taxonomic range, the results still show that the starnosed mole has a large PrV even when compared to other somatosensory specialists (**Figure 2**).

The size of the PrV in the star-nosed mole is likely related to the exceptional spatial resolution of the star. Multi-unit receptive fields in the cortex average 0.82 mm² in the nonfoveal part of the star and 0.52 mm² in the foveal regions, which are both smaller than receptive fields reported for primate fingertips⁷⁶⁻⁷⁸. If the PrV is the nucleus for fine touch, the extraordinary resolution of the star would be expected to distinguish this nucleus. There is already evidence that this is the case: within the PrV, as in the cortex, the medial ventral rays have a larger representation than the other rays. Interestingly, the greater size of the representation of the foveal rays in the cortex and PrV is not explained by greater innervation of these rays¹⁴. Combined with the smaller receptive fields in foveal than in nonfoveal areas in S1, this suggests that within the lemniscal pathway foveal afferents converge less than the afferents for other rays.



Finding that a behaviorally important area of skin is overrepresented in the central nervous system is not new⁷⁹. But finding that the size of the somatosensory representation cannot be predicted by counting the number of fibers innervating that structure and multiplying by a constant "afferent scaling factor" is special⁸⁰. This result is important because it suggests the mole PrV, and perhaps the rest of the lemniscal pathway, could be used to address questions about how the central over-representation of a foveal area of a sensory epithelium comes about.

Conclusion

The role of the PrV, compared to other regions of the STC is relatively understudied. The oversight is surprising given its vital role in organizing the somatosensory cortex. The lack of focus on the PrV might be due to the relatively unimpressive PrV in rodents compared to other trigeminal somatosensory specialists. There are many unresolved questions that could be addressed with comparative work. Just some include: Anatomically, what contributes to a hypertrophied PrV? Are there unique afferents? Is there less convergence? Within a nucleus, what contributes to the "foveal" area of higher resolution? Understanding these points will inform us on the forces that link the evolution of sensory surfaces and their central representations.

References

- 1. Dehnhardt G, Mauck B, Hanke W and Bleckmann H (2001). Hydrodynamic trail-following in harbor seals (Phoca vitulina). Science. 293 (5527): 102-104.
- 2. Ahl AS (1986). The role of vibrissae in behavior: a status review. Vet Res Commun. 10 (4): 245-268.
- 3. Dubbeldam JL (1980). Studies on the somatotopy of the trigeminal system in the mallard, Anas platyrhynchos L. II. Morphology of the principal sensory nucleus. J Comp Neurol. 191 (4): 557-571.
- 4. Pettigrew JD and Frost BJ (1985). A tactile fovea in the Scolopacidae? Brain Behav Evol. 26 (3-4): 105-195.
- Proske U, Gregory JE and Iggo A (1998). Sensory receptors in monotremes. Philos Trans R Soc Lond B Biol Sci. 353 (1372): 1187-1198.
- 6. Leitch DB and Catania KC (2012). Structure, innervation and response properties of integumentary sensory organs in crocodilians. J Exp Biol. 215 (Pt 23): 4217-4230.
- 7. Catania KC (1996). Ultrastructure of the Eimer's organ of the star-nosed mole. J Comp Neurol. 365 (3): 343-354.
- 8. Catania KC (2005). Evolution of sensory specializations in insectivores. Anat Rec A Discov Mol Cell Evol Biol. 287 (1):

1038-1050.

- 9. Gutierrez-Ibanez C, Iwaniuk AN and Wylie DR (2009). The independent evolution of the enlargement of the principal sensory nucleus of the trigeminal nerve in three different groups of birds. Brain Behav Evol. 74 (4): 280-294.
- 10. Van der Loos H and Dorfl J (1978). Does the skin tell the somatosensory cortex how to construct a map of the periphery? Neurosci Lett. 7 (1): 23-30.
- 11. Krubitzer LA and Seelke AMH (2012). Cortical evolution in mammals: The bane and beauty of phenotypic variability. Proc Natl Acad Sci U S A. 109: 10647-10654.
- 12. Woolsey TA and Van der Loos H (1970). The structural organization of layer IV in the somatosensory region (SI) of mouse cerebral cortex. The description of a cortical field composed of discrete cytoarchitectonic units. Brain Res. 17 (2): 205-242.
- 13. Ashwell KW, Hardman CD and Paxinos G (2006). Cyto- and chemoarchitecture of the sensory trigeminal nuclei of the echidna, platypus and rat. J Chem Neuroanat. 31 (2): 81-107.
- 14. Catania KC, Leitch DB and Gauthier D (2011). A Star in the Brainstem Reveals the First Step of Cortical Magnification. Plos One. 6 (7).

This article reports on a star representation in the hypertrophied principal sensory nucleus of the star-nosed mole. Importantly, the authors find that the size of the behaviorally important "fovea" rays is magnified at the level of the brainstem.

- 15. Sarko DK, Johnson JI, Switzer RC, 3rd, Welker WI and Reep RL (2007). Somatosensory nuclei of the manatee brainstem and thalamus. Anat Rec (Hoboken). 290 (9): 1138-1165.
- Olszewski J (1950). On the anatomical and functional organization of the spinal trigeminal nucleus. J Comp Neurol. 92 (3): 401-413.
- 17. Ma PM (1991). The barrelettes--architectonic vibrissal representations in the brainstem trigeminal complex of the mouse. I. Normal structural organization. J Comp Neurol. 309 (2): 161-199.
- 18. Gerard MW (1923). Afferent impulses of the trigeminal nerve. AMA Arch Neurol Psychiat. 9: 306–338.
- 19. Darian-Smith I, Phillips G and Ryan RD (1963). Functional Organization in the Trigeminal Main Sensory and Rostral Spinal Nuclei of the Cat. J Physiol. 168: 129-146.
- Dubner R and Bennett GJ (1983). Spinal and Trigeminal Mechanisms of Nociception. Annual Review of Neuroscience. 6: 381-418.
- 21. Ramon y Cajal S (1896). Beitrag zum Studium der Medulla oblongata, des Kleinhirns uud des Ursprungs der Gehirnnerven. Leipzig: J. A. Barth.
- 22. Anstrom KE (1953). On the central course of afferent fibres in the trigeminal, facial, glossopharyngeal, and vagal nerves and their nuclei in the mouse. Acta physiol scand Suppl. 106: 209-320.
- 23. Windle WF (1926). Non-bifurcating nerve fibers of the trigeminal nerve. The Journal of Comparative Neurology 40 (1): 229-240.

This article reports on the projection patterns of trigeminal sensory afferents. Windle finds non-bifurcating assending axons, an observation which is the basis for the modern perception that such axons exsist. The finding has not been replicated, though it is not clear that the hypothesis that these axons exist has been well tested.

- 24. Butler AB and Hodos W (2005). Comparative vertebrate neuroanatomy : evolution and adaptation, 2nd Edition. Hoboken, N.J.: Wiley-Interscience.
- 25. Martin JH (2003). Neuroanatomy : text and atlas, 3rd Edition. New York, N.Y.: McGraw-Hill.
- 26. Bosman LW, Houweling AR, Owens CB, Tanke N, Shevchouk OT, Rahmati N, Teunissen WH, Ju C, Gong W, Koekkoek SK and De Zeeuw CI (2011). Anatomical pathways involved in generating and sensing rhythmic whisker movements. Front Integr Neurosci. 5: 53.
- 27. Usunoff KG, Marani E and Schoen JH (1997). The trigeminal system in man. Adv Anat Embryol Cell Biol. 136: I-X, 1-126.
- Hayashi H (1980). Distributions of vibrissae afferent fiber collaterals in the trigeminal nuclei as revealed by intra-axonal injection of horseradish peroxidase. Brain Res. 183 (2): 442-446.
- 29. Hayashi H (1985). Morphology of terminations of small and large myelinated trigeminal primary afferent fibers in the cat. J Comp Neurol. 240 (1): 71-89.
- Jacquin MF, Renehan WE, Rhoades RW and Panneton WM (1993). Morphology and topography of identified primary afferents in trigeminal subnuclei principalis and oralis. J Neurophysiol. 70 (5): 1911-1936.
- Shortland PJ, Demaro JA, Shang F, Waite PM and Jacquin MF (1996). Peripheral and central predictors of whisker afferent morphology in the rat brainstem. J Comp Neurol. 375 (3): 481-501.
- 32. Chiaia NL, Hess PR, Hosoi M and Rhoades RW (1987). Morphological characteristics of low-threshold primary afferents in the trigeminal subnuclei interpolaris and caudalis (the medullary dorsal horn) of the golden hamster. J Comp Neurol. 264 (4): 527-546.
- 33. Tsuru K, Otani K, Kajiyama K, Suemune S and Shigenaga Y (1989). Central terminations of periodontal mechanoreceptive and tooth pulp afferents in the trigeminal principal and oral nuclei of the cat. Brain Res. 485 (1): 29-61.
- 34. Shigenaga Y, Otani K and Suemune S (1990). Morphology of central terminations of low-threshold trigeminal primary afferents from facial skin in the cat--intra-axonal staining with HRP. Brain Res. 523 (1): 23-50.
- 35. Jacquin MF, Hu JW, Sessle BJ, Renehan WE and Waite PM (1992). Intra-axonal Neurobiotin injection rapidly stains the long-range projections of identified trigeminal primary afferents in vivo: comparisons with HRP and PHA-L. J Neurosci Methods. 45 (1-2): 71-86.
- 36. Arvidsson J (1982). Somatotopic organization of vibrissae afferents in the trigeminal sensory nuclei of the rat studied by transganglionic transport of HRP. J Comp Neurol. 211 (1): 84-92.
- 37. da Silva S, Hasegawa H, Scott A, Zhou X, Wagner AK, Han BX and Wang F (2011). Proper formation of whisker barrelettes requires periphery-derived Smad4-dependent TGF-beta signaling. Proc Natl Acad Sci U S A. 108 (8): 3395-3400.

- Jacquin MF, Renehan WE, Mooney RD and Rhoades RW (1986). Structure-function relationships in rat medullary and cervical dorsal horns. I. Trigeminal primary afferents. J Neurophysiol. 55 (6): 1153-1186.
- Sjöqvist O (1938). Studies on pain conduction in the trigeminal nerve; a contribution to the surgical treatment of facial pain. Helsingfors: Mercators tryckeri.
- 40. Furuta T, Timofeeva E, Nakamura K, Okamoto-Furuta K, Togo M, Kaneko T and Deschenes M (2008). Inhibitory gating of vibrissal inputs in the brainstem. J Neurosci. 28 (8): 1789-1797.
- 41. Timofeeva E, Lavallee P, Arsenault D and Deschenes M (2004). Synthesis of multiwhisker-receptive fields in subcortical stations of the vibrissa system. J Neurophysiol. 91 (4): 1510-1515.
- 42. Aronoff R, Matyas F, Mateo C, Ciron C, Schneider B and Petersen CC (2010). Long-range connectivity of mouse primary somatosensory barrel cortex. Eur J Neurosci. 31 (12): 2221-2233.
- 43. Haque T, Akhter F, Kato T, Sato F, Takeda R, Higashiyama K, Moritani M, Bae YC, Sessle BJ and Yoshida A (2012). Somatotopic direct projections from orofacial areas of secondary somatosensory cortex to trigeminal sensory nuclear complex in rats. Neuroscience. 219: 214-233.
- 44. Lee S, Carvell GE and Simons DJ (2008). Motor modulation of afferent somatosensory circuits. Nat Neurosci. 11 (12): 1430-1438.
- 45. Timofeeva E, Dufresne C, Sik A, Zhang ZW and Deschenes M (2005). Cholinergic modulation of vibrissal receptive fields in trigeminal nuclei. J Neurosci. 25 (40): 9135-9143.
- 46. Beak SK, Hong EY and Lee HS (2010). Collateral projection from the forebrain and mesopontine cholinergic neurons to whisker-related, sensory and motor regions of the rat. Brain Res. 1336: 30-45.
- 47. Lee SB, Lee HS and Waterhouse BD (2008). The collateral projection from the dorsal raphe nucleus to whisker-related, trigeminal sensory and facial motor systems in the rat. Brain Res. 1214: 11-22.
- 48. Moore RY and Bloom FE (1979). Central catecholamine neuron systems: anatomy and physiology of the norepinephrine and epinephrine systems. Annual Review of Neuroscience. 2: 113-168.
- 49. Veinante P and Deschenes M (1999). Single- and multiwhisker channels in the ascending projections from the principal trigeminal nucleus in the rat. J Neurosci. 19 (12): 5085-5095.
- 50. Furuta T, Kaneko T and Deschenes M (2009). Septal neurons in barrel cortex derive their receptive field input from the lemniscal pathway. J Neurosci. 29 (13): 4089-4095.
- 51. Van der Loos H (1976). Barreloids in mouse somatosensory thalamus. Neurosci Lett. 2 (1): 1-6.
- 52. Kolmac CI, Power BD and Mitrofanis J (1998). Patterns of connections between zona incerta and brainstem in rats. J Comp Neurol. 396 (4): 544-555.
- 53. Steindler DA (1985). Trigeminocerebellar, trigeminotectal, and trigeminothalamic projections: a double retrograde

axonal tracing study in the mouse. J Comp Neurol. 237 (2): 155-68. 175.

- 54. Kirkpatrick DB and Kruger L (1975). Physiological properties of neurons in the principal sensory trigeminal nucleus of the cat. 69. Exp Neurol. 48 (3 Pt 1): 664-690.
- 55. Zeigler HP and Witkovsky P (1968). The main sensory trigeminal nucleus in the pigeon: a single-unit analysis. J Comp Neurol. 134 (3): 255-264.
- 56. Gordon G, Landgren S and Seed WA (1961). The functional characteristics of single cells in the caudal part of the spinal nucleus of the trigeminal nerve of the cat. J Physiol. 158: 544-559.
- 57. Silver R and Witkovsky P (1973). Functional characteristics of single units in the spinal trigeminal nucleus of the pigeon. Brain Behav Evol. 8 (4): 287-303.
- Molenaar GJ, Fizaan-Oostveen JL and van der Zalm JM (1979). Infrared and tactile units in the sensory trigeminal system of python reticulatus. Brain Res. 170 (2): 372-376.
- 59. Azerad J, Woda A and Albe-Fessard D (1982). Physiological Properties of neurons in different parts of the cat trigeminal sensory complex. Brain Res. 246 (1): 7-21.
- 60. Mosso JA and Kruger L (1972). Spinal trigeminal neurons excited by noxious and thermal stimuli. Brain Res. 38 (1): 206-210.
- Dallel R, Clavelou P and Woda A (1989). Effects of tractotomy 76. on nociceptive reactions induced by tooth pulp stimulation in the rat. Exp Neurol. 106 (1): 78-84.
- 62. Li LS, Rutlin M, Abraira VE, Cassidy C, Kus L, Gong SC, Jankowski MP, Luo WQ, Heintz N, Koerber HR, Woodbury CJ and Ginty DD (2011). The Functional Organization of Cutaneous Low-Threshold Mechanosensory Neurons. Cell. 147 (7): 1615-1627.

The authors use transgenic mice and immunohistochemistry to selectively label 4 classes of low-threshold mechanoreceptors. This allows them to trace the peripheral and central projections of somatosensory neurons, a goal that has been largely elusive in somatosensory research.

- 63. Kamitani T, Kuroiwa Y and Hidaka M (2004). Isolated hypesthesia in the right V2 and V3 dermatomes after a midpontine infarction localised at an ipsilateral principal sensory trigeminal nucleus. J Neurol Neurosurg Psychiatry. 75 (10): 1508-1509.
- 64. Nash PG, Macefield VG, Klineberg IJ, Murray GM and Henderson LA (2009). Differential activation of the human trigeminal nuclear complex by noxious and non-noxious orofacial stimulation. Hum Brain Mapp. 30 (11): 3772-3782.
- 65. Craig AD (2003). Pain mechanisms: labeled lines versus convergence in central processing. Annual Review of Neuroscience. 26: 1-30.
- 66. Welker C (1971). Microelectrode delineation of fine grain somatotopic organization of (SmI) cerebral neocortex in albino rat. Brain Res. 26 (2): 259-275.
- 67. Killackey HP and Fleming K (1985). The role of the principal sensory nucleus in central trigeminal pattern formation. Brain Res. 354 (1): 141-145.

- Pierret T, Lavallee P and Deschenes M (2000). Parallel streams for the relay of vibrissal information through thalamic barreloids. J Neurosci. 20 (19): 7455-7462.
- Lo FS, Guido W and Erzurumlu RS (1999). Electrophysiological properties and synaptic responses of cells in the trigeminal principal sensory nucleus of postnatal rats. J Neurophysiol. 82 (5): 2765-2775.
- Vincent SB (1912). The function of vibrissae in the behavior of the white rat. Behav Monogr. 1: 1–82.

70.

71.

73.

74.

80.

- Dorfl J (1982). The musculature of the mystacial vibrissae of the white mouse. J Anat. 135 (Pt 1): 147-154.
- 72. Hartmann MJ (2011). A night in the life of a rat: vibrissal mechanics and tactile exploration. Ann N Y Acad Sci. 1225: 110-118.
 - Catania KC and Kaas JH (1995). Organization of the somatosensory cortex of the star-nosed mole. J Comp Neurol. 351 (4): 549-567.
 - Catania KC and Remple FE (2005). Asymptotic prey profitability drives star-nosed moles to the foraging speed limit. Nature. 433 (7025): 519-522.
- 75. Catania KC and Kaas JH (1997). Somatosensory fovea in the starnosed mole: behavioral use of the star in relation to innervation patterns and cortical representation. J Comp Neurol. 387 (2): 215-233.
 - Sachdev RN and Catania KC (2002). Receptive fields and response properties of neurons in the star-nosed mole's somatosensory fovea. J Neurophysiol. 87 (5): 2602-2611.
- 77. Pons TP, Wall JT, Garraghty PE, Cusick CG and Kaas JH (1987). Consistent features of the representation of the hand in area 3b of macaque monkeys. Somatosens Res. 4 (4): 309-331.
- 78. Xerri C, Merzenich MM, Peterson BE and Jenkins W (1998). Plasticity of primary somatosensory cortex paralleling sensorimotor skill recovery from stroke in adult monkeys. J Neurophysiol. 79 (4): 2119-2148.
- 79. Penfield W and Rasmussen T (1950). The cerebral cortex of man; a clinical study of localization of function. New York,: Macmillan.
 - Welker E and Van der Loos H (1986). Quantitative correlation between barrel-field size and the sensory innervation of the whiskerpad: a comparative study in six strains of mice bred for different patterns of mystacial vibrissae. J Neurosci. 6 (11): 3355-3373.

This article finds tests the relationship between whisker innervation and cortical barrel area in six strains of mice breed for their different whisker patterns. The authors find that within a strain there is a linear relationship between the variables, but the slope of the relationship differs between strains. The existence of a linear relationship can be contrasted with case in the star-nosed mole.

Correspondence: eva.k.sawyer@vanderbilt.edu

Further information: http://as.vanderbilt.edu/catanialab