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LETTER FROM THE EDITORS

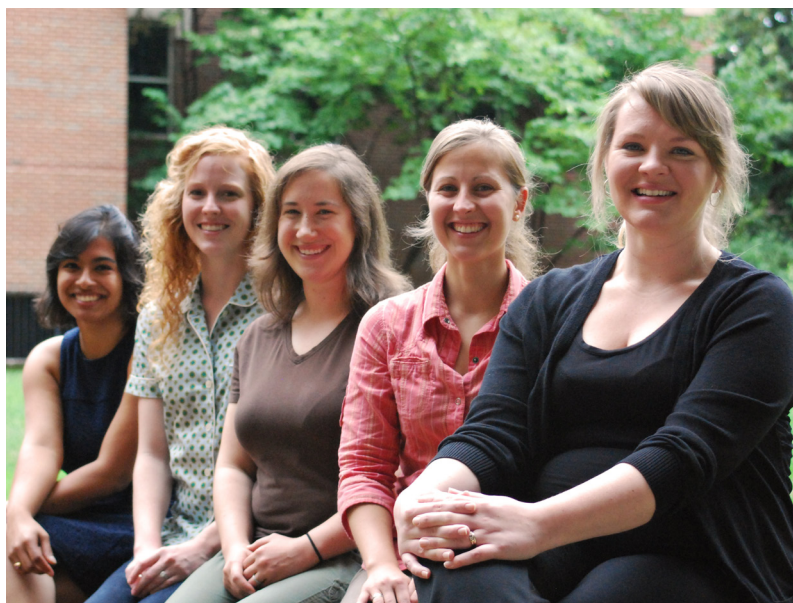
Dear Friends and Colleagues of the Vanderbilt Neuroscience Community,

The *Vanderbilt Reviews Neuroscience* is still alive and kicking, and we're excited to bring you another exciting issue this year! With preserving our recent style and formatting makeover, we hope to promote this layout as the permanent template for all future issues. A new feature of this year's issue is our wonderful cover art, a scientific homage to the cartoonist style of one of our favorite publications, *The New Yorker*. Special thanks to Eva Sawyer for her artistic (and scientific) talents! We also included a new section in this issue called "Grad Student Corner" that is dedicated to current student issues and potential resources that may be valuable to our students. Regardless, our goal remains the same in continuing to highlight the talented writing and research interests of the newest Ph.D. candidates in the Vanderbilt Neuroscience Program. This issue features twelve candidate reviews covering topics from both systems and molecular neuroscience, including: the role of neurotransmitter transporter regulation in diseases; circuit processing in anxiety and autism, and the role of the developing amygdala in childhood maltreatment; circuit refinement and neurotransmitter signaling during development; touch mechanisms; and new potential CNS drug targets, as well as a patient-derived stem cell model for Parkinson's disease. We also highlight a collection of first-author papers published by Vanderbilt Neuroscience students over the past calendar year. Our students excel in publishing in high impact journals, making insightful contributions that are highly regarded by external peer review. Keep up the good work, and thanks for making our jobs easier by providing such great content for the *VRN*!

This *VRN* issue would not have been possible without the painstaking contributions from our tenacious editing board. Our Associate Editors, Suzanne Avery, Hayley Clay and Barbara O'Brien (all pictured below), are incredibly talented and hardworking. For the first time, our associate editing board consisted of all females (girl power!), and the future of *VRN* remains bright under their leadership for next year. We had a blast with you guys! We would also like to recognize the outgoing Co-Editors-in-Chief, Drs. Andrew Hardaway and Maureen McHugo, as well as Founder/Editor-in-Chief, Dr. Chris Ciarleglio, for helping establish the *VRN* and equipping us with a valuable network of resources to successfully produce this year's issue. Our sincerest thanks also go to both Drs. Mark Wallace and Doug McMahon, who have helped to guide the role of the *VRN* in sculpting each qualifying student's graduate experience. Their support and enthusiasm for a student-run endeavor like the *VRN* is only one example of the Vanderbilt Neuroscience Program's ideals in promoting the development of successful, independent thinkers. Finally, we would like to thank the 2012 qualifying class for their hard work in generating the true foundation of this issue and tolerating our many edits and emails. This would not have been possible without you!

Your Co-Editors-in-Chief,

Sudipta Chakraborty and Juliane Krueger Fister



From left: Sudipta Chakraborty, Hayley Clay, Barbara O'Brien, Juliane Krueger Fister, and Suzanne Avery



Juliane Krueger Fister and Sudipta Chakraborty

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Review Process

All reviews submitted for doctoral qualification must be approved by a committee of at least four tenured or tenure-track faculty members (Phase I). All approved reviews are accepted by *VRN*.

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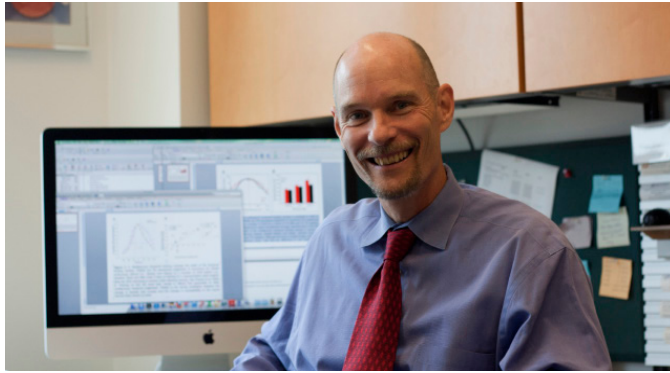
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A Message from the Director of the Vanderbilt Brain Institute

It is with a sense of deep pride that I review this fifth issue of *VRN*. Once again I am very impressed with the quality and diversity of the reviews that make up this volume, as well as with the synopses that illustrate the exceptional year our trainees have had in the realm of scholarship.

This past month has seen the VBI staff assemble the proposal for renewal of our training grant, and in gathering the materials necessary to make the strongest possible arguments for continuing (and increased) support, I was pleased to be able to point to the numerous successes that we have had over the past several years. Along with continuing our exceptional record of scholarship, with numerous papers in high profile journals authored by our trainees, we have had a remarkable record of success in garnering extramural support in the form of individual fellowship grants and foundation awards. Past students of our program are now transitioning to faculty positions at prestigious institutions such as Harvard, Yale, and Duke (and Vanderbilt of course!), and our newly minted PhDs continue to do postdoctoral fellowships in some of the world's leading neuroscience laboratories. With these accomplishments to reflect on, I feel confident that our mission to train the future leaders in the neurosciences is well on track!

Yours in science,

Mark T. Wallace, PhD

A Message from the Neuroscience Program Director of Graduate Studies

A neuroscience journal run, edited and contributed entirely by graduate students - how cool is that? Plenty, when the quality of the work published and the care with which it is assembled is as special as *VRN*. *VRN* showcases the scholarly work of our outstanding students as they assemble and synthesize their knowledge base and move toward fully implementing their dissertation research. The original articles highlight the insight and intellectual curiosity of Neuroscience graduate students at Vanderbilt and foreshadow their novel contributions to solving the mysteries of the brain. Each issue is a special pleasure to behold.

As ever,

Douglas McMahon, PhD



An Update from the Neuroscience Student Organization President

First, I would like to welcome all incoming students to our wonderful Neuroscience community. Be sure to get to know your new fellow graduate students as we will be each other's future peers and colleagues. Second, I want to congratulate our twelve new PhD candidates, whose qualifying reviews are featured in this issue. This group of talented young scientists is now moving on to phase II of the qualifying process, which includes a NRSA-style research thesis proposal.

As the Neuroscience Student Organization (NSO) president it is my distinct honor to highlight here some of the achievements of our student body. We currently have 14 active Ruth L. Kirchstein National Research Awardees, as well as one award each from Autism Speaks, the National Science Foundation and the American Heart Association. Furthermore, since the last VRN issue, 10 students have successfully defended their PhD and moved on to bigger and better things. This year, we have a total of seven Clinical Neuroscience Scholars (CNS) Program recipients. The CNS program is now in its second year and has been incredibly successful in connecting scientist and medical doctors to bridge the gap between the basic and clinical research sciences. Last year, we all had the pleasure to listen to the CNS scholars' experiences during a series of CNS research forums.

A big thanks goes out to the organizers of our Neuroscience Boot Camp. This boot camp has now taken place for two years in a row and has been a tremendous success in connecting the incoming students, as well as providing senior graduate students with teaching experiences and creating an informal atmosphere to discuss fundamental neuroscience topics. Jeff Jones, Terry Jo Bichell, and Ben Dean were instrumental in developing and executing a curriculum for this camp, which was held one to two weeks prior to the first day of classes. Our student body comes from diverse educational backgrounds, and the boot camp is designed to bring the students together on the same level as they enter the program. Lectures and paper discussions are prepared by upper level students, and incoming students are encouraged to participate by drawing on a whiteboard as they would during the qualifying exam process. Overall, I believe the camp has been a great success and I truly hope it will be continued in the

upcoming years.

Next, I would like to point out a few changes to the Neuroscience curriculum. In order to better streamline classes, our fundamental neuroscience courses, Systems Neuroscience and Cell and Molecular Neuroscience, are now called Fundamentals I and II and are building upon each other and further integrate the dependency of molecular and systems neurosciences on each other. A host of elective courses have also been added to meet the demands of the student body as well as the ever-growing field of neuroscience. This year, we also have our inaugural class of graduate students through the Educational Neuroscience track. I am looking forward to their progress and contributions to science and educational research.

Another yearly staple is the student run Neuroscience Retreat. The 16th annual retreat will be held again at the Joe C. Davis YMCA Outdoor Center. This year our goal is to promote interactions between new and current students as well as the overall neuroscience community. The retreat will include short talks from Dr. Kim Petrie and Dr. Ruth Schemmer regarding important concepts for early, mid and later career development, including topics ranging from effective communication, resume writing do's and don'ts, and opportunities for intern and externships here at Vandderbilt. We will also get a first-hand account of what it is like to start a lab from a Vanderbilt graduate, Dr. Brad Greuter. A poster session open to students, staff and post-docs will be held to allow the Neuroscience community to get a taste of the exciting science occurring on campus. We are also happy to host NIH Distinguished Investigator Dr. Kenneth Fischbeck, who will give the Keynote lecture.

Lastly, I will echo my predecessors, and encourage everyone to get involved in any facet of our student organization and neuroscience community. We are an organization of students for students and can only grow with your support and commitment.

Yours truly,

Juliane Krueger Fister



OUTREACH+ EDUCATION

This year's **Brain Blast** was held Saturday, March 2, and children from the community were introduced to hands-on neuroscience research with graduate students, post-doctoral fellow, and principal investigators.

During Brain Blast, children explored brain research by experiencing optical, somatotopic, and sensorimotor illusions with the Tong and Wallace labs. They used their own brain waves to control objects with the Roe Lab's MindFlex demonstration, extracted DNA from strawberries with the Emeson Lab, and touched a real human brain at the Tick-Tock station! Other labs brought animals so that the community could learn about how we use animal models to investigate different aspects of brain function. The circadian group brought their Madagascar hissing cockroaches, and many children (including our very own Jeff Jones) loved handling the creepy crawly critters. Less hands-on, Eva Sawyer and Duncan Leitch from the Catania Lab brought an American alligator to show the highly sensitive receptors lining the jaw of this fantastic reptile. Children even had some microscope time with Ben Dean from the Gamse Lab looking at fluorescent neurons in transparent fish. Other hands-on activities also encouraged interest in brain research. Student-run booths included putting together 3-D brain puzzles, building neurons, making brain hats, or guessing objects via smell, sound, or touch. Be sure to keep a look out for updates on next year's Brain Blast.



Top right: Brain Blast booths at One Hundred Oaks Mall.

Middle right: Jeff Jones showing off a Madagaskar hissing coackroach.

Bottom right: A very interative touch screen was a big hit with the audience, demonstrated here by Dan Bermingham.

Top left: A little girl very obviously enthralled with the Catania lab alligator.

Bottom left: Loui, Terry Jo Bichell's son checking out the zebrafish through the microscope with help from Ben Dean.



** photos provided by Barbara O'Brien and Eva Sawyer*

A Message From Your Middle Tennessee SfN Chapter

Dear Members of the Vanderbilt Neuroscience Community,

Thanks to you, it has been another exciting and successful year for the Middle Tennessee Chapter of the Society for Neuroscience. Our chapter is one of well over 100 spanning the globe and is gaining increasing attention from the national organization for its dynamic character and exciting initiatives. Last year we were invited to give a presentation on some of our innovative activities (such as our pumpkin decorating social) at the Chapters Workshop at the Annual Society for Neuroscience meeting.

Since its inception more than 10 years ago, our chapter has continued to grow and expand its membership beyond Vanderbilt to include a number of local institutions such as Meharry, Austin Peay, Fisk and TSU. At this year's Annual Meeting, faculty from several of these schools treated us to short talks about their research, followed by our Data Blitz presentations by local trainees and a social jointly sponsored by the Neuroscience Student Organization. We just completed our 3rd Summer Enrichment Research Program in Education and Neuroscience Training, or SERPENT, with a student from TN Technical University, Matthew Defenderfer. This program allows an undergraduate from a local institution to come to Vanderbilt and do research over the summer. It also includes an outreach component, since the summer student is tasked with developing a brief educational lesson/experience for the public or local K-12 schools. Matt spent his summer working in Mark Wallace's lab and for his outreach project he developed an interactive optical illusion game that we will highlight at next year's Brain Blast. Matt will be applying to Vanderbilt's PhD program in neuroscience after graduating next year. This would be our 3rd student to matriculate into a graduate program, making our rate of SERPENT students who continue on to advanced research 100%!

If you are not a member of the chapter already, I would strongly encourage you to join us and become an active part of the local neuroscience community. Being part of the chapter increases your networking opportunities, provides a forum for sharing scientific ideas, allows you to engage in educating the public about neuroscience research, and there are several travel and lecture awards available. Of course, there are fun activities as well (recall the bal-

loon "flocking" of Mark's office). What the chapter is and what it does is determined by its membership. It is the goal of the current leadership team to make the Middle TN Chapter a showcase example of what can be done through the local chapters. At present, we continue to focus on expanding our membership through increasing the involvement of other local institutions and developing new funding mechanisms so that we can create innovative and novel programming initiatives.

Do not miss the opportunity to be a part of a vibrant and dynamic community of neuroscientists right here in your neighborhood! We can't realize the full potential of our chapter and make it into the national treasure that we aspire to without your involvement and support. You are just one click away from joining and/or making a donation! Don't wait, do it now! (<http://www.mtncsf.org/>).



Bruce D. Carter, PhD

President of the Middle Tennessee Chapter of the Society for Neuroscience



Modern Science Chaos: Taking New Initiatives

The changing climate of today's modern science world has resulted in a unique shift in career paths and job outcomes around the country. According to the NIH Biomedical Workforce Working Group Report, the number of Ph.D. trainees has significantly increased since 2004, in parallel with the doubling of the NIH budget. This allowed for an increase in the number of predoctoral research grants, whereas placement on training grants were previously a major source of Ph.D. recruitment. Unlike training grants, individual research grants do not necessarily require the same level of training that is specific for professional development. Concurrently, universities began to put a focus on increasing the number of non-tenure-track positions that are dependent on outside funding and would not require the university to cover their salaries. Together, this brews the perfect storm of chaos: the increased production of Ph.D.'s only trained to excel in the academic sector, in an environment where there are not nearly enough positions available for the majority of them.

Therefore, a strong demand exists for better training PhD candidates for "non-traditional" career routes outside of academia. Clearly, the focus of Ph.D. trainees remains to successfully complete thesis projects and learn to become successful, independent scientists. However, while some programs may require teaching experiences for graduation, a strong absence of exposure to any other career paths is highly evident - students are only trained to conduct and communicate research, apply for funding, direct and maintain projects, and perhaps teach to a minor degree. With the current state of science, this proves to be disadvantageous to the biomedical field, as candidates enter alternative career paths without any previous training or necessary knowledge to become successful in these sectors. Yet, Ph.D. trainees are able to critically question, evaluate and solve problems, making them highly marketable across many fields.

This rising concern is widespread, as even the National Science Foundation (NSF), a prominent scientific funding and research agency, put out a call this year for innovative ideas that would enhance STEM (science, technology, engineering, and mathematics) graduate education. Our own Neuroscience Student Organization (NSO) even put in an

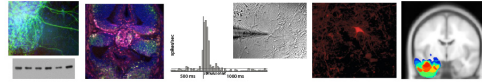
application of ideas (geared towards women and minorities in the STEM fields), in addition to other PhD students around campus. Vanderbilt's BRET (Biomedical Research, Education and Training) office is currently looking into revolutionary methods that may integrate training experiences into the graduate education system. We optimistically hope for a new system for future graduate students, where official Ph.D. candidates are able to participate in short-term, unpaid intern/externships that promote immersion into various sectors outside of academia that are actively part of modern science. In the meantime, I encourage you to reach out on your own towards sectors that may be of interest to you as a career path, be it consulting, scientific writing, regulatory affairs, biotechnology project management, clinical trials, etc. Speaking from personal experience, even sending a simple email to someone in your area of interest about their careers and their training experiences along the way can prove to be fruitful in both gaining new information and forming new professional contacts. In fact, Vanderbilt has several clinical research labs that can provide a wealth of information in terms of how this sector operates, in addition to the presence of other biotech, regulatory and writing agencies around Nashville. I also urge you to get involved on committees and programs around campus that pertain to your interests. There are so many to offer, including the Saturday Academy at Vanderbilt for the Young (SAVY) and Weekend Academy at Vanderbilt (WAVU) programs for promoting education for children in grades K-10; the Scientist Educator Program; the Editors' Club; the HHMI/ VUMC Certificate Program in Molecular Medicine (CPMM); and even our own VRN editing board. Until agreements develop between graduate programs and funding agencies concerning the implementation of new training experiences, we all need to "take the bull by its horns" and prepare ourselves the best we can for an exciting new wave of science careers that will impact the world in ways previous generations have never seen!

Sudipta Chakraborty

Resources for Neuroscience Students Compiled by Neuroscience Students:

Campus Resources

- Vanderbilt Research News: <http://news.vanderbilt.edu/research>
- Neuroscience Student Organization: <http://www.mc.vanderbilt.edu/root/vumc.php?site=NSO>
- Vanderbilt Depolarized (Student Blog): <https://my.vanderbilt.edu/vanderbiltdepolarized>
- BRET Office: <https://medschool.vanderbilt.edu/bret>
- Center for Teaching: <http://cft.vanderbilt.edu>
- Graduate School: <http://www.vanderbilt.edu/gradschool>
- Eskind Library: <http://www.mc.vanderbilt.edu/diglib>
- Vanderbilt CORES: <http://www.mc.vanderbilt.edu/root/vumc.php?site=CFUIS>
- SAVY/WAVU Teaching Programs: <http://pty.vanderbilt.edu/contact/employment/for-vu-faculty-staff-and-students>
- Vanderbilt Brain Institute: <http://braininstitute.vanderbilt.edu/index.php>
- Middle TN Chapter for the Society of Neuroscience (MTNCSfN): <http://www.mtncsfn.org>



MTNCSfN

A Society for Neuroscience Chapter

Serving the Middle Tennessee Area

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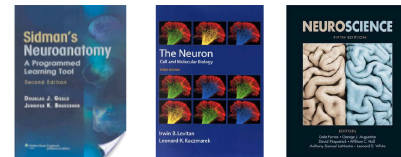
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Professional networks: www.linkedin.com and <http://www.researchgate.net>



Teach yourself molecular biology concepts through youtube:

- Central dogma: <http://www.youtube.com/watch?v=J3HVV2k2No>
- Post-translational modifications: http://www.neb.com/nebecomm/tech_reference/epigenetics/epigenetics.asp#UCZCAI7lpqg
- Vesicle dance: <http://www.youtube.com/watch?v=BdKbRgT4hn8>

Seriously teach yourself molecular biology concepts:

- NCBI main page: www.ncbi.nlm.nih.gov/About/index.html
- Basic Cell Biology: www.ncbi.nlm.nih.gov/About/primer/genetics_cell.html
- Great review lectures by noted scientists: www.ibioseminars.org



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- Protocol database: www.sciclips.com/sciclips/bio-protocols.do
- Current Protocols in Electrophysiology: <http://www.sciclips.com/sciclips/bio-protocols.do?catName=Neuroscience&l2CatName=Electrophysiology>
- See videos of techniques through The Journal of Visual Experiments: www.jove.com



Get help with calculations:

- Dilution calculator: www.tocris.com/dilutionCalculator
- Molarity calculator: www.graphpad.com/quickcalcs/Molarityform.cfm
- App calculator: <http://www.sigmaaldrich.com>



Neuroanatomy:

- www.brain-map.org
- <http://mouse.brainarchitecture.org>
- www.brainmuseum.org



RESEARCH HIGHLIGHTS

Overcoming Dravet syndrome by overcoming the GABRG2(Q40X) mutation

Xuan Huang, Tian, M, Hernandez, CC, Hu, N, & MacDonald, RL (2012). The GABRG2 nonsense mutation, Q40X, associated with Dravet syndrome activated NMD and generated a truncated subunit that was partially rescued by aminoglycoside induced stop codon read through, *Neurobiology of Disease* 48(1):115-123.

Epilepsy is one of the most common neurological disorders, and can arise from brain injury or genetic mutation. Commonly, mutations causing epilepsy occur in genes key to maintaining the electrical balance of neurons, including voltage-gated ion channels and GABA_A receptors. In Huang et al. (2012), the Macdonald lab investigates a mutation seen in one of the more insidious types of idiopathic genetic epilepsy, Dravet syndrome. This form of epilepsy involves frequent seizures and progressive intellectual decline, and many cases can be traced back to nonsense mutations resulting in premature stop codons.

The mutation of interest in this study, GABRG2(Q40X), causes a premature stop codon the GABA_A γ 2S subunit. Human embryonic kidney (HEK 293T) cells were transfected with wild-type γ 2S or γ 2S(Q40X) and probed for changes in γ 2S mRNA expression after knockdown of a component of the nonsense-mediated decay machinery. Many mRNAs containing the Q40X mutation were indeed degraded via nonsense-mediated decay, and those mRNAs that did make it to protein translation produced only a truncated peptide. This peptide did not mature, as evidenced by a lack of glycosylation, and was not expressed at the cell surface, as a wild-type γ 2S subunit normally would be.

In addition to its problems with mRNA stability, protein expression, and protein maturation, γ 2S subunits containing the Q40X mutation were not incorporated into GABA_A receptors. HEK 293T cells transfected with the α 1, β 2, and γ 2S(Q40X) subunits exhibited no γ 2S(Q40X) surface expression, and the α 1 and β 2 subunits displayed similar surface protein expression patterns as cells that were transfected with α 1 and β 2 subunits but not γ 2S. Assays for whole-cell receptor levels indicated that the lack of γ 2S(Q40X) surface expression was due to an overall lack

of γ 2S(Q40X) expression, rather than a defect in its secretion to the cell surface.

Cells transfected with the γ 2S(Q40X) subunit also displayed functional abnormalities consistent with their exclusion from GABA_A receptors. Compared to wild-type, receptors on γ 2S(Q40X)-transfected cells displayed reduced peak amplitudes in GABA-evoked currents and enhanced sensitivity to zinc-mediated inhibition. These phenotypes, however, matched those of cells expressing only the α 1 and β 2 subunits, indicating that GABA_A receptors in cells transfected with γ 2S(Q40X) were actually α 1 β 2 receptors.

The antibiotic gentamicin can be used to promote read-through of premature stop codons in mRNA. Gentamicin treatment of cells transfected with γ 2S(Q40X) had increased overall expression of the mutant γ 2S protein. This phenotype was enhanced when the premature stop codon was changed to a TGA stop codon, which increased the cell's gentamicin-mediated read-through abilities. In addition to increasing overall protein expression of γ 2S(Q40X), gentamicin treatment enhanced surface expression of γ 2S(Q40X). These γ 2S(Q40X) subunits at the cell surface were incorporated into functional GABA_A receptors, as electrophysiological recordings subsequently showed two hallmarks of γ 2S-containing GABA_A receptors: zinc insensitivity and diazepam-mediated current amplitude potentiation.

These data not only provide insight into the mechanism of Dravet syndrome cases stemming from the GABRG2(Q40X) mutation, but also a mechanism by which the mutation's effects may be overcome. With the ability to partially rescue the mutant phenotype with premature stop codon read-through, this work provides a promising trajectory for future investigations into how to treat genetic forms of epilepsy.

He may be rough around the edges, but that reptile is highly sensitive

Duncan B Leitch, & Catania, KC (2012). Structure, innervation and response properties of integumentary sensory organs in crocodilians, *Journal of Experimental Biology* 215: 4217-4230.

Crocodilians may have thick skin, but that skin is more sensitive than your fingertips. The cover of the December 2012 issue of the *Journal of Experimental Biology* promi-

nently featured Duncan Leitch's article on somatosensory receptors of crocodilians, experts at detecting and catching prey. What gives them this incredible ability to so swiftly and precisely detect the exact location of their target? Leitch argues that it begins with the fine mechanoreceptors in the skin that are able to detect the slightest vibration at the water's surface.

The integumentary sensory organs (ISOs) visibly located on the skin of alligators and crocodiles have been speculated to serve many functions from sensing electric or magnetic properties of water to secreting protective oils. Leitch and Catania (2012) designed a study that would examine the distribution, structure, and physiological responses of ISOs for alligators and crocodiles to provide more insight into the function of these unique receptors. Using a behavioral paradigm, they ruled out the hypotheses of these being electro- or magneto- receptors and showed that somatosensory input alone is sufficient for the animal to locate its target.

While crocodiles have ISOs distributed on their entire bodies, ISOs on alligators are restricted to the head, more specifically in and around the mouth and jaw. Each ISO is composed of free nerve endings, discoid receptors, and terminals that appeared to be similar to the Paciniform corpuscles, all of which contribute to the ISO's extreme sensitivity. At the center of each ISO is a dermal Merkel cell column. To examine the sensitivity of this receptor, the authors recorded in vivo extracellular single- and multi-unit activity from the trigeminal ganglion while stimulating the skin with fine wooden probes or calibrated von Frey hairs. The smallest and most numerous receptive fields were located at the rostral portion of the face. Thresholds of ISOs varied, but in some cases, the threshold could not be recorded because it was smaller than the smallest calibrated force that could be applied. The most sensitive ISOs were found near the teeth and mandible. When a sea salt solution was applied to the receptors, no activity was detected. Similarly, activity was not detected in response to a 9V battery placed in the water. These results indicate that the receptors do not detect ionic changes in the water. Similar results were shown for recordings from the spinal nerves that innervated ISOs on the limbs and digits.

In examining the structure and function of ISOs, this article supports the hypothesis that ISOs are mechanoreceptors that specify the location of a disturbance in the water, based on the strength of the vibration signal. To correlate

these findings with behavior, the authors recorded orientation to a food pellet dropped in water or to freely swimming fish. In complete darkness and with a white noise masker, crocodilians were able to orient to and efficiently capture prey, suggesting the sensitivity of ISOs plays a large role in identifying the location of a target, even when auditory and visual cues are absent.

The semantics of ecstasy use

Tristan J Watkins, Raj, V, Lee J, Dietrich, MS, Cao, A, Blackford, JU, Salmono, RM, Park, S, Benningfield, MM, Di Iorio, CR, & Cowan, RL (2013). Ecstasy (MDMA) polydrug users have altered brain activation during semantic processing, *Psychopharmacology* 227(1): 41-54.

Recreational experimentation with ecstasy (3,4-methylenedioxyamphetamine or MDMA) has risen dramatically in the United States in recent years, with first-time users increasing 23% between 2008 and 2009. This trend is especially concerning due to the potential for long-term negative impact on the central nervous system—ecstasy use has been associated with both neuroanatomical and neurophysiological effects, such as degeneration of presynaptic axon terminals and dysregulation of serotonergic (5-HT) function, as well as long-term neurocognitive deficits, particularly within the realm of memory ability. Although long-term abstinence may be associated with partial improvement, several studies suggest sustained effects despite temperance. Given rising use and the potential for long-term negative impacts, it is critical that we develop a better understanding of the effects of ecstasy on the human brain.

The most consistent neurocognitive deficits found in ecstasy users have been in verbal memory, suggesting that verbal memory ability may be a sensitive marker of ecstasy-related impairments. As such, probing brain regions associated with verbal memory impairments in ecstasy users may be a useful approach for understanding the neural basis of ecstasy toxicity. Therefore, in a recent Psychopharmacology report, Tristan Watkins and colleagues examined the association of lifetime ecstasy use with semantic memory performance and brain activation. In this study, abstinent ecstasy users (n=23) and controls (n=11) performed a two-part semantic encoding and recognition task while acquiring functional magnetic resonance imaging (fMRI) data. During the encoding task, subjects mem-

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orized both words and non-words (pronounceable Dutch words that would have no meaning for non-Dutch speaking subjects). During the recognition task, subjects were randomly presented with previously-shown words and non-words paired with phonological homophones that had not been previously shown, and asked to choose the prior studied word. Functional MRI data were collected during both parts of the semantic encoding and recognition task, and semantically-relevant brain activations were investigated using the contrast of word > non-word. During the encoding phase, ecstasy users had greater fMRI activity than controls in bilateral language processing areas, including Brodmann areas 7, 39, and 40. However, there were no between-group differences in semantic memory ability during the recognition task, nor were there differences in fMRI activity during recognition. These findings suggest that in order to achieve normal memory ability, ecstasy users' brains may use compensatory mechanisms, whereby greater neuronal activity during encoding is necessary to preserve verbal memory performance.

These conclusions, although preliminary, are in line with known effects of ecstasy-induced 5-HT toxicity in the brain. Ecstasy use is associated with degeneration of presynaptic axon terminals, 5-HT depletion, reduced binding to 5-HT reuptake transporters (5-HTT), reduced levels of 5-HT breakdown product 5-hydroxyindoleacetic acid (5-HIAA), and chronic upregulation of 5-HT_{2A} receptors. Since the net effect of 5-HT in the cortex is inhibitory, reduced 5-HT cortical signaling in ecstasy users—whether from axon loss or due to chronic reductions in 5-HT signaling—would be consistent with increased cortical activation relative to control subjects. This study highlights potential consequences of ecstasy-induced 5-HT neurotoxic effects and will guide future studies in human ecstasy users.

RESEARCH BRIEFS

Behavioral inhibition is a strong risk factor for social anxiety disorder

Jacqueline Clauss, & Blackford, JU (2012). Behavioral Inhibition and Risk for Developing Social Anxiety Disorder: A Meta-Analytic Study, *Journal of the American Academy of Child & Adolescent Psychiatry* 51(10): 1066-1075.

lescent Psychiatry 51(10): 1066-1075.

Social anxiety disorder (SAD)—the persistent and intense fear of being scrutinized by others—is a highly prevalent anxiety disorder that affects approximately 1 in 10 people in their lifetime. SAD is chronic, typically develops in adolescence, and can cause profound functional impairment at school and with peers, eventually resulting in lower educational attainment and wages, and fewer quality relationships during adulthood. SAD also confers risk for development of comorbid psychiatric disorders; 70% of adults with SAD will eventually develop a comorbidity such as major depression or substance abuse. Given the significant disability associated with SAD, early identification and intervention of at-risk adolescents would be ideal. One of the strongest known risk factors is behavioral inhibition (BI)—the chronic tendency to respond to novel people, places, and objects with wariness and avoidance behaviors. BI is heritable, emerges in childhood, and is present in 15-20% of individuals. However, the degree of risk associated with BI varies from study to study, making the size and potential clinical significance of the association unclear. Therefore, Clauss and colleagues conducted a meta-analysis to quantify the association between childhood BI and risk for SAD. The results of this analysis show that BI confers a greater than sevenfold increase in risk of developing SAD (odds ratio = 7.59), meaning that over 40% of children with BI (as opposed to 10% of the general population) will go on to eventually develop SAD. This indicates that BI is a principal and clinically significant predictor of risk for SAD, and an important indicator for early intervention and prevention.

Swimming worms provide insight into new modifiers of dopamine signaling

J Andrew Hardaway, & Hardie, SL, Whitaker, SM, Baas, SR, Zhang, B, Birmingham, DO, Lichtenstein, AJ, & Blakely, RD (2012). Forward Genetic Analysis to Identify Determinants of Dopamine Signaling in *Caenorhabditis elegans* Using Swimming-Induced Paralysis, *G3 (Bethesda)* 2(8): 961-975.

The dopamine (DA) pathway is complex in its control of both cognitive and motor behavior, with altered signaling resulting in several different brain diseases. Despite the current knowledge on key components of the pathway, investigation into novel modifiers of the pathway may illuminate further mechanisms behind DA-associated brain

diseases. Hardaway and colleagues took advantage of an unbiased, forward genetics approach using the genetically amenable, invertebrate model system, *Caenorhabditis elegans* (*C. elegans*). These forward genetic screens were based on a previously established dopamine transporter (DAT, or DAT-1 in worms)-associated phenotype known as Swip, or *Swimming-induced paralysis*, that requires the function of the DA receptor DOP-3 in worms. Hardaway and colleagues isolated two novel *dat-1* alleles (*vt21* and *vt25* lines) that displayed altered protein expression and trafficking to the cell surface. Interestingly, they also discovered two new lines, *vt25* and *vt29*, that do not contain mutations in *dat-1*. While *vt25* animals did not show a strong swip phenotype but was dependent on DOP-3, *vt29* was nearly 100% swip penetrant in a CAT-2 (tyrosine hydroxylase in worms)-dependent manner. Moreover, *vt25* and *vt29* also showed differential swip rescue responses to DAT overexpression, with both lines showing reduced dopamine content. Interestingly, *vt29* showed full resistance to 6-OHDA similar to *dat-1* animals, while *vt25* only showed partial resistance, indicating that the Swip-responsive gene in this line modulates DAT-1 activity. Therefore, this study further validates the Swip phenotype as an appropriate readout of DAT activity, while also offering novel genomic loci that may provide insight into therapeutics targeting new DA-modulating genes.

Modifier genes in epilepsy show sex-specificity and transcriptome differences

Nicole A Hawkins, & Kearney, JA (2012). Confirmation of an epilepsy modifier locus on mouse chromosome 11 and candidate gene analysis by RNA-Seq, *Genes Brain Behavior* 11(4): 452-460.

About 1% of the worldwide population experiences some form of epilepsy, which is a neurological disorder that often affects voltage-gated sodium channels. Several genes have been linked to the syndrome, particularly mutations in genes associated with voltage-gated sodium channels such as *SCN1A* and *SCN2A*. Using a transgenic mouse model (*Scna2^{Q54}*) characterized by slowed channel inactivation and assiduous sodium current, Nicole Hawkins and JA Kearney investigated in their 2012 study how variability in clinical phenotype relates to genetic strain background as additional genes modulate the mutated gene. In fact depending on the genetic strain, *Scna2^{Q54}* mice show varying degrees of epileptic activity including delayed sei-

zure onsets and improved survivals. Moreover, two loci on chromosome 11 (*Moe1*) and 19 (*Moe2*) have been characterized to influence severity of *Scna2^{Q54}* and fine mapping of *Moe2* has lead to *Kcnv2* being identified as a candidate gene. In the present study, focus on *Moe1* revealed two modifiers, which are sex-specific and include transcriptome differences and the encoding of single-nucleotide polymorphisms. Subsequent analysis of gene function and expression patterns suggested several modifier genes such as voltage-gated calcium channel subunits *Cacna 1g* and *Cacnb 1* as well as a transcription factor *Hlf*.

Sex-specific differences in the biologic mechanism of posttraumatic stress disorder

Asante Kamkwala, Norrholm, SD, Poole, JM, Brown, A, Donley, S, Duncan, E, Bradley, B, Ressler, KJ, & Jovanovic, T (2012). Dark-enhanced startle responses and heart rate variability in a traumatized civilian sample: putative sex-specific correlates of posttraumatic stress disorder, *Psychosomatic Medicine* 74(2): 153-159.

Prior research has linked trauma with an increased risk for posttraumatic stress disorder (PTSD). In a study in 2012, Asante Kamkwala and colleagues sought out to investigate characteristics of the biologic mechanisms of PTSD. They specifically looked at the dark-enhanced startle response (indicator for anxiety) and measures of heart rate variability and found that both measures may be sex-specific. They recruited 14 highly traumatized adults and recorded psychophysical responses of the eye bling reflex and heart rate variability during a startle paradigm with light and dark phases. Their data shows that female patients exhibited higher startle response magnitudes. Although there appeared to be no sex-difference in heart rate variability, male patients had increased high frequency heart rate variability compared to male controls. Kamkwala and colleagues suggest that the observed sex-differences may be associated with the limbic system, specifically with the sexually dimorphic bed nucleus of the stria terminalis, which is about 2.5 times larger in men.

CaMKII inhibition regulates medium spiny neuron activity

Jason R Klug, Mathur, BN, Kash, TL, Wang, HD, Matthews, RT, Robison, AJ, Anderson, ME, Deutch, AY, Lovinger, DM, Colbran, RJ, & Winder, DG (2012). Genetic inhibition of CaM-

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KII in dorsal striatal medium spiny neurons reduces functional excitatory synapses and enhances intrinsic excitability, *PLoS ONE* 7(9): e45323.

CaMKII is a kinase important for both the presynaptic and postsynaptic neuron functions. Though its role in learning and memory are well known, a more unexplored territory is the role of CaMKII in the striatum, where it is highly expressed in GABAergic medium spiny neurons (MSNs). The striatum is important in the pathophysiology of two dopamine-related diseases: Parkinson's disease (PD) and addiction. Since dopaminergic neurons modulate the firing of striatal MSNs, identifying the factors that regulate MSN firing behavior is important to understanding disease mechanisms. In Klug et al. (2012), the Winder lab explores the role of CaMKII in striatal MSNs using a transgenic mouse model that expresses an EGFP-linked CaMKII inhibitor peptide (EAC3I) in MSNs.

Inhibiting CaMKII resulted in a reduction in activity-dependent excitatory postsynaptic currents (mEPSCs), which was attributable to reduced synapse number. CaMKII promotes membrane trafficking of the AMPA receptor GluA1 subunit, which is important for activity-dependent synapse potentiation. MSNs from GluA1 knockout mice displayed similar problems with firing frequency. Additionally, CaMKII inhibition resulted in increased resting membrane potential and input resistance, markers of the cell's intrinsic excitability.

The alterations in MSN firing behavior and membrane potential, and possible differences in AMPA receptor trafficking that come with CaMKII inhibition, demonstrate that CaMKII is important for the membrane properties that allow MSNs to fire and propagate signals. In disorders such as PD and addiction, the firing of dopaminergic neurons in the striatum can be severely depleted, thereby altering the behavior of postsynaptic MSNs. Previous work has shown that CaMKII inhibition ameliorates disease symptoms in animal models of PD and addiction. A possible mechanism could be that the upregulation of MSN firing after CaMKII inhibition compensates for reduced input from dopaminergic neurons, indicating that CaMKII inhibition could someday serve as a therapy for PD and addiction.

A precise measure of manganese levels allows more accurate detection of modulators

Kevin K Kumar, Aboud, AA, Patel, DK, Aschner, M, & Bowman AB (2013). Optimization of Fluorescence Assay of Cellular Manganese Status for High Throughput Screening, *Journal of Biochemical and Molecular Toxicology* 27(1): 42-49.

The Bowman Lab has previously developed the Cellular Fura-2 Manganese Extraction Assay (CFMEA) for quantifying intracellular manganese (Mn) levels. In his article in the *Journal of Biochemical and Molecular Toxicity*, Kevin Kumar and colleagues explain how he has optimized this assay for high throughput screening (HTS) by adjusting concentrations and analysis tools for efficiency in identifying modulators with increasing signal-to-background ratios. In addition, Kumar and colleagues identified and decreased variability among samples, achieving a Z-factor required for HTS.

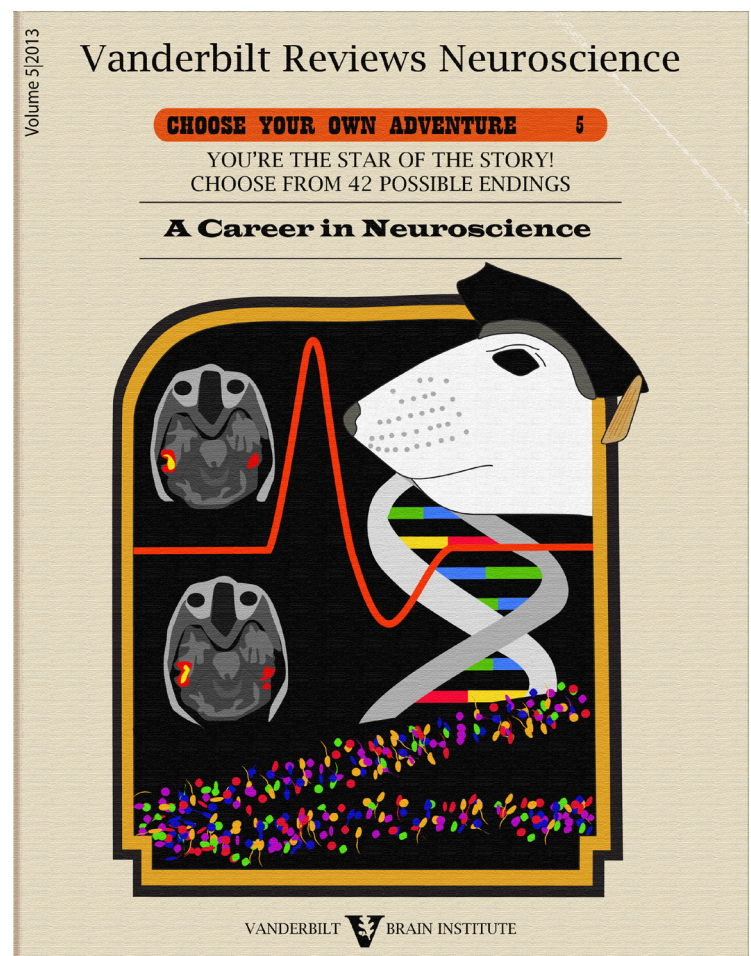
Mn neurotoxicity causes basal ganglia dysfunction and neurodegeneration. In addition, Mn exposure is an environmental risk factor for Parkinson's disease (PD), and modulation of neuronal Mn status has been observed in the pathogenesis of Huntington's disease (HD). Identification of modulators of Mn accumulation in neurons is a potential route to identify novel treatments for these debilitating neurodegenerative disorders. The authors used an immortalized murine striatal cell line exposed with physiologically relevant Mn concentrations. The first steps was to determine a cell density (cells/well) and exposure paradigm that would yield 50% quenching of the vehicle fura-2 signal. This was found to be 10,000 cells/well exposed for 60 minutes. Subsequently, the authors identified that variance was least in a fura-2 concentration of 0.75 μ M and that precision was highest in an extraction volume of 125 μ L. Finally, analyzing normalized fluorescent levels, rather than raw levels, decreased batch-to-batch variability. With these new parameters, CFMEA for HTS reliably detects cellular Mn levels. CFMEA is currently being utilized for HTS of small molecule libraries for modulators of manganese accumulation and toxicity.

On the Cover

A career in neuroscience is multi-faceted. This year's cover art was specifically chosen to represent all the wonderful career directions the field of neuroscience in general and our program in particular offers. Eva Sawyer graciously agreed to design a cover inspired by the *Choose Your Own Adventure* book series, as our students and faculty choose their own adventure, too.

Eva is a fourth year graduate student in Dr. Ken Catania's laboratory, where her research meets her interest in how evolution and ecology shape sensory systems. In Dr. Catania's lab, she studies the somatosensory system in the star-nosed mole, a small subterranean animal with an elaborate somatosensory organ on its nose. The star-nosed mole has 22 rays on its nose

that it uses to navigate its world. Eva's specific focus is on the ray representation along the somatosensory pathway and the preserved somatosensory map with emphasis on subcortical somatosensory nuclei such as the trigeminal brainstem complex and the ventral posterior medial nucleus of the thalamus. Eva investigates the ray-map using both neuroanatomical techniques such as specific immunohistochemical stains as well as neurophysiological techniques looking at multi-unit extracellular neuronal recordings during somatosensory stimulation of the mole rays. Eva also recently received a Ruth L. Kirschstein National Research Service Award to support her research endeavors.



Regulation of the Dopamine Transporter Through Trafficking-Dependent and -Independent Mechanisms

Daniel Bermingham

The dopamine transporter (DAT) is a vital protein involved in maintaining dopamine (DA) homeostasis in the brain by mediating reuptake of synaptic DA back into the presynaptic terminal. The regulation of this protein is critical for ensuring proper dopaminergic signaling, and its dysregulation can have dire consequences for a number of behaviors and neurological processes that are modulated by DA signaling. DAT activity can be regulated in both a positive and negative manner; the mechanisms of regulation involve both trafficking of the protein to and from the surface, as well as modulation of intrinsic transport activity independent of trafficking. Discussion of these trafficking-dependent and -independent modes of regulation will be the focus of this review.

Keywords: *Dopamine, transporter, presynaptic, trafficking, monoamines*

Introduction

The neurotransmitter dopamine (DA) is important for modulating many aspects of physiology and behavior, including motivation¹, movement², reward³ and attention⁴. Importantly, perturbations to the central dopaminergic systems are associated with many disease states such as Parkinson's disease², attention deficit/hyperactivity disorder (ADHD)⁵, schizophrenia² and addiction⁶. Therefore, understanding regulation of dopaminergic signaling is vital for understanding the pathophysiology of these diseases, as well as for gaining insights into the behaviors modulated by this neurotransmitter.

Presynaptic regulation of DA signaling is a dynamic process that can occur at many levels. These include regulation of DA synthesis by the enzyme tyrosine hydroxylase⁷, excitability of the presynaptic neuron⁸, and, most importantly for this review, regulation of the activity of the dopamine transporter (DAT), which acts to reuptake synaptic DA back into the presynaptic terminal to halt its signaling⁹. DAT is a twelve transmembrane domain protein that belongs to the sodium- and chloride-dependent (SLC6) family of transporters, of which the serotonin, norepinephrine, and GABA transporters are also members¹⁰. This family of transporters couple movement of their substrates across membranes to movement of sodium and chloride ions down their electrochemical gradients, allowing for

energy-independent movement of substrate across the membrane¹⁰. DAT itself is the target for therapeutic agents such as the ADHD medications methylphenidate (Ritalin) and amphetamine formulations (e.g., Adderall)¹¹, and variants in this protein have been found to be associated with ADHD¹²⁻¹⁵. This, along with the fact that DAT is the target of addictive psychostimulants such as cocaine⁹, amphetamine¹¹, and methamphetamine¹⁶, implicates the importance of this protein for the pathophysiology of many of the disease states associated with altered DA signaling.

Importantly, DAT is not a static protein, and tight regulation of its activity has been observed experimentally¹⁷⁻²⁰. Much research has demonstrated trafficking-dependent regulation of DAT through movement of the protein to and away from the surface by a number of interacting proteins and post-translational modifications, many of which will be discussed below. However, trafficking-independent regulation, where the intrinsic activity of DAT is modulated without trafficking to other compartments, has not been described as extensively. This review will focus on discussing these two broad classes of regulation that likely work in concert to achieve tight regulation of the activity of this vital protein.

Trafficking-dependent downregulation of DAT

The most robustly observed mechanism of DAT regulation,

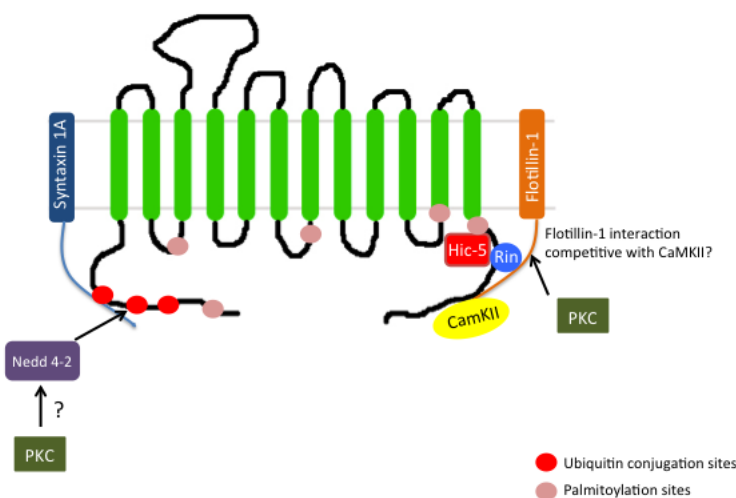


Figure 1: Modulation of DAT activity. DAT activity can be modulated by a number of interacting proteins, though which of these proteins may act together, or which may be exclusive and/or competitive is not entirely clear. Importantly, the site of interaction of Flotillin-1 has not been mapped, but has been placed based on evidence of potential competition with CaMKII. Post-translational modifications such as palmitoylation and ubiquitination also play important roles in regulating DAT activity and surface expression. Direct phosphorylation of the transporter has not been shown to regulate activity per se, but is important for regulating efflux-competent states of DAT.

downregulation of transport through trafficking of the transporter, is achieved through a large network of interacting proteins, the list of which is ever-expanding. Downregulation of DAT by Protein Kinase C (PKC) activation is among the most frequently studied mechanisms of DAT regulation, and is mediated by a number of proteins²¹⁻²⁵. Activation of PKC by phorbol esters such as PMA^{23, 24, 26} or by activation of G_q-coupled G-protein coupled receptors²⁷ induces a rapid reduction in surface DAT and a concomitant increase in intracellular DAT²⁸. This redistribution of DAT is mediated by a clathrin-dependent mechanism, as knockdown of either clathrin heavy chain or dynamin II inhibits both constitutive as well as PKC-triggered endocytosis^{26, 29}. However, direct phosphorylation of DAT by PKC is not required for internalization of the protein, suggesting that potentially another DAT-interacting protein is phosphorylated either by PKC or a downstream kinase to mediate this effect³⁰. It was recently reported by Cremona et al. that the membrane raft-associated protein Flotillin-1 was required for internalization of DAT upon PKC activation, and that phosphorylation of this protein at Ser315 is also required for this effect³¹. These results suggest that Flotillin-1 may be either a direct or indirect target for phosphorylation by the PKC signaling cascade. It was also observed that Flotillin-1 was required for maintaining membrane raft-association of DAT, potentially pointing toward the importance of this local-

ization for PKC-regulated endocytosis.

Further evidence of the importance of membrane microdomain targeting in PKC-regulated endocytosis has recently been reported. An ADHD-associated variant in the dopamine transporter (R615C) was shown to constitutively internalize and recycle back to the membrane at an accelerated rate, and was insensitive to PKC- and amphetamine (AMPH)-stimulated endocytosis¹². Importantly, this variant showed reduced association with Flotillin-1 and reduced membrane raft localization, as measured by colocalization with an Alexa 647-conjugated cholera toxin B, which marks membrane raft-enriched GM1 Ganglioside. This reduction in association with Flotillin-1 and membrane rafts is contrasted by a significant increase in association with Ca²⁺/calmodulin-dependent protein kinase II (CaMKII). It is possible that the reduced association between the 615C variant and Flotillin-1 may be due to the fact that CamKII and Flotillin-1 interactions with DAT are competitive due to overlapping sites of interaction. Therefore, an increased affinity for CaMKII might prevent Flotillin-1 interactions and raft association, though this is just speculation at this point since the site of interaction between Flotillin-1 and DAT has not been mapped (**Figure 1**). Regardless, these altered associations in this PKC- and AMPH-induced trafficking-insensitive DAT variant certainly provide further support for the importance of the Flotillin-1 interaction and raft localization for

Membrane rafts:

Cholesterol-rich microdomains that are thought to organize signaling molecules into discrete regions of the plasma membrane.

Clathrin:

A protein that mediates endocytosis of vesicles, a process that is important for many cellular physiological processes, including internalization of cell-surface proteins.

PKC and AMPH-triggered endocytosis of the transporter.

Another recent study identified another DAT-interacting protein, the plasma membrane-associated GTPase, Rin, which is involved in mediating the effects of PKC on DAT³². Utilizing yeast two-hybrid, GST pull down, and FRET approaches, it was demonstrated that Rin directly associates with residues 587-596 of the DAT C-terminus, and that expression and activity of Rin is required for PKC-triggered endocytosis of the transporter. It was also shown that Rin and DAT interactions increase with PKC activation. Furthermore, Rin/DAT interactions were shown to occur preferentially in membrane rafts, again pointing toward the importance of this localization for PKC-regulated DAT trafficking. Unfortunately, attempts at demonstrating an interaction *in vivo* in dopaminergic neurons were unsuccessful due to technical issues, so the relevance of this interaction is unknown. Importantly, Rin is present in striatal tissue and a number of DAergic cell lines, supporting a potential role in endogenous DAT regulation. The role that Rin plays in DAT regulation is unclear, though it is possible that it helps regulate movement of DAT into or out of membrane raft microdomains, a process that has been suggested to be important for PKC regulation of DAT. However, more work is necessary to clarify just what role this interaction plays in DAT trafficking by PKC-dependent pathways.

It has been also been shown that PKC activation results in ubiquitination of DAT, and that three lysine residues in the N-terminus of DAT that act as ubiquitin conjugation sites are required for PKC-dependent endocytosis of the transporter³³. Utilizing a large-scale RNAi screen for genes involved in PKC-dependent endocytosis, Sorkina et al. identified the E3 ubiquitin ligase, Nedd4-2, as being necessary for this process, as loss of this protein by siRNA-mediated knockdown abolished DAT ubiquitination^{34,35}. These data suggest that PKC activation induces ubiquitination of DAT by Nedd4-2, and that this process is necessary for PKC-regulated endocytosis of the transporter. It is possible that PKC itself regulates Nedd4-2 activity or recruitment to DAT, though this has not been shown experimentally. Also, the interplay between this Nedd4-2 dependent mechanism and the interaction with Flotillin-1 or Rin has yet to be investigated.

Another interesting DAT-interacting protein that appears to be involved in trafficking of the transporter is the focal adhesion protein Hic-5. This interaction was initially identified by the yeast two-hybrid system, and verified by co-

immunoprecipitation from striatal extracts and immunostaining that showed presynaptic colocalization of these two proteins³⁶. Co-expression of DAT and Hic-5 in HEK293 cells resulted in a decrease in transport and surface expression of DAT compared to expression of DAT alone. These results suggest that the association between DAT and Hic-5 may be important for surface trafficking of DAT. In further support of this, work done in platelets by Carneiro and Blakely has shown that Hic-5 also associates with the serotonin transporter (SERT)³⁷. Interestingly, treatment with PMA, which also induces internalization of SERT, increases association between SERT and Hic-5 at times that correlate with decreases in SERT surface expression. If a similar mechanism occurs with Hic-5 and DAT, it may be the case that Hic-5 is involved in the network of proteins mediating PKC-regulated DAT endocytosis, though direct evidence for this has yet to be presented. Also worth noting is the fact that this interaction between Hic-5 and DAT occurs in the membrane-proximal region of the C-terminus DAT, very close to the interaction site of Rin. It is possible that these proteins function together to regulate DAT trafficking, or may be competitive for binding to DAT and may mediate different forms of DAT regulation, though these possibilities have yet to be investigated.

Importantly, it should be noted that the timing and cellular contexts of these various PKC-related mechanisms of DAT endocytosis remain unclear. For instance, it is unknown if these proteins work in tandem in DAergic terminals through a single PKC-related mechanism of DAT endocytosis, or if there is exclusivity between these different interactors depending on factors such as the active signaling pathways or the membrane microdomain localization of DAT. Foster et al. demonstrated that treatment with β -PMA causes PKC- α to be recruited to membrane rafts and may preferentially regulate endocytosis of DAT in these fractions through some of the mechanisms mentioned above. However, it is possible that activation of other PKC isoforms by other signaling pathways may lead to modulation of DAT endocytosis by entirely separate mechanisms. Sakrikar et al. have proposed a model based on work with the R615C DAT variant that postulates that DAT endocytosis occurs in both regulated and constitutive manners that depend upon microdomain localization. In this model, PKC regulates endocytosis from membrane rafts, whereas DAT localized outside of these rafts endocytoses in a constitutive manner. This is consistent with the finding that PKC- α moves into raft fractions after PMA treatment, but Foster et al. also showed that PKC activation does not

reduce surface DAT in raft fractions. These inconsistencies may reflect the different cell types used or the methods employed, or it may be the case that PKC- α mediates a trafficking-independent mode of DAT regulation, and other isoforms, such as PKC- β underlie mechanisms of trafficking-dependent regulation of DAT. Clearly, the picture of PKC regulation of DAT is quite complicated at this point, and this complication is made worse by the inconsistency in cell lines and techniques employed by various groups. Regardless, it certainly seems as if PKC regulation of DAT surface expression may actually occur through a number of mechanisms; future work on the interaction of these various PKC-related mechanisms, as well as the PKC subtypes mediating these effects, may clarify where and when these different pathways may modulate DAT surface expression *in vivo*.

The neuronal SNARE protein Syntaxin 1A is another DAT regulatory partner whose interaction appears to have a number of functional consequences for DAT activity. Tissue treatment with Botulinum Neurotoxin C (BoNT/C) results in degradation of Syntaxin 1A, and treatment of rat striatal tissue with this toxin causes an increase in DA uptake and DAT surface levels³⁸. Conversely, heterologous co-expression of these two proteins results in decreased DA uptake and DAT surface levels compared to DAT expression alone. Together, these results suggest that the interaction between these two proteins promotes suppression of DA uptake through reduced DAT surface levels. Importantly, PMA-induced endocytosis of DAT was intact even with BoNT/C treatment, suggesting that the Syntaxin 1A/DAT interaction is not required for PKC-triggered endocytosis of the transporter. This interaction is interesting because it may have relevance for the localization of DAT to sites of release, as Syntaxin 1A is an important member of the SNARE complex that mediates vesicular fusion and neurotransmitter release. What the relevance of this interaction is remains unclear, but it is important to note that Syntaxin 1A is also essential for facilitating an

efflux-competent state of DAT and decreases DAT channel currents, which, combined with the apparent effects on trafficking, likely have important functional consequences for how DAT behaves endogenously.

Trafficking-dependent upregulation of DAT

There are a number of signaling pathways that have been shown to promote surface expression of DAT through trafficking-dependent mechanisms. For instance, insulin increases dopamine uptake by increasing surface expression of DAT, and this increase can be blocked by inhibition of Phosphatidylinositol 3-kinase (PI3K)³⁹. Importantly, PI3K inhibition also reduces basal uptake and surface expression in a dynamin-dependent manner, suggesting that this regulation is vital for opposing endocytosis of the protein by other pathways such as PKC, etc³⁹. Also vital for this process is the kinase Akt, whose level of phosphorylation increases upon insulin stimulation⁴⁰. Constitutively active Akt prevents AMPH-induced endocytosis of DAT, and inhibition of Akt reduces DAT uptake and surface expression, supporting its role in the insulin/PI3K pathway that promotes surface expression of DAT.

DAT activity can also be modulated by interactions with the D2 dopamine receptor (D2R). D2Rs are expressed both postsynaptically and presynaptically, and can be alternatively spliced into short and long isoforms (D2S and D2L), with the D2S isoform as the predominant presynaptic isoform. It has been observed that D2-deficient animals have reduced DAT function⁴¹, and heterologous expression of D2 and DAT suggests that this D2 effect on DAT occurs cell-autonomously⁴². By co-expressing D2 and DAT, it has been shown that these proteins directly interact^{43,44}, and increases uptake and DAT surface expression in heterologous systems compared to singly transfected DAT⁴³. In addition to this direct interaction, there also appears to be a functional interaction between these two proteins. When DAT and D2S were co-expressed in cells, the D2 agonist quinpirole significantly increased dopamine

SNARE:

A family of proteins that mediate docking and fusion of vesicles to the plasma membrane, which allows for the release of neurotransmitter.

CANDIDATE REVIEWS

uptake and DAT surface expression, and a specific ERK1/2 antagonist could block this increase⁴⁴. This suggests that D2 modulation of DAT activity likely occurs via at least two mechanisms: a direct interaction of the proteins promoting increased surface expression of the transporter, as well as a functional interaction that involves D2-dependent ERK1/2 signaling cascades.

An important thing to keep in mind when considering how these signaling pathways influence DAT regulation is that these pathways overlap and interact in many ways. For instance, in addition to activating PI 3-kinase, insulin can also activate PKC signaling pathways via PLC gamma, which would presumably oppose PI 3-kinase upregulation of DAT. It is likely that compartmentalization of DAT with its regulatory partners helps organize these regulatory mechanisms in manners that aren't well understood, and it should always be kept in mind that much of the work on DAT regulation is done in heterologous expression systems that may not accurately reflect the environment in which DAT is endogenously expressed. Because of this, care must be taken in how the results of such studies are interpreted until they are repeated in endogenous DAT-expressing systems.

Trafficking-independent downregulation of DAT

In recent years, it has become clear that trafficking of DAT alone cannot explain regulation of its activity, and that there are trafficking-independent modes of regulation of this protein. Foster et al. initially reported findings that suggest that reduction in DAT activity by PMA-induced PKC activation could only partially be explained by internalization of the transporter⁴⁵. They demonstrated this by inhibiting clathrin-mediated endocytosis using either the chemical inhibitor Concanavalin A (Con A) or a dominant-negative dynamin. This inhibition was sufficient to prevent internalization of the protein, but only partially prevented PKC-induced downregulation of transport activity. Also, using a cholesterol depletor, methyl- β -cyclodextrin (M β C), it was demonstrated that PKC-induced downregulation was also partially inhibited by loss of cholesterol, but DAT internalization was about equivalent to PMA treatment alone, suggesting a loss of PKC-induced downregulation that was independent of trafficking. In further support of this, another study showed that PKC-induced downregulation of DAT in synaptosomes occurs even in the presence of high sucrose, which blocks endocytosis⁴⁶. These lines of evidence suggest that PKC causes a decrease in DAT activity via a mechanism that does not require internalization of

the protein.

The association of DAT with cholesterol-rich membrane rafts has been well characterized, and this association can be decreased by M β C treatment and increased by treatment with water-soluble cholesterol (wsChol). By augmenting membrane cholesterol of DAT-transfected HEK cells, as well as striatal synaptosomes with wsChol, Hong and Amara demonstrated that binding of cocaine-analogs [¹²⁵I] RTI-55 and [³H]WIN35428 was significantly increased compared to untreated controls⁴⁷. They also showed that binding of Maleimide-PEO₂-biotin to DAT, which specifically recognizes sulfhydryl (-SH) moieties on surface-accessible cysteine residues, is increased with no change in total surface DAT. Using site-directed mutagenesis, the site of increased reactivity was found to be cysteine 306, and its increased availability to the -SH-specific biotin was attributed to an increase in outward conformation of the DAT protein. This cholesterol-dependent change in DAT conformation may underlie some of the modulation of DAT activity by altered membrane cholesterol content, and may represent a mechanism through which altered cholesterol-rich membrane raft association of DAT can regulate transport, independent of trafficking to and away from the cell surface.

Trafficking-independent upregulation of DAT

Though very little direct evidence for trafficking-independent upregulation of DAT activity has been observed, there are a few findings that indicate that this may occur. Foster and Vaughn have shown that DAT is palmitoylated, and that this modification has functional consequences for transport⁴⁶. By inhibiting palmitoyl acyltransferase using 2-bromopalmitate (2BP), which prevents protein palmitoylation, they showed that blocking palmitoylation of DAT induces a rapid reduction in transport with no changes in DAT protein or surface levels at early time points. It is important to note that 2BP inhibited palmitoylation of DAT by about 40% within 45 minutes, suggesting that palmitate turnover is quite rapid, and that this modification may be used to acutely regulate DAT activity. If this is the case, then palmitoylation and depalmitoylation may represent mechanisms by which DAT kinetics can be rapidly up- or downregulated, independent of trafficking of the protein. Additionally, since palmitoyl groups can mediate interactions between proteins and membrane lipids, the palmitoylation status of DAT may impact its membrane microdomain localization and, therefore, its regulation, providing a potential mechanism through which activity

and surface expression may be altered by this modification.

Further evidence from the related transporters SERT, as well as the norepinephrine transporter (NET), demonstrates that trafficking-independent upregulation of transporters does occur, and it is tempting to think that this mode of regulation may generalize to other family members such as DAT. For instance, activation of adenosine A3 receptors on serotonergic neurons increases SERT activity by a mechanism that is partially independent of trafficking to the surface⁴⁸⁻⁵⁴. The signaling pathways involved have been worked out, and it appears that activation of PKG and p38 MAP kinase underlies this upregulation⁵⁰⁻⁵³. In the case of NET, it has been demonstrated that insulin stimulation also activates p38 MAP kinase, and this activation induces upregulation of transport without any significant increase in surface expression^{55,56}. This raises the question of whether mechanisms that upregulate DAT activity may be doing so via similar mechanisms that may be due in part to trafficking-independent activation. It seems reasonable to think that perhaps trafficking-independent upregulation of DAT has not been reported because of a lack of temporal resolution in monitoring DAT surface expression. For example, many studies that showed increased uptake and a concomitant increase in DAT surface levels only looked at DAT surface levels at later time points, and may have missed an earlier increase in DAT activity prior to trafficking of DAT to the surface. Hopefully, future work will begin to clarify whether this mode of regulation is indeed employed to regulate DAT, as has been shown for related transporters.

Conclusions

Understanding how DA homeostasis is maintained and dynamically regulated is essential for gaining insights into how DA mediates its effects on behavior, and how dysfunction of this system can lead to diseases such as ADHD, schizophrenia, and addiction. At the center of DA signaling regulation is DAT, whose activity is vital for controlling the proper amount of synaptic DA during neurotransmission. Because DAT dysfunction has been shown to be associated with a subset of individuals with diseases such as ADHD, it is imperative that there be a focus on studying how this protein is regulated in order to understand its role in these disease states, as well as in normal cognition and behavior. As this review has shown, there are many different modes of DAT regulation involving trafficking and modulation of intrinsic transport activity, though when and where these pathways may regulate DAT *in vivo* and

how they may interact with one another remains poorly understood. As these regulatory networks of proteins are worked out further, it will hopefully expand our understanding of DA signaling in the brain and open up avenues for treating individuals with DA-related brain disorders.

References

1. Wise RA. Dopamine, learning and motivation. (2004) *Nat Rev Neurosci.* 5 (6): 483-494
2. Seeman P, Niznik HB. Dopamine receptors and transporters in parkinson's disease and schizophrenia. (1990) *The FASEB Journal.* 4 2737-2744
3. Fiorillo CD, Tobler PN, Schultz W. Discrete coding of reward probability and uncertainty by dopamine neurons. (2003) *Science.* 299 (5614): 1898-1902
4. Puumala T, Sirvio J. Changes in activities of dopamine and serotonin systems in the frontal cortex underlie poor choice accuracy and impulsivity of rats in an attention task. (1998) *Neuroscience.* 83 (2): 489-499
5. Cook EH, Jr., Stein MA, Krasowski MD, Cox NJ, Olkon DM, Kieffer JE, Leventhal BL. Association of attention-deficit disorder and the dopamine transporter gene. (1995) *Am J Hum Genet.* 56 (4): 993-998
6. Berke JD, Hyman SE. Addiction, dopamine, and the molecular mechanisms of memory. (2000) *Neuron.* 25 (3): 515-532.
7. Zhang D, Kanthasamy A, Yang Y, Anantharam V, Kanthasamy A. Protein kinase cdelta negatively regulates tyrosine hydroxylase activity and dopamine synthesis by enhancing protein phosphatase-2a activity in dopaminergic neurons. (2007) *J Neurosci.* 27 (20): 5349-5362
8. Hu G, Duffy P, Swanson C, Ghasemzadeh MB, Kalivas PW. The regulation of dopamine transmission by metabotropic glutamate receptors. (1999) *J Pharmacol Exp Ther.* 289 (1): 412-416
9. Giros B, Mestikawy SE, Bertrand L, Caron MG. Cloning and functional characterization of a cocaine-sensitive dopamine transporter. (1991) *Federation of Experimental Biological Sciences.* 295 149-154
10. Chen NH, Reith ME, Quick MW. Synaptic uptake and beyond: The sodium- and chloride-dependent neurotransmitter transporter family slc6. (2004) *Pflugers Arch.* 447 (5): 519-531
11. Seeman P, Madras BK. Anti-hyperactivity medication: Methylphenidate and amphetamine. (1998) *Mol Psychiatry.* 3 (5): 386-396
12. Sakrikar D, Mazei-Robison MS, Mergy MA, Richtand NW, Han Q, Hamilton PJ, Bowton E, Galli A, Veenstra-Vanderweele J, Gill M, Blakely RD. Attention deficit/hyperactivity disorder-derived coding variation in the dopamine transporter disrupts microdomain targeting and trafficking regulation. (2012) *J Neurosci.* 32 (16): 5385-5397

This paper is important because it demonstrates how the signaling pathways and regulation of a ADHD-associated DAT variant are altered, and potentially brings the field closer to understanding how alterations in DA signaling regulation can contribute to this disease.

CANDIDATE REVIEWS

It also furthers our understanding of the importance of membrane microdomain localization for DAT function.

13. Bowton E, Saunders C, Erreger K, Sakrikar D, Matthies HJ, Sen N, Jessen T, Colbran RJ, Caron MG, Javitch JA, Blakely RD, Galli A. Dysregulation of dopamine transporters via dopamine d2 autoreceptors triggers anomalous dopamine efflux associated with attention-deficit hyperactivity disorder. (2010) *J Neurosci.* 30 (17): 6048-6057
14. Friedel S, Saar K, Sauer S, Dempfle A, Walitza S, Renner T, Romanos M, Freitag C, Seitz C, Palmason H, Scherag A, Winde-muth-Kieselbach C, Schimmelmann BG, Wewetzer C, Meyer J, Warnke A, Lesch KP, Reinhardt R, Herpertz-Dahlmann B, Linder M, Hinney A, Remschmidt H, Schafer H, Konrad K, Hubner N, Hebebrand J. Association and linkage of allelic variants of the dopamine transporter gene in adhd. (2007) *Mol Psychiatry.* 12 (10): 923-933
15. Asherson P, Brookes K, Franke B, Chen W, Gill M, Ebstein RP, Buitelaar J, Banaschewski T, Sonuga-Barke E, Eisenberg J, Manor I, Miranda A, Oades RD, Roeyers H, Rothenberger A, Sergeant J, Steinhausen HC, Faraone SV. Confirmation that a specific haplotype of the dopamine transporter gene is associated with combined-type adhd. (2007) *Am J Psychiatry.* 164 (4): 674-677
16. Fleckenstein AE, Metzger RR, Wilkins DG, Gibb JW, Hanson GR. Rapid and reversible effects of methamphetamine on dopamine transporters. (1997) *J.Pharmacol.Exp.Ther.;* 282 (2): 834-838
17. Mortensen OV, Amara SG. Dynamic regulation of the dopamine transporter. (2003) *Eur J Pharmacol.* 479 (1-3): 159-170
18. Zahniser NR, Sorkin A. Rapid regulation of the dopamine transporter: Role in stimulant addiction? (2004) *Neuropharmacology.* 47 Suppl 1 80-91
19. Gulley JM, Zahniser NR. Rapid regulation of dopamine transporter function by substrates, blockers and presynaptic receptor ligands. (2003) *Eur J Pharmacol.* 479 (1-3): 139-152
20. Foster JD, Cervinski MA, Gorentla BK, Vaughan RA. Regulation of the dopamine transporter by phosphorylation. (2006) *Handb Exp Pharmacol.* (175): 197-214
21. Zhang L, Coffey LL, Reith MEA. Regulation of the functional activity of the human dopamine transporter by protein kinase c. (1997) *Biochemical Pharmacology.* 53 (5): 677-688
22. Doolen S, Zahniser NR. Conventional protein kinase c isoforms regulate human dopamine transporter activity in xenopus oocytes. (2002) *FEBS Lett.* 516 (1-3): 187-190
23. Loder MK, Melikian HE. The dopamine transporter constitutively internalizes and recycles in a protein kinase c-regulated manner in stably transfected pc12 cell lines. (2003) *J Biol Chem.* 278 (24): 22168-22174
24. Vaughan RA, Huff RA, Uhl GR, Kuhar MJ. Protein kinase c-mediated phosphorylation and functional regulation of dopamine transporters in striatal synaptosomes. (1997) *The Journal of Biological Chemistry.* 272 (24): 15541-15546
25. Zhu SJ, Kavanaugh MP, Sonders MS, Amara SG, Zahniser NR. Activation of protein kinase c inhibits uptake, currents and binding associated with the human dopamine transporter expressed in xenopus oocytes. (1997) *J. Pharmacol. Exp. Ther.;* 282 (3): 1358-1365
26. Daniels GM, Amara SG. Regulated trafficking of the human dopamine transporter. Clathrin-mediated internalization and lysosomal degradation in response to phorbol esters. (1999) *J Biol Chem.* 274 (50): 35794-35801
27. Granas C, Ferrer J, Loland CJ, Javitch JA, Gether U. N-terminal truncation of the dopamine transporter abolishes phorbol ester- and substance p receptor-stimulated phosphorylation without impairing transporter internalization. (2003) *J Biol Chem.* 278 (7): 4990-5000
28. Pristupa ZB, McConkey F, Liu F, Man HY, Lee FJ, Wang YT, Niznik HB. Protein kinase-mediated bidirectional trafficking and functional regulation of the human dopamine transporter. (1998) *Synapse.* 30 (1): 79-87
29. Sorkina T, Hoover BR, Zahniser NR, Sorkin A. Constitutive and protein kinase c-induced internalization of the dopamine transporter is mediated by a clathrin-dependent mechanism. (2005) *Traffic.* 6 (2): 157-170
30. Chang MY, Lee SH, Kim JH, Lee KH, Kim YS, Son H, Lee YS. Protein kinase c-mediated functional regulation of dopamine transporter is not achieved by direct phosphorylation of the dopamine transporter protein. (2001) *J Neurochem.* 77 (3): 754-761
31. **Cremona ML, Matthies HJ, Pau K, Bowton E, Speed N, Lute BJ, Anderson M, Sen N, Robertson SD, Vaughan RA, Rothman JE, Galli A, Javitch JA, Yamamoto A. Flotillin-1 is essential for plkc-triggered endocytosis and membrane microdomain localization of dat. (2011) *Nat Neurosci.***
This is a landmark paper because it is one of the first to really demonstrate the importance of membrane microdomain localization of DAT for its regulation and function, and identifies Flotillin-1 as a potential target for PKC that may play a role in mediating endocytosis of DAT by this pathway.
32. Navaroli DM, Stevens ZH, Uzelac Z, Gabriel L, King MJ, Lifshitz LM, Sitte HH, Melikian HE. The plasma membrane-associated gtpase rin interacts with the dopamine transporter and is required for protein kinase c-regulated dopamine transporter trafficking. (2011) *The Journal of Neuroscience.* 31 (39): 13
33. Miranda M, Dionne KR, Sorkina T, Sorkin A. Three ubiquitin conjugation sites in the amino terminus of the dopamine transporter mediate protein kinase c-dependent endocytosis of the transporter. (2007) *Mol Biol Cell.* 18 (1): 313-323
34. Sorkina T, Miranda M, Dionne KR, Hoover BR, Zahniser NR, Sorkin A. Rna interference screen reveals an essential role of nedd4-2 in dopamine transporter ubiquitination and endocytosis. (2006) *J Neurosci.* 26 (31): 8195-8205
35. Vina-Vilaseca A, Sorkin A. Lysine 63-linked polyubiquitination of the dopamine transporter requires ww3 and ww4 domains of nedd4-2 and ube2d ubiquitin-conjugating enzymes. (2010) *J Biol Chem.* 285 (10): 7645-7656
36. Carneiro AM, Ingram SL, Beaulieu JM, Sweeney A, Amara SG, Thomas SM, Caron MG, Torres GE. The multiple lim domain-containing adaptor protein hic-5 synaptically colocalizes and interacts with the dopamine transporter. (2002) *J Neurosci.* 22 (16): 7045-7054
37. Carneiro AM, Blakely RD. Serotonin-, protein kinase c-, and hic-5-associated redistribution of the platelet serotonin transporter. (2006) *J Biol Chem.* 281 (34): 24769-24780
38. Cervinski MA, Foster JD, Vaughan RA. Syntaxin 1a regulates dopamine transporter activity, phosphorylation and surface expres-

- sion. (2010) *Neuroscience*. 170 408-416
39. Carvelli L, Moron JA, Kahlig KM, Ferrer JV, Sen N, Lechleiter JD, Leeb-Lundberg LM, Merrill G, Lafer EM, Ballou LM, Shippenberg TS, Javitch JA, Lin RZ, Galli A. Pi 3-kinase regulation of dopamine uptake. (2002) *J Neurochem*. 81 (4): 859-869.
40. Garcia BG, Wei Y, Moron JA, Lin RZ, Javitch JA, Galli A. Akt is essential for insulin modulation of amphetamine-induced human dopamine transporter cell-surface redistribution. (2005) *Mol Pharmacol*. 68 (1): 102-109
41. Dickinson SD, Sabeti J, Larson GA, Giardina K, Rubinstein M, Kelly MA, Grandy DK, Low MJ, Gerhardt GA, Zahniser NR. Dopamine d2 receptor-deficient mice exhibit decreased dopamine transporter function but no changes in dopamine release in dorsal striatum. (1999) *J Neurochem*. 72 (1): 148-156
42. Mayfield RD, Zahniser NR. Dopamine d2 receptor regulation of the dopamine transporter expressed in xenopus laevis oocytes is voltage-independent. (2001) *Mol Pharmacol*. 59 (1): 113-121
43. Lee FJ, Pei L, Moszczynska A, Vukusic B, Fletcher PJ, Liu F. Dopamine transporter cell surface localization facilitated by a direct interaction with the dopamine d2 receptor. (2007) *Embo J*. 26 (8): 2127-2136
44. **Bolan EA, Kivell B, Jaligam V, Oz M, Jayanthi LD, Han Y, Sen N, Urizar E, Gomes I, Devi LA, Ramamoorthy S, Javitch JA, Zapata A, Shippenberg TS. D2 receptors regulate dopamine transporter function via an extracellular signal-regulated kinases 1 and 2-dependent and phosphoinositide 3 kinase-independent mechanism. (2007) *Mol Pharmacol*. 71 (5): 1222-1232**
- D2 autoreceptor regulation of DAT mediates an important feedback loop for regulating DA signaling, and this paper expands our understanding of the functional interaction between these proteins, and uncovers the signaling pathways involved in activity-dependent regulation of DAT by this receptor.*
45. **Foster JD, Adkins SD, Lever JR, Vaughan RA. Phorbol ester induced trafficking-independent regulation and enhanced phosphorylation of the dopamine transporter associated with membrane rafts and cholesterol. (2008) *J Neurochem*. 105 1683-1699.**
- This paper challenges the dogma that PKC regulation of DAT occurs solely through endocytosis of the transporter, and opens up the conversation on trafficking-independent regulation of this transporter, even through modes of regulation that were once thought to be trafficking-dependent.*
46. Foster JD, Vaughan RA. Palmitoylation controls dopamine transporter kinetics, degradation, and protein kinase c-dependent regulation. (2011) *J Biol Chem*. 286 (7): 5175-5186
47. Hong WC, Amara SG. Membrane cholesterol modulates the outward facing conformation of the dopamine transporter and alters cocaine binding. (2010) *J Biol Chem*. 285 (42): 32616-32626
48. Miller KJ, Hoffman BJ. Adenosine a3 receptors regulate serotonin transport via nitric oxide and cgmp. (1994) *J Biol Chem*. 269 (44): 27351-27356
49. Okada M, Kawata Y, Murakami T, Wada K, Nizuno K, Kondo T, Kaneko S. Differential effects of adenosine receptor subtypes on release and reuptake of hippocampal serotonin. (1999) *European Journal of Neuroscience*. 11 1-9
50. Blakely RD, Zhu C, Hewlett W, Dostmann WR, Buck E, Jayanthi LD, Ramamoorthy S. Protein kinase g-mediated phosphorylation of sert is required for adenosine receptor triggered stimulation of serotonin transporters. (2004).
51. Daws LC, Blakely RD, Munn JL, Zhu CB, Davis N, Owens WA. Evidence that adenosine receptor-linked protein kinase g and p38mapk acutely regulate the serotonin transporter in vivo. (2004) *American College of Neuropsychopharmacology*.
52. Zhu CB, Hewlett WA, Feoktistov I, Biaggioni I, Blakely RD. Adenosine receptor, protein kinase g, and p38 mitogen-activated protein kinase-dependent up-regulation of serotonin transporters involves both transporter trafficking and activation. (2004) *Mol Pharmacol*. 65 (6): 1462-1474
53. Zhu CB, Steiner JA, Munn JL, Daws LC, Hewlett WA, Blakely RD. Rapid stimulation of presynaptic serotonin transport by a3 adenosine receptors. (2007) *J Pharmacol Exp Ther*. 322 (1): 332-340
54. Zhu C, Lindler KM, Campbell NG, Sutcliffe JS, Hewlett WA, Blakely RD. Colocalization and regulated physical association of presynaptic serotonin transporters with a3 adenosine receptors. (2011) *Mol Pharm*. in press
55. Filgewicz DP, Bentson K, Ocrant I. The effect of insulin on norepinephrine uptake by pc12 cells. (1993) *Brain Research Bulletin*. 32 425-431
56. Apparsundaram S, Sung U, Price RD, Blakely RD. Trafficking-dependent and -independent pathways of neurotransmitter transporter regulation differentially involving p38 mitogen-activated protein kinase revealed in studies of insulin modulation of norepinephrine transport in sk-n-sh cells. (2001) *J Pharmacol Exp Ther*. 299 (2): 666-677

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The Waiting Game: How Studying Anticipatory Processing Can Provide New Insight into the Neural Circuit of Dysfunction in Anxiety Disorders

Jacqueline Clauss

Anticipation of upcoming aversive or threatening events is a highly adaptive psychological process. During anticipation, individuals consider the current environment and prior memories to initiate cognitive and motor processes. Anticipation of aversive stimuli has been studied in a number of neuroimaging paradigms, and results have shown that anticipation results in increased activation in a network of brain regions. This network includes emotion reactivity regions—the amygdala and insula, which signal threat and arousal—and emotion regulation regions—the dorsal anterior cingulate cortex and dorsolateral prefrontal cortex, which signal action and cognitive control. Balance between emotion reactivity regions and emotion regulation regions is necessary for the successful preparation for an upcoming aversive event. Anticipation can also be maladaptive, such as in anxiety disorders, and understanding anticipatory anxiety can provide new insight into dysfunction in the neural circuit in anxiety disorders. Patients with anxiety disorders may engage anticipatory processes to neutral or mildly aversive stimuli. When patients with anxiety disorders anticipate aversive stimuli, they have greater activation of emotion reactivity regions and less activity of emotion regulation regions, relative to controls. Among patients with anxiety disorders, patients who have greater activation of emotion regulation regions typically experience fewer symptoms. Individuals with social phobia typically experience extreme anticipatory anxiety prior to social situations; anticipatory anxiety has been studied using speech anticipation paradigms. In patients with social phobia, relative to controls, anticipation of social stimuli is associated with increased activity of emotion reactivity regions, and less activity of emotion regulation regions in patients with social phobia, relative to controls. Studying anticipation of social stimuli in patients with social phobia may lead to greater understanding of the pathophysiology of social phobia and help to identify new targets for treatment and prevention.

Keywords: *Anxiety, anticipation, fear, amygdala, dorsal anterior cingulate, dorsolateral prefrontal cortex, insula, social phobia*

Anticipation of Aversive Events

An upcoming aversive event, such as a having a thesis committee meeting, giving a scientific talk, or taking a test triggers anticipation of that event. Anticipation is often adaptive; during the anticipation of an event, mental and physical preparation takes place. Anticipation of aversive stimuli promotes survival in a changing environment¹. Anticipation of aversive events can be broken down into a number of overlapping processes (see **Figure 1**): A.) orienting and threat detection;² B.) memory recall and evaluation;³ and C.) motor preparation and cognitive control⁴. During anticipation of emotional stimuli, physiologic reactivity is heightened, suggesting emotional arousal^{5,6}. Anticipatory processes may modify behavior, and avoid, prepare for, or

alter the aversive event. Cognitive processes, such as reappraisal, distraction, or emotion suppression, may also reduce anxiety and allow for mental preparation for the aversive event. Better understanding of the neural processes of anticipation of aversive stimuli may provide insight into these adaptive responses

Anticipation of aversive events is mediated by emotion reactivity and emotion regulation brain regions. During anticipation of aversive events, common emotion reactivity regions activated include the amygdala^{2,4,7-11} and the insula^{2,4,8-17}, and common emotion regulation regions activated include the dorsolateral prefrontal cortex (dlPFC)^{1,4,9,10,13,14,17} and the dorsal anterior cingulate cortex (dACC)^{1,2,4,11-13,15,16} (see **Figure 2**). The amygdala detects and attends to aver-

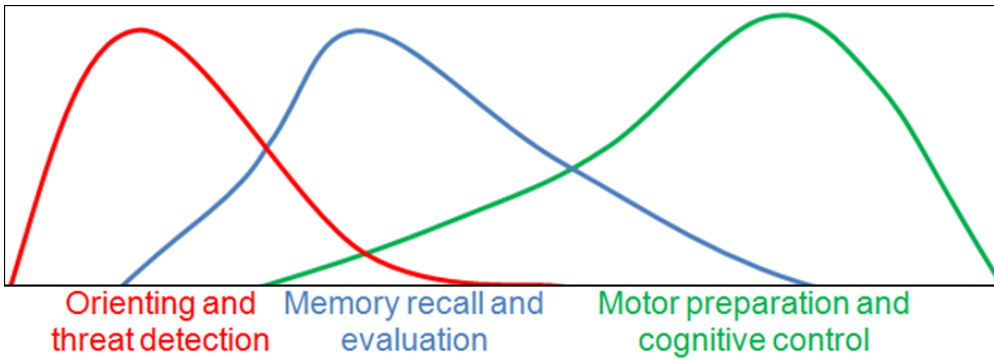


Figure 1: *Phases of anticipatory processing.* Anticipation of aversive events involves several distinct, yet overlapping processes, including orienting and threat detection, memory recall and evaluation, and motor preparation and cognitive control.

sive stimuli,^{18,19} activates fear responses,¹⁸ and engages emotional memory²⁰. The insula detects and responds to emotional, interoceptive, and autonomic responses,²¹ and may represent internal feelings of anticipatory anxiety⁹, and amygdala-insula structural connections²² may play an important role in anticipatory processing. During anticipation of aversive stimuli, emotion regulation regions have a dual role – initiating behavioral and cognitive responses and regulating activation in emotion reactivity regions. The dACC integrates cognitive, motor, and affective processes, and plans future behavioral or emotional responses.^{21,23} During anticipation, the dACC may be particularly important in integrating emotional responses and cognitive processes.²¹ The dlPFC also engages emotional working memory²⁴ and exerts cognitive control over emotional responses^{25,26}. The dACC and dlPFC are reciprocally connected,^{27,28} and dACC projections to the amygdala^{29–31} and insula^{32,33} may regulate emotion reactivity.

During anticipation of aversive events, cognitive control processes may regulate emotional responses. Cognitive control processes may be implicit (engaged spontaneously) or explicit (instructed); common cognitive control processes include reappraisal, reality checking, and even the use of a placebo, in studies examining response to pain. Cognitive control processes commonly activate the prefrontal cortex, including the dACC and dlPFC^{34,35} and suppress activation in emotion reactivity regions, including the amygdala and insula³⁶. More frequent use of cognitive reappraisal strategies is related to larger dACC volume³⁷. Placebo analgesics are associated with increased activation in the dlPFC and reduced activation in the insula; the magnitude of the placebo effect correlates with the degree of change of neural response in the dlPFC and insula.³⁶ Individuals who use reappraisal more often during daily life have less amygdala activity during anticipation of aversive stimuli,²⁵ suggesting that individuals who are more practiced at reappraisal may more effectively suppress emotion reactivity responses.

es. Importantly, during anticipation of negative stimuli, functional connectivity between dACC and left amygdala is increased,^{28,30} suggesting that anticipation is a key time for emotion regulation and preparation. Cognitive control strategies may regulate emotional responses³⁸ and may be adaptive during anticipation of aversive stimuli.

Anticipation of Aversive Events in Anxiety Disorders

In patients with anxiety disorders, anticipation can be maladaptive. Patients often have heightened feelings of anticipation, which can lead to fear, helplessness, and feelings of uncontrollable future threat.³⁹ Heightened anticipation of aversive stimuli may result in symptoms of excessive worry and avoidance of otherwise safe situations. Greater anticipatory anxiety, and not severity or frequency of symptoms,⁴⁰ is related to more avoidance behavior, one of the most detrimental features of anxiety disorders⁴¹. Patients with anxiety disorders may also display inappropriate anticipation, resulting in anticipation of both phobic stimuli and of relatively minor events^{24,42,43}. Patients with anxiety disorders have increased attention bias to threat⁴⁴ and are more likely to interpret neutral events as negative⁴⁵, both of which may result in increased anticipation. Anxiety disorders are also characterized by ineffective emotion regulation,⁴⁶ which may result in withdrawal or avoidance behaviors, rather than the more adaptive anticipation - cognitive control and approach behaviors. Differences in anticipatory processing are likely related to two mechanisms – increased threat detection and less effective emotion regulation.

In support of these hypotheses, patients with anxiety disorders also have increased amygdala^{11,24,43} and insula^{42,43,47,48} activation during anticipation of aversive stimuli. Most studies have also found less activation of emotion regulation regions during anticipation, including the dorsal anterior cingulate cortex (dACC)⁴³ and dorsolateral prefrontal cortex (dlPFC)^{43,47,48} activation; however, results are not

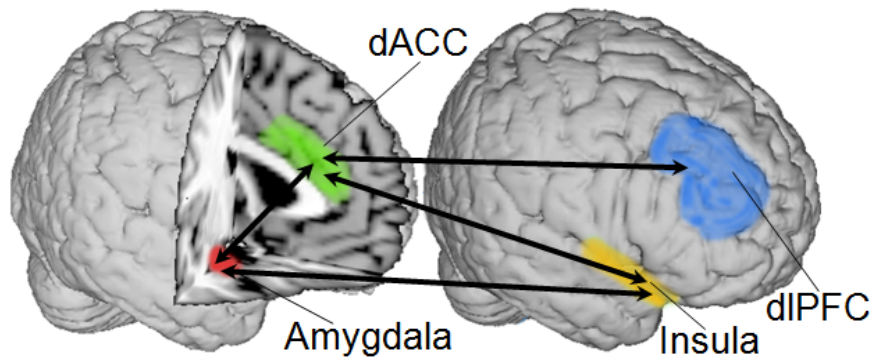


Figure 2: *Neural network of anticipation.* Regions of the brain activated during anticipation of aversive stimuli include the amygdala, insula, dorsal anterior cingulate cortex (dACC), and dorsolateral prefrontal cortex (dlPFC). Connectivity between emotion reactivity regions (amygdala and insula) and emotion regulation regions (dlPFC and dACC) has been observed.

consistent, and a few studies have found increased dACC⁴² and dlPFC²⁴ activation during anticipation of aversive stimuli. Prefrontal cortex regions that are close in anatomic position, may have distinct functions and engage separate processes,⁴⁹ and our understanding of the function of prefrontal cortex regions is evolving over time (for example, see ⁵⁰ and ⁵¹). Further study is needed to determine the precise location and function of changes in prefrontal cortex activity in anxiety disorders. Unlike healthy controls,²⁵ when patients with social phobia engage cognitive control mechanisms during anticipation of aversive stimuli, they do not show increased activation of prefrontal cortex regulation regions⁵².

Patients with anxiety disorders who are better able to engage emotion regulation regions during anticipation of aversive stimuli may have higher functioning, fewer symptoms, and better response to treatment. For example, greater dlPFC activation during anticipation of negative images is associated with fewer PTSD symptoms and better executive functioning.⁴⁷ In patients with generalized anxiety disorder, greater dACC activation during anticipation, was associated with greater reduction in anxiety and depression symptoms following treatment.¹¹ In summary, in patients with anxiety, greater activity of emotion regulation regions during anticipation of aversive events is associated with higher functioning and better treatment response.

Individuals with high trait anxiety are at high risk for developing anxiety disorders, and have increased attention bias to threat⁵³. High trait anxious individuals show similar patterns of anticipation of aversive stimuli, including increased activation of the amygdala,⁵⁴ insula,^{14,48} and dlPFC¹⁴. Degree of trait anxiety was positively associated with activation in the amygdala and insula during anticipation of aversive stimuli,⁹ and high trait anxiety is associated with increased connectivity between the insula and emo-

tion regulation regions, including the dlPFC and dACC.¹⁴ In individuals with high trait anxiety, greater activation of emotion regulation regions and greater coupling between emotion regulation regions and emotion reactivity regions may compensate for heightened emotion reactivity in anxiety. Greater activation of emotion regulation regions during anticipation may protect high trait anxious people from engaging in withdraw or avoidance behavior, and may “protect” against development of anxiety disorders; however, this has not been explicitly tested.

Focus on Anticipation of Aversive Social Stimuli in Social Phobia

Social phobia is defined as the fear or avoidance of one or more performance or social evaluative situations, especially those in which an individual is exposed to social evaluation or scrutiny by others. Symptoms of social phobia are often most prominent during the anticipation of social situations and is accompanied by heightened physiological arousal,^{24,55} anticipatory anxiety,^{24,55} and negative self-beliefs^{56,57}. Understanding the neurobiology of anticipation of social situations has the potential to advance our understanding of the pathophysiology of social phobia.

Anticipation of aversive social stimuli in social phobia has typically been measured using a speech anticipation task, which reflects some of the common triggers of social phobia – fear of social evaluation. Subjects are given several minutes to prepare a speech on a given topic, and then they deliver the speech to a group of experimenters or in front of a video camera. Anticipation of public speaking increases state anxiety,^{24,55} negative affect,⁵⁵ and physiological arousal^{24,55}. During speech anticipation, degree of social anxiety is correlated with increased negative self-beliefs,⁵⁶ self-reported anticipatory anxiety,^{56,57} and physiologic arousal^{56,57}. During anticipation of giving a speech, activa-

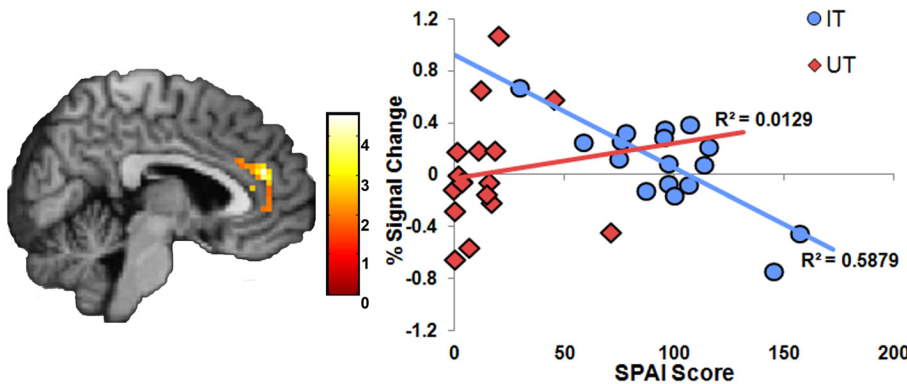


Figure 3: Increased activation in the rostral anterior cingulate in subjects with fewer social anxiety symptoms. During anticipation of fear faces, greater anterior cingulate activation in the inhibited temperament group (IT) was associated with fewer social anxiety symptoms ($R^2 = 0.59$), but not in the uninhibited temperament group, (UT; $R^2 = 0.01$).

tion is increased in regions associated with anticipation of aversive stimuli, including in the amygdala^{24,43} and insula⁴³, and decreased in the dACC and dlPFC⁴³, in patients with social phobia, compared with controls. While studies of anticipation of “protect” against development of anxiety disorders; however, this has not been explicitly tested.

Focus on Anticipation of Aversive Social Stimuli in Social Phobia

Social phobia is defined as the fear or avoidance of one or more performance or social evaluative situations, especially those in which an individual is exposed to social evaluation or scrutiny by others. Symptoms of social phobia are often most prominent during the anticipation of social situations and is accompanied by heightened physiological arousal,^{24,55} anticipatory anxiety,^{24,55} and negative self-beliefs^{56,57}. Understanding the neurobiology of anticipation of social situations has the potential to advance our understanding of the pathophysiology of social phobia.

Anticipation of aversive social stimuli in social phobia has typically been measured using a speech anticipation task, which reflects some of the common triggers of social phobia – fear of social evaluation. Subjects are given several minutes to prepare a speech on a given topic, and then they deliver the speech to a group of experimenters or in front of a video camera. Anticipation of public speaking increases state anxiety,^{24,55} negative affect,⁵⁵ and physiological arousal^{24,55}. During speech anticipation, degree of social anxiety is correlated with increased negative self-beliefs,⁵⁶ self-reported anticipatory anxiety,^{56,57} and physiologic arousal^{56,57}. During anticipation of giving a speech, activation is increased in regions associated with anticipation of aversive stimuli, including in the amygdala^{24,43} and insula⁴³, and decreased in the dACC and dlPFC⁴³, in patients with social phobia, compared with controls. While studies

of anticipation of public speaking show similar results to studies of aversive anticipation in other anxiety disorders, public speaking tasks do not target symptoms of the most disabling subtype of social phobia.

Social phobia has two subtypes: generalized and non-generalized. Generalized social phobia includes multiple social fears; non-generalized social anxiety disorder is confined to a single social fear (usually public speaking), is less disabling, and is rarely brought to medical attention⁵⁸. Generalized social phobia is highly impairing and can result in fear of interacting with other people,⁵⁸ dropping out of school,⁵⁸ losing a job,⁵⁸ and psychiatric comorbidities, including other anxiety disorders and depression⁵⁹. While studying the anticipation of public speaking is important in understanding non-generalized social anxiety disorder, giving talks in front of large groups of people can often be avoided. In generalized social anxiety disorder, encounters with unfamiliar individuals on a day-to-day basis cause impairment and anxiety, and cannot be avoided. Therefore, studying differences in brain activation to more common social stimuli, such as novel faces, is important in understanding the neural basis of generalized social phobia and to prevent and treat this disorder. One way to understand the pathophysiology of generalized social phobia might be to examine brain function during the anticipation of mildly aversive social stimuli.

One study by our lab (Clauss & Blackford, in preparation) has examined the neural correlates of anticipation of fear faces. To study social anxiety disorder, we study a high-risk group, individuals with an inhibited temperament. Inhibited temperament is associated with a 7-fold increased odds of developing social phobia.⁶⁰ In individuals with an inhibited temperament, anticipation of fear faces is associated with greater activation in the dlPFC and dACC, relative to those with an uninhibited temperament. In the inhibited

temperament group, dACC activation was negatively correlated with social phobia symptoms, as measured by the Social Phobia and Anxiety Inventory⁶¹ (see **Figure 3**). The uninhibited group showed no significant change in activation from baseline during anticipation of faces, suggesting that an upcoming mild social stimulus evokes anticipatory activity in the inhibited temperament group only; additionally, there was no relationship between dACC activation and symptoms in the uninhibited temperament group (see **Figure 3**). These results suggest that anticipation of a mildly aversive social stimulus may be a unique probe for social anxiety symptoms.

Implications for Treatment

Anticipation of aversive stimuli is a key process in anxiety disorders and should be targeted in treatment of the disorders.⁴ Neural responses during anticipation of aversive stimuli have been shown to be sensitive to treatment effects. Anxiolytic medications, including selective serotonin reuptake inhibitors and pregabalin, decrease insula and amygdala activity during anticipation.^{62,63} Additionally, non-pharmacologic treatments are effective in modulating activity of emotion reactivity and regulation regions. In patients with social anxiety disorder⁵², engaging in emotion regulation strategies during anticipation reduced insula and amygdala activity and increased ACC activity during anticipation and perception of aversive stimuli; in healthy controls, emotion regulation strategies also increased activity in the dlPFC and individual differences in emotion regulation were negatively correlated with amygdala activity during anticipation²⁵. Greater pre-treatment anterior cingulate activation during anticipation in patients with generalized anxiety disorder was associated with better treatment response, suggesting that individuals who engage emotion regulation areas more at baseline may be more responsive to treatment.¹¹ Therapies that engage emotion regulation regions during aversive anticipation may be effective treatments for anxiety disorders.

Summary

Anticipation is a key psychological process and is highly adaptive by allowing individuals to avoid or modify upcoming aversive events. A network of brain regions is activated during anticipation of aversive stimuli, including the amygdala, insula, dorsal anterior cingulate cortex, and dorsolateral prefrontal cortex. In individuals with anxiety disorders, this network is disrupted; typically, activation

of emotion reactivity regions is higher and activation of emotion regulation regions is lower. Anticipation of public speaking has been studied extensively in social phobia, but anticipation of more mild aversive social stimuli, such as single fear faces, has only been studied in a high-risk group. Considering the disability associated with generalized social phobia, which includes fear of daily social interactions, anticipation of more mild aversive social stimuli should be investigated and targeted for treatment. Enhanced activity of emotion regulation regions is associated with compensatory activity in high-risk, but high-functioning individuals, better outcomes, and better treatment response, suggesting that emotion regulation regions may be an important target for treatment of anxiety disorders.

References

1. Herwig U, Abler B, Walter H and Erk S (2007). Expecting unpleasant stimuli-An fMRI study. *Psychiatry Research: Neuroimaging*. 154 (1): 1–12.
2. Onoda K, Okamoto Y, Toki S, Ueda K, Shishida K, Kinoshita A, Yoshimura S, Yamashita H and Yamawaki S (2008). Anterior cingulate cortex modulates preparatory activation during certain anticipation of negative picture. *Neuropsychologia*. 46 (1): 102–110.
3. Mackiewicz KL, Sarinopoulos I, Cleven KL and Nitschke JB (2006). The effect of anticipation and the specificity of sex differences for amygdala and hippocampus function in emotional memory. *Proceedings of the National Academy of Sciences of the United States of America*. 103 (38): 14200–14205.
4. Nitschke JB, Sarinopoulos I, Mackiewicz KL, Schaefer HS and Davidson RJ (2006). Functional neuroanatomy of aversion and its anticipation. *NeuroImage*. 29 (1): 106–116.
5. Nitschke JB, Larson CL, Smoller MJ, Navin SD, Pederson AJ, Ruffalo D, Mackiewicz KL, Gray SM, Victor E and Davidson RJ (2002). Startle potentiation in aversive anticipation: evidence for state but not trait effects. *Psychophysiology*. 39 (2): 254–258.
6. Grillon C, Baas JP, Lissek S, Smith K and Milstein J (2004). Anxious responses to predictable and unpredictable aversive events. *Behavioral Neuroscience*. 118 (5): 916.
7. Ueda K, Okamoto Y, Okada G, Yamashita H, Hori T and Yamawaki S (2003). Brain activity during expectancy of emotional stimuli: an fMRI study. *Neuroreport*. 14 (1): 51.
8. **Phelps EA, O'Connor KJ, Gatenby JC, Gore JC, Grillon C, Davis M and others (2001). Activation of the left amygdala to a cognitive representation of fear. *Nature Neuroscience*. 4 (4): 437–441.**
9. Carlson JM, Greenberg T, Rubin D and Mujica-Parodi LR (2011). Feeling anxious: anticipatory amygdalo-insular response predicts the feeling of anxious anticipation. *Social, Cognitive, and Affective Neuroscience*. 6 (1): 74–81.

10. Somerville LH, Whalen PJ and Kelley WM (2010). Human bed nucleus of the stria terminalis indexes hypervigilant threat monitoring. *Biological Psychiatry*. 68 (5): 416–424.
11. **Nitschke JB, Sarinopoulos I, Oathes DJ, Johnstone T, Whalen PJ, Davidson RJ and Kalin NH (2009). Anticipatory activation in the amygdala and anterior cingulate in generalized anxiety disorder and prediction of treatment response. *American Journal of Psychiatry*. 166 (3): 302.**
12. Chua P, Krams M, Toni I, Passingham R and Dolan R (1999). A functional anatomy of anticipatory anxiety. *NeuroImage*. 9 (6): 563–571.
13. Herwig U, Kaffenberger T, Baumgartner T and Jancke L (2007). Neural correlates of a ‘pessimistic’ attitude when anticipating events of unknown emotional valence. *NeuroImage*. 34 (2): 848–858.
14. Simmons AN, Stein MB, Strigo IA, Arce E, Hitchcock C and Paulus MP (2011). Anxiety positive subjects show altered processing in the anterior insula during anticipation of negative stimuli. *Human Brain Mapping*. 32 (11): 1836–1846.
15. Drabant EM, Kuo JR, Ramel W, Blechert J, Edge MD, Cooper JR, Goldin PR, Hariri AR and Gross JJ (2010). Experiential, autonomic, and neural responses during threat anticipation vary as a function of threat intensity and neuroticism. *NeuroImage*. 55 (1): 401–10.
16. Holtz K, Pané-Farré CA, Wendt J, Lotze M and Hamm AO (2012). Brain activation during anticipation of interoceptive threat. *NeuroImage*. 61 (4): 857–65.
17. **Simmons A, Matthews SC, Stein MB and Paulus MP (2004). Anticipation of emotionally aversive visual stimuli activates right insula. *Neuroreport*. 15 (14): 2261–65.**
18. Davis M (1992). The role of the amygdala in fear and anxiety. *Annual Review of Neuroscience*. 15 (1): 353–375.
19. Davis M and Whalen PJ (2001). The amygdala: vigilance and emotion. *Molecular Psychiatry*. 6 (1): 13–34.
20. Phelps EA and Anderson AK (1997). Emotional memory: what does the amygdala do? *Current Biology*. 7 (5): R311–R314.
21. Critchley HD (2005). Neural mechanisms of autonomic, affective, and cognitive integration. *The Journal of Comparative Neurology*. 493 (1): 154–166.
22. Mufson EJ, Mesulam MM and Pandya DN (1981). Insular interconnections with the amygdala in the rhesus monkey. *Neuroscience*. 6 (7): 1231–1248.
23. Phillips ML, Drevets WC, Rauch SL and Lane R (2003). Neurobiology of emotion perception I: The neural basis of normal emotion perception. *Biological Psychiatry*. 54 (5): 504–514.
24. **Tillfors M, Furmark T, Marteinsdottir I and Fredrikson M (2002). Cerebral blood flow during anticipation of public speaking in social phobia: a PET study. *Biological Psychiatry*. 52 (11): 1113–1119.**
25. Herwig U, Baumgartner T, Kaffenberger T, Brühl A, Kottlow M, Schreiter-Gasser U, Ablner B, Jäncke L and Rufer M (2007). Modulation of anticipatory emotion and perception processing by cognitive control. *NeuroImage*. 37 (2): 652–662.
26. Wager TD, Jonides J and Reading S (2004). Neuroimaging studies of shifting attention: a meta-analysis. *NeuroImage*. 22 (4): 1679–1693.
27. Petrides M and Pandya DN (1999). Dorsolateral prefrontal cortex: comparative cytoarchitectonic analysis in the human and macaque brain and corticocortical connection patterns. *European Journal of Neuroscience*. 11 1011–1036.
28. Selemon LD and Goldman-Rakic PS (1988). Common cortical and subcortical targets of the dorsolateral prefrontal and posterior parietal cortices in the rhesus monkey: evidence for a distributed neural network subserving spatially guided behavior. *The Journal of Neuroscience*. 8 (11): 4049.
29. Bracht T, Tüscher O, Schnell S, Kreher B, Rüscher N, Glauche V, Lieb K, Ebert D, Il’yasov KA, Hennig J, Weiller C, Van Elst LT and Saur D (2009). Extraction of prefronto-amygdalar pathways by combining probability maps. *Psychiatry Research: Neuroimaging*. 174 (3): 217–222.
30. Ghashghaie HT, Hilgetag CC and Barbas H (2007). Sequence of information processing for emotions based on the anatomic dialogue between prefrontal cortex and amygdala. *NeuroImage*. 34 (3): 905–923.
31. Ray RD and Zald DH (2012). Anatomical insights into the interaction of emotion and cognition in the prefrontal cortex. *Neuroscience & Biobehavioral Reviews*. 36 (1): 479–501.
32. Mesulam M and Mufson EJ (1982). Insula of the old world monkey. III: Efferent cortical output and comments on function. *The Journal of Comparative Neurology*. 212 (1): 38–52.
33. Mufson EJ, Mesulam M and others (1982). Insula of the old world monkey. II: Afferent cortical input and comments on the claustrum. *The Journal of Comparative Neurology*. 212 (1): 23–37.
34. Goldin PR, McRae K, Ramel W and Gross JJ (2008). The neural bases of emotion regulation: reappraisal and suppression of negative emotion. *Biological Psychiatry*. 63 (6): 577–586.
35. Ochsner KN, Ray RD, Cooper JC, Robertson ER, Chopra S, Gabrieli JD, and Gross JJ (2004). For better or for worse: neural systems supporting the cognitive down-and up-regulation of negative emotion. *NeuroImage*. 23 (2): 483–499.
36. Wager TD, Rilling JK, Smith EE, Sokolik A, Casey KL, Davidson RJ, Kosslyn SM, Rose RM and Cohen JD (2004). Placebo-induced changes in FMRI in the anticipation and experience of pain. *Science*. 303 (5661): 1162–1167.
37. Giuliani NR, Drabant EM and Gross JJ (2011). Anterior cingulate cortex volume and emotion regulation: is bigger better? *Biological Psychology*. 86 (3): 379–382.
38. Denny BT, Ochsner KN, Weber J and Wager TD (In press). Anticipatory brain activity predicts the success or failure of subsequent emotion regulation. *Social Cognitive and Affective Neuroscience*. doi:10.1093/scan/nss148
39. Barlow DH, Chorpita BF and Turovsky J Fear, panic, anxiety, and disorders of emotion. *Perspectives on Anxiety, Panic, and Fear*. 43 251–328.
40. Margraf J, Taylor CB, Ehlers A and Roth WT Panic attacks in the natural environment. *The Journal of Nervous and Mental Disease*. 175 (9): 558–565.
41. Adler CM, Craske MG, Kirshenbaum S and Barlow DH (1989). ‘Fear of panic’: An investigation of its role in panic occurrence,

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- phobic avoidance, and treatment outcome. *Behaviour Research and Therapy*. 27 (4): 391–396.
42. Straube T, Mentzel HJ and Miltner WH. (2007). Waiting for spiders: brain activation during anticipatory anxiety in spider phobics. *NeuroImage*. 37 (4): 1427–1436.
43. Lorberbaum JP, Kose S, Johnson MR, Arana GW, Sullivan LK, Hamner MB, Ballenger JC, Lydiard RB, Brodrick PS, Bohning DE and others (2004). Neural correlates of speech anticipatory anxiety in generalized social phobia. *Neuroreport*. 15 (18): 2701.
44. Bradley BP, Mogg K, White J, Groom C and Bono J (1999). Attentional bias for emotional faces in generalized anxiety disorder. *British Journal of Clinical Psychology*. 38 (3): 267–278.
45. Yoon KL and Zinbarg RE (2008). Interpreting neutral faces as threatening is a default mode for socially anxious individuals. *Journal of Abnormal Psychology*. 117 (3): 680.
46. Rodebaugh T and Heimberg R (2008). Emotion regulation and the anxiety disorders: adopting a self-regulation perspective. *Emotion Regulation*. 140–149.
47. Aupperle RL, Allard CB, Grimes EM, Simmons AN, Flagan T, Behrooznia M, Cissell SH, Twamley EW, Thorp SR, Norman SB, Paulus MP and Stein MB (2012). Dorsolateral prefrontal cortex activation during emotional anticipation and neuropsychological performance in posttraumatic stress disorder. *Archives of General Psychiatry*. 69 (4): 360–371.
48. Simmons A, Strigo I, Matthews SC, Paulus MP and Stein MB (2006). Anticipation of aversive visual stimuli is associated with increased insula activation in anxiety-prone subjects. *Biological Psychiatry*. 60 (4): 402–409.
49. Myers-Schulz B and Koenigs M (2011). Functional anatomy of ventromedial prefrontal cortex: implications for mood and anxiety disorders. *Molecular Psychiatry*. 17 (2): 132–141.
50. Bush G, Luu P, Posner MI and others (2000). Cognitive and emotional influences in anterior cingulate cortex. *Trends in Cognitive Sciences*. 4 (6): 215–222.
51. Etkin A, Egner T and Kalisch R (2011). Emotional processing in anterior cingulate and medial prefrontal cortex. *Trends in Cognitive Sciences*. 15 (2): 85–93.
52. Bruhl AB, Herwig U, Delsignore A, Jancke L and Rufer M (In press). General emotion processing in social anxiety disorder: neural issues of cognitive control. *Psychiatric Research: Neuroimaging*.
53. Mogg K, Bradley BP and Hallowell N (1994). Attentional bias to threat: roles of trait anxiety, stressful events, and awareness. *The Quarterly Journal of Experimental Psychology Section A: Human Experimental Psychology*. 47 (4): 841–864.
54. Indovina I, Robbins TW, Núñez-Elizalde AO, Dunn BD and Bishop SJ (2011). Fear-conditioning mechanisms associated with trait vulnerability to anxiety in humans. *Neuron*. 69 (3): 563–571.
55. Davidson RJ, Marshall JR, Tomarken AJ and Henriques JB (2000). While a phobic waits: Regional brain electrical and autonomic activity in social phobics during anticipation of public speaking. *Biological Psychiatry*. 47 (2): 85–95.
56. Schulz SM, Alpers GW and Hofmann SG (2008). Negative self-focused cognitions mediate the effect of trait social anxiety on state anxiety. *Behaviour Research and Therapy*. 46 (4): 438–449.
57. Cornwell BR, Johnson L, Berardi L and Grillon C (2006). Anticipation of public speaking in virtual reality reveals a relationship between trait social anxiety and startle reactivity. *Biological Psychiatry*. 59 (7): 664–666.
58. Stein MB (1996). How shy is too shy? *The Lancet*. 347 (9009): 1131–1132.
59. Kessler RC, Stein MB and Berglund P (1998). Social phobia subtypes in the National Comorbidity Survey. *American Journal of Psychiatry*. 155 (5): 613–619.
60. Clauss JA and Blackford JU (2012). Behavioral inhibition and risk for developing social anxiety disorder: a meta-analytic study. *Journal of the American Academy of Child & Adolescent Psychiatry*. 51 (10): 1066–1075.
61. Turner SM, Beidel DC, Dancu CV and Stanley MA (1989). An empirically derived inventory to measure social fears and anxiety: The Social Phobia and Anxiety Inventory. *Psychological Assessment: A Journal of Consulting and Clinical Psychology*. 1 (1): 35.
62. Simmons AN, Arce E, Lovero KL, Stein MB and Paulus MP (2009). Subchronic SSRI administration reduces insula response during affective anticipation in healthy volunteers. *International Journal of Neuropsychopharmacology*. 12 (8): 1009–20.
63. Aupperle RL, Ravindran L, Tankersley D, Flagan T, Stein NR, Simmons AN, Stein MB and Paulus MP (2011). Pregabalin influences insula and amygdala activation during anticipation of emotional images. *Neuropsychopharmacology*. 36 (7): 1466–1477.

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The $\alpha 1$ Subunit in Sickness and in Health: Properties of the Most Predominant GABA_A Receptor Subunit and Implications of Its Dysfunction in Human Epilepsy

Elizabeth Deel

The aim of this review is to highlight the role of the $\alpha 1$ subunit of the GABA_A receptor under normal conditions and to examine the consequences of its dysfunction in the context of epilepsy. First, background information relevant to GABA_A receptors is discussed, followed by a summary of the biophysical properties conferred by the $\alpha 1$ subunit and its developmental expression patterns. Next, key findings from $\alpha 1$ subunit knockout mice are reviewed. Lastly, the important role of the $\alpha 1$ subunit in regulating inhibitory tone in the CNS is highlighted by examining consequences of mutations in the $\alpha 1$ subunit implicated in generalized human epilepsy syndromes.

Keywords GABA, GABA_A receptor, heterogeneity, alpha-1 subunit, epilepsy

Introduction

GABA_A receptors (GABA_ARs) are a family of chloride-selective, ligand-gated ion channels that mediate the majority of fast inhibition in the adult central nervous system¹. GABA_ARs belong to a larger superfamily of ligand-gated ion channels called Cys-loop receptors, which also includes nicotinic acetylcholine receptors, glycine receptors, and serotonin type III receptors^{2,3}. The GABA_AR gene family is comprised of at least 19 different subunits which are classified by sequence homology into 8 subtypes. These subunits can assemble in various combinations to produce functional GABA_ARs: $\alpha 1$ -6, $\beta 1$ -3, $\gamma 1$ -3, δ , ϵ , π , θ , and $\rho 1$ -3⁴. Based on the large number of different GABA_AR subunits, there are seemingly myriad different subunit combinations possible, although only a subset of these theoretical combinations has been identified *in vivo*⁵. The cache of possible subunit combinations coupled with differing spatial and temporal expression patterns provides considerable structural and functional heterogeneity to GABA_ARs. Distinct subunit combinations produce distinct receptor isoforms which display highly variable properties throughout development and in adulthood⁶⁻⁹. This topic will be discussed in further detail below.

The vast majority of GABA_ARs exist as a combination of two α subunits, two β subunits, and either a γ or δ ^{5,10} subunit arranged as shown in **Figure 1A-B**. The ϵ , π , and $\rho 1$ -3 subunits are far less common and are generally positioned

in place of the γ or δ subunit, while the θ subunit can assume the position of the β subunit. The morphology of each GABA_AR subunit includes several characteristic features. A large extracellular domain at the N-terminus contains the characteristic disulfide bridge between two cysteine residues that creates the “Cys-loop” for which the receptor family is named. There are four helical transmembrane domains termed M1-M4, with the M2 of each subunit lining the ion pore. Connecting M3 and M4, there is a large intracellular domain, and beyond M4 is a very small extracellular C-terminal domain (**Figure 1C**)^{1,2,11}.

Full activation of GABA_ARs requires the binding of two molecules of the neurotransmitter γ -amino butyric acid (GABA)—one at each α/β subunit interface^{12,13}. Upon activation of the receptor, the channel opens and chloride (Cl⁻) flows down its electrochemical gradient through the pore. In the mature brain, this leads to an influx of Cl⁻, which causes the membrane potential of the cell to hyperpolarize and thus inhibits the generation of action potentials.

As previously mentioned, the wide variety of GABA_AR isoforms supports extensive functional heterogeneity, perhaps best demonstrated by the existence of two distinct forms of GABA_AR-mediated inhibition: tonic and phasic¹⁴. Tonic inhibition is mediated by extrasynaptically localized GABA_ARs largely comprised of a δ subunit with an $\alpha 4$ and/or an $\alpha 6$ subunit¹⁵, though some extrasynaptic GABA_ARs containing $\alpha 5$ and lacking a δ subunit are known to func-

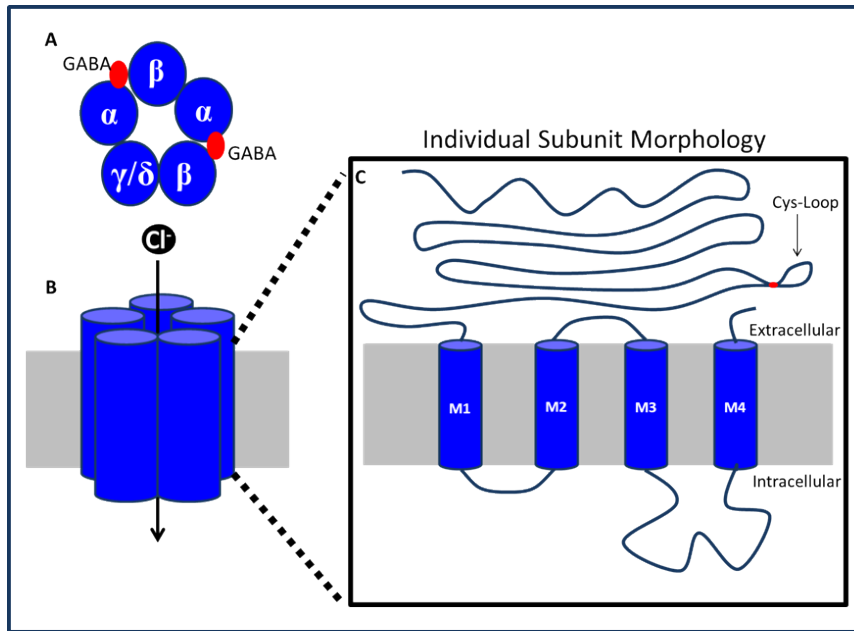


Figure 1: Structure of GABA_A receptors. A: Top down view of typical subunit arrangement with GABA binding site indicated by red ovals. B: Side view of receptor depicting chloride flowing through the pore. C: Morphology of an individual subunit.

tion extrasynaptically as well¹⁶. Extrasynaptic GABA_ARs are continuously activated by low levels (1M) of ambient GABA typically overflowing from the synaptic cleft. The properties of these receptors are conducive to persistent extrasynaptic activation, as they are highly sensitive to GABA, activate relatively slowly, and desensitize minimally¹⁷. The functional role of tonic inhibition seems to be the regulation of neuronal excitability. Conversely, phasic inhibition is mediated by synaptic GABA_ARs, which most often contain a γ subunit with $\alpha 1$, $\alpha 2$, and/or $\alpha 3$ subunits. Unlike their extrasynaptic counterparts, synaptic GABA_ARs are transiently activated by much higher (1mM) concentrations of GABA released from the presynaptic neuron into the synaptic cleft. In contrast to extrasynaptic GABA_ARs, synaptic GABA_ARs mediate tonic inhibition to allow for fast transmission of a fleeting signal and are thus less sensitive to GABA, activate rapidly, and desensitize extensively¹⁸. The short-lived activation of these receptors produces a transient inhibitory post-synaptic current (IPSC) which translates presynaptic GABA release into a post-synaptic signal^{19,20}. The remainder of this review will focus on the most predominant subtype of synaptic receptors: those containing the $\alpha 1$ subunit. The distinct receptor properties and known roles of the $\alpha 1$ subunit, as well as important findings from $\alpha 1$ subunit knockout mice and recently discovered epilepsy-associated mutations in the $\alpha 1$ subunit are discussed below.

Biophysical Properties of the $\alpha 1$ Subunit Relative to Other Synaptic α Subunits

The $\alpha 1$ subunit is the most predominant α subunit in the adult brain and is most often assembled into the $\alpha 1\beta 2\gamma 2$ GABA_AR isoform comprising 60% of all GABA_ARs⁸. Further heterogeneity exists among synaptic receptors. The specific α subtype influences biophysical properties of GABA_ARs including GABA sensitivity and the rates of activation, deactivation, and desensitization²¹⁻²⁵. The primary strategy employed to determine the contribution of each α subunit to a particular property is to express different α subunits with the same β and γ subunit partners in heterologous cells and explore the properties of interest. One such property, GABA sensitivity, is defined by the concentration of GABA that is required to produce a given response. A common measure of GABA sensitivity is the concentration of GABA required to elicit a half-maximal response in a given receptor subtype, known as the EC₅₀. A low EC₅₀ indicates higher sensitivity and vice versa. The activation rate of a particular GABA_AR is the rate at which receptor current increases from 10% to 90% of the maximal or peak current, and the deactivation rate is the rate at which current amplitude decreases after GABA is removed.

The desensitized state of the receptor is a high-affinity state in which GABA is bound but the ion channel is closed. The desensitization rate is the rate at which the response diminishes in continued presence of GABA. Each of the biophysical properties defined above influence the shape and time course of GABA_AR-mediated IPSCs. Thus, the identity of the α subunit within the receptor in large part

GABA Sensitivity	$\alpha 1 > \alpha 2 > \alpha 3$
Activation Rate	$\alpha 2 > \alpha 1 > \alpha 3$
Deactivation Rate	$\alpha 1 > \alpha 2 > \alpha 3$
Desensitization Rate	$\alpha 1 = \alpha 2 > \alpha 3$

Table 1: *Biophysical properties by synaptic α subunits.*

dictates the properties of inhibitory currents. The biophysical properties of $\alpha 1$ containing GABA_ARs relative to other synaptic α subunits are summarized in **Table 1**^{21–24}.

Interestingly, the properties of GABA_AR mediated IPSCs are known to change throughout development^{26,27}—namely the decay kinetics which are heavily influenced by the deactivation and desensitization rates of the GABA_ARs^{28,29}. In fetal and early postnatal development, GABA_AR-mediated IPSCs decay relatively slowly; later in development, the IPSCs decay much more rapidly. The timing of this change coincides with the timing of a well-established developmental change in GABA_AR α subunit expression^{6,30,31}. Early in development, the $\alpha 2$ and $\alpha 3$ subunits predominate, but soon after birth their expression begins to wane while the expression of the $\alpha 1$ subunit steadily increases to become the most abundant α subunit by postnatal day 12 in mice⁶. A comparison of juvenile and mature GABA_AR-mediated IPSCs is shown in **Figure 2**. Given that the identity of the α subunit impacts IPSC properties, it is feasible that the developmental changes in α subunit expression and IPSC decay kinetics are causally linked. Indeed, it has been shown that in mice lacking the $\alpha 1$ subunit juvenile IPSC kinetics persist into adulthood^{32–34}. The functional role of this developmental switch in α subunit expression and the concomitant change in IPSC kinetics is currently not well understood.

Findings From $\alpha 1$ Subunit Knockout Mice

In 2001, transgenic mice lacking the $\alpha 1$ subunit of the GABA_AR exhibited a 50–60% decrease in the total number of GABA_ARs in the brain^{33,35}. Consistent with this finding, the expression of the $\beta 2/3$ and $\gamma 2$ subunits—the most common binding partners of the $\alpha 1$ subunit—is also

decreased in $\alpha 1$ knockout mice^{36,37}. Given that the $\alpha 1$ subunit is the most abundant subunit and its loss results in a loss of the majority of GABA_ARs in the brain, it is very surprising that these animals are viable and lack any obvious phenotypic abnormalities aside from a slight handling-induced tremor. The fact that the mice are overtly normal could suggest that changes occur within the GABA_AR system to compensate for the loss of $\alpha 1$. Indeed, post-transcriptional increases in the expression of the other α subunits have been observed^{36,37}, but the nature and extent of the compensation seems to vary among brain regions and has not been systematically quantified in the entire brain.

One study suggests that neurons upregulate the subunits they normally express rather than expanding their subunit repertoire³⁶, which is consistent with a post-transcriptional mode of upregulation. The consequences of these compensatory changes are not completely understood, but $\alpha 1$ knockout mice fail to develop mature IPSC kinetics^{32–34} and exhibit a lower threshold for pharmacologically induced seizures³⁸. This phenotype could indicate a decrease in inhibitory tone, although it was reported that $\alpha 1$ knockout mice did not experience spontaneous seizures³⁵. However, it is important to note that these conclusions were drawn by visual inspection only rather than EEG analysis. Certain types of seizures, such as absence seizures, result in very subtle alterations in behavior that are difficult to detect even in a human, much less in a mouse. Thus it is possible that these animals did have seizures but did not display any easily detectable seizure behavior.

The $\alpha 1$ Subunit and Epilepsy

Because GABA_ARs are the primary source

Electroencephalogram (EEG):

Method to measure electrical activity of the brain using electrodes on the surface of the scalp.

Missense Mutation:

Non-synonymous point mutation in which the identity of a single nucleotide is changed resulting in a codon that codes for an amino acid that differs from that of the WT protein at that particular location.

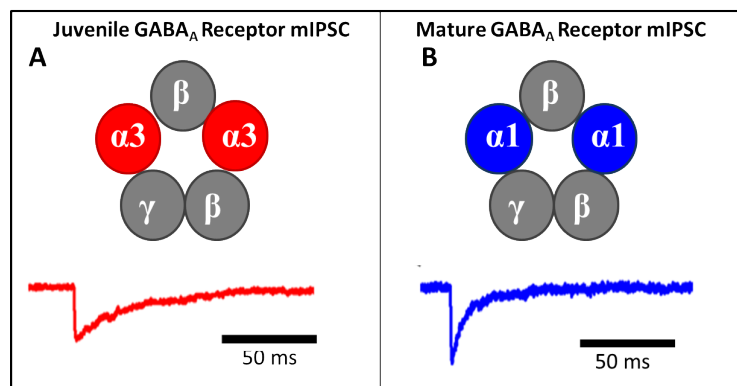


Figure 2: Developmental change in α subunit expression and IPSC kinetics. A: Juvenile $\alpha 3$ containing receptor with much slower IPSC delay kinetics. B: Mature $\alpha 1$ containing receptor with fast IPSC decay kinetics.

Endoplasmic Reticulum Associated Degradation:

A cellular process that marks misfolded proteins within the ER for ubiquitination followed by degradation via the proteasome.

Nonsense Mediated Decay:

Cellular mechanism to detect nonsense mutations (premature stop codons) and prevent translation of truncated proteins by degrading the mutant mRNA.

of inhibition in the central nervous system, it is not surprising that several mutations in various GABA_AR subunits have been identified in patients with idiopathic generalized epilepsy (IGE) syndromes such as childhood absence epilepsy and juvenile myoclonic epilepsy^{39–41}. Epilepsy is a large collection of syndromes diagnosed upon the occurrence of two or more unprovoked seizures. Epilepsy may be classified as IGE if the cause is thought to be genetic and the seizures appear to involve the entire brain simultaneously with no obvious focal origin. Most IGEs are thought to be multigenic, which renders developing animal models fairly difficult. However, the identification of monogenic forms of IGE has permitted the generation of animal models based on human disease-associated mutations. To date, there have been four mutations identified specifically in the $\alpha 1$ subunit in human patients suffering from idiopathic generalized epilepsy (IGE.) Two of these mutations, K353delins18X and D219N⁴², were only recently reported and have not been fully characterized. The other two however, A322D⁴³ and S326fs328X⁴⁴, have been extensively studied.

$\alpha 1$ (K353delins18X) Mutation

The $\alpha 1$ (K353delins18X) mutation was identified in 1 unaffected and 3 affected individuals with IGE exhibiting generalized tonic-clonic seizures. These seizures manifest as a sudden tensing of skeletal muscles followed by rapid contractions and relaxations resulting in characteristic convulsions⁴⁵. This mutation involves the insertion of 25 base pairs into intron

10 which interrupts splicing and causes the retention of intron 10 in the transcript. The inclusion of intron 10 leads to an 18-amino acid insertion into the protein as well as the truncation of the fourth transmembrane domain due to a premature stop codon. Work in heterologous expression systems revealed that the protein is localized to the ER with complete loss of cell surface expression. In agreement with these findings, GABA-mediated currents were absent in these cells⁴². The fate of the mutant protein and the mechanism by which the $\alpha 1$ (K353delins18X) mutation leads to epilepsy is yet to be determined.

$\alpha 1$ (D219N) Missense Mutation

The D219N missense mutation was identified in 4 of 5 affected individuals with IGE in a French-Canadian family exhibiting IGE or febrile seizures(FS)—so aptly named due to their coincidence with fever⁴⁵. Two of the four individuals with FS also reported a single generalized tonic clonic seizure. Studies conducted in heterologous expression systems indicate that surface expression of the mutant subunit is reduced as compared to WT $\alpha 1$ subunit, consistent with observations of decreased GABA-evoked peak current amplitudes. Additionally, $\alpha 1$ (D219N) subunit-containing receptors were shown to desensitize more rapidly than WT $\alpha 1$ subunit-containing receptors⁴². Further studies characterizing the effects of this mutation both *in vitro* and *in vivo* will be required to elucidate the mechanism by which it promotes the development of epilepsy.

$\alpha 1$ (A322D) Missense Mutation

The A322D missense mutation was identified in 8 affected individuals within a large French-Canadian family suffering from a type of IGE called juvenile myoclonic epilepsy (JME). Myoclonic seizures are characterized by sudden, brief, involuntary jerks of the arms or legs⁴⁵. This mutation is autosomal dominant and results in the insertion of a charged aspartate residue in place of a highly conserved alanine within the M3 domain^{39–41,43}. Experiments in heterologous cells indicate that this mutation disrupts the insertion of M3 into the lipid bilayer which results in its retention in the endoplasmic reticulum (ER) and subsequent partial degradation through ER-associated degradation^{39–41,46} (ERAD). However, the mutant subunit is not completely degraded, as total and surface mutant protein is detectable, albeit at significantly lower levels than the WT $\alpha 1$ subunit^{39–41,46}. It has also been postulated that the $\alpha 1$ (A322D) mutation exerts a dominant negative effect by oligomerizing with and trapping WT subunits in the ER which are then subject to ERAD^{39,47}. Consistent with reduced surface expression, the peak GABA-evoked current through receptors containing the $\alpha 1$ (A322D) subunit was reduced 88%⁴⁷. In the mutant receptors, open time of the channel was considerably reduced. Additionally, $\alpha 1$ (A322D) subunit-containing receptors exhibited substantially reduced sensitivity to GABA with a nearly 100-fold increase in the GABA EC₅₀^{39,40}. It would be particularly enlightening to study the effects of this mutation *in vivo* and to that end, our group is in the process of generating an $\alpha 1$ (A322D) knock-in mouse line.

$\alpha 1$ (S326fs328X) Frameshift Mutation

The $\alpha 1$ (S326fs328X) mutation is an autosomal dominant *de novo* mutation identified in a patient with childhood absence epilepsy. In contrast to the previously mentioned seizure types, absence seizures are not associated with any sort of convulsions or motor movements. Rather, they are characterized by sudden brief lapses in consciousness often accompanied by a blank stare⁴⁵. A single base pair deletion leads to a frameshift and premature termination codon (PTC) in the eighth exon which corresponds to the third transmembrane domain of the protein⁴⁴. The PTC has been shown to induce nonsense mediated decay (NMD) of the mutant mRNA, albeit incomplete. Mutant mRNA that escapes NMD is subsequently translated into truncated protein which is retained in the ER and subjected to ERAD^{48,49}. Thus, the $\alpha 1$ (S326fs328X) mutant subunit is not incorporated into the cell membrane and GABA-evoked currents are absent^{44,48}.

Based on these findings, it is thought that the epilepsy phenotype is a result of haploinsufficiency in the WT $\alpha 1$ gene. This raises the intriguing possibility that heterozygous $\alpha 1$ knockout animals could serve as a model for this disease. Indeed, our group recently revealed through EEG analysis that heterozygous $\alpha 1$ knockout mice do in fact experience seizures, though they are absence seizures rather than convulsive seizures⁵⁰. This may explain why it was previously reported that $\alpha 1$ knockout mice did not exhibit an epileptic phenotype as mentioned above. The seizures were greatly attenuated by treatment with ethosuximide (ETX), the most commonly prescribed drug for absence seizures in human patients. These findings represent a novel model of absence epilepsy based on a mutation identified in a human epilepsy patient.

Conclusions

GABA_ARs are a heterogeneous population of receptors and their properties are heavily influenced by their α subunit composition. The $\alpha 1$ subunit is the most predominant subunit in the adult brain and is largely responsible for the maintenance of inhibitory tone in the CNS. As evidenced by the consequences of the dysfunction or loss of the $\alpha 1$ subunit, it also seems to be involved in epilepsy susceptibility. The transgenic $\alpha 1$ (A322D) knock-in and $\alpha 1$ knockout mouse lines that our group focuses on represent the first mouse models of genetic epilepsy based on mutations in the $\alpha 1$ subunit identified in human epilepsy patients. These two mutations were both identified in patients suffering from generalized epilepsy, but their seizure phenotypes were distinct. Based on our preliminary analyses, the epileptic phenotypes of the two mouse lines also differ. By dissecting the similarities and differences in pathogenesis behind these two models, we aim to identify common themes among all generalized epilepsies and also delineate important differences that contribute to distinct disease manifestations. While epilepsy is an exceptionally complicated and heterogeneous condition, the advent of animal models reflecting specific disease-associated mutations in GABA_ARs represents a promising and riveting new avenue for advancing our understanding of the pathogenesis of generalized epilepsy.

References

1. Sieghart, W. Structure, Pharmacology, and Function of GABA_A Receptor Subtypes. *Advances in Pharmacology* **54**, 231-263 (2006).

CANDIDATE REVIEWS

2. Connolly, C. & Wafford, K. The Cys-Loop Superfamily of Ligand-gated Ion Channels: The Impact of Receptor Structure and Function. *Biochem Soc Trans* **32**, 529-534 (2004).
3. Unwin, N. Neurotransmitter action: opening of ligand-gated ion channels. *Cell* **72**, 31-41 (1993).
4. Sieghart, W. & Sperk, G. Subunit composition, distribution and function of GABA(A) receptor subtypes. *Current Topics in Medicinal Chemistry* **2**, 795-816 (2002).
5. McKernan, R.M. & Whiting, P.J. Which GABA A receptor subtypes really occur in the brain? *Trends in Neuroscience* **19**, 139-143 (1996).
6. Laurie, D.J., Wisden, W. & Seeburg, P.H. The Distribution of Thirteen GABA (A) Receptor Subunit mRNAs in the Rat Brain III: Embryonic and Postnatal Development. *The Journal of Neuroscience* **12**, 4151-4172 (1992).
7. Olsen, R.W. & Sieghart, W. GABA A receptors: subtypes provide diversity of function and pharmacology. *Neuropharmacology* **56**, 141-8 (2009).
8. Möhler, H. GABA(A) receptor diversity and pharmacology. *Cell and Tissue Research* **326**, 505-16 (2006).
9. Hevers, W. & Lüddens, H. The diversity of GABAA receptors: Pharmacological and electrophysiological properties of GABAA channel subtypes. *Molecular Neurobiology* **18**, 35-86 (1998).
10. Baumann, S.W., Baur, R. & Sigel, E. Subunit arrangement of gamma-aminobutyric acid type A receptors. *The Journal of Biological Chemistry* **276**, 36275-80 (2001).
11. Macdonald, R.L. & Olsen, R.W. GABAA receptor channels. *Annual Review of Neuroscience* **17**, 569-602 (1994).
12. Baumann, S.W., Baur, R. & Sigel, E. Individual properties of the two functional agonist sites in GABA(A) receptors. *The Journal of Neuroscience* **23**, 11158-66 (2003).
13. Kash, T.L., Jenkins, A., Kelley, J.C., Trudell, J.R. & Harrison, N.L. Coupling of agonist binding to channel gating in the GABA(A) receptor. *Nature* **421**, 272-5 (2003).
14. Mody, I. Distinguishing between GABA(A) receptors responsible for tonic and phasic conductances. *Neurochemical Research* **26**, 907-13 (2001).
15. Sur, C., Farrar, S., Kerby, J., Whiting, P., Atack, J., McKernan, R. Preferential Coassembly of Alpha 4 and Delta Subunits of the GABAA Receptor in Rat Thalamus. *Molecular pharmacology* **56**, 110-115 (1999).
16. Caraiscos, V.B., Elliott, E., You-Ten, K., Cheng, V., Beelli, D., Newell, J., Jackson, M., Lambert, J., Rosahl, T., Wafford, K., MacDonald, J., Orser, B. Tonic inhibition in mouse hippocampal CA1 pyramidal neurons is mediated by alpha5 subunit-containing gamma-aminobutyric acid type A receptors. *Proceedings of the National Academy of Sciences of the United States of America* **101**, 3662-7 (2004).
17. Mchedlishvili, Z. & Kapur, J. High-Affinity, Slowly Desensitizing GABA A Receptors Mediate Tonic Inhibition in Hippocampal Dentate Granule Cells. *Molecular Pharmacology* **69**, 564-575 (2006).
18. Haas, K.F. & Macdonald, R.L. GABA A receptor subunit gamma2 and delta subtypes confer unique kinetic properties on recombinant GABA A receptor currents in mouse fibroblasts. *Journal of Physiology* **514**, 27-45 (1999).
19. Farrant, M. & Nusser, Z. Variations on an inhibitory theme: phasic and tonic activation of GABA(A) receptors. *Nature Reviews Neuroscience* **6**, 215-29 (2005).
20. Nusser, Z., Sieghart, W. & Somogyi, P. Segregation of Different GABA A Receptors to Synaptic and Extrasynaptic Membranes of Cerebellar Granule Cells. *Journal of Neuroscience* **18**, 1693-1703 (1998).
21. Gingrich, K.J., Roberts, W. a & Kass, R.S. Dependence of the GABAA receptor gating kinetics on the alpha-subunit isoform: implications for structure-function relations and synaptic transmission. *The Journal of Physiology* **489**, 529-43 (1995).
22. Eyre, M.D., Renzi, M., Farrant, M. & Nusser, Z. Setting the Time Course of Inhibitory Synaptic Currents by Mixing Multiple GABAA Receptor Alpha Subunit Isoforms. *Journal of Neuroscience* **32**, 5853-5867 (2012).
23. Picton, A.J. & Fisher, J.L. Effect of the alpha subunit subtype on the macroscopic kinetic properties of recombinant GABA(A) receptors. *Brain Research* **1165**, 40-9 (2007).
24. Lavoie, a M., Tingey, J.J., Harrison, N.L., Pritchett, D.B. & Twyman, R.E. Activation and deactivation rates of recombinant GABA(A) receptor channels are dependent on alpha-subunit isoform. *Biophysical Journal* **73**, 2518-26 (1997).
25. Böhme, I., Rabe, H. & Lüddens, H. Four amino acids in the alpha subunits determine the gamma-aminobutyric acid sensitivities of GABAA receptor subtypes. *The Journal of Biological Chemistry* **279**, 35193-200 (2004).
26. Okada, M., Onodera, K., Van Renterghem, C., Sieghart, W. & Takahashi, T. Functional correlation of GABA(A) receptor alpha subunits expression with the properties of IPSCs in the developing thalamus. *The Journal of Neuroscience* **20**, 2202-8 (2000).
27. Dunning, D., Hoover, C., Soltesz, I., Smith, M., Dowd, D., Magueresse, C., Alfonso, J., Khodosevich, K., Martín, Á., Bark, C., Monyer, H. GABA A Receptor Mediated Miniature Postsynaptic Currents and α -Subunit Expression in Developing Cortical Neurons. *The Journal of Neurophysiology* **82**: 3286-3297 (1999).
28. Jones, M.V. & Westbrook, G.L. Desensitized states prolong GABAA channel responses to brief agonist pulses. *Neuron* **15**, 181-91 (1995).
29. Ortinski, P.I., Lu, C., Takagaki, K., Fu, Z. & Vicini, S. Expression of distinct alpha subunits of GABAA receptor regulates inhibitory synaptic strength. *Journal of Neurophysiology* **92**, 1718-27 (2004).
30. Hashimoto, T., Nguyen, Q., Rotaru, D., Keenan, T., Arion, D., Beneyto, M., Gonzalez-Burgos, G., Lewis, D. Protracted developmental trajectories of GABAA receptor alpha1 and alpha2 subunit expression in primate prefrontal cortex. *Biological Psychiatry* **65**, 1015-23 (2009).
31. Fritschy, J.-M., Paysan, J., Enna, A. & Mohler, H. Switch in the Expression of Rat GABAA-Receptor Subtypes During Postnatal Development: An Immunohistochemical Study. *Journal of Neuroscience* **14**, 5302-5324 (1994).
32. Bosman, L.W.J., Heinen, K., Spijker, S. & Brussaard, A.B. Mice lacking the major adult GABAA receptor subtype have normal number of synapses, but retain juvenile IPSC kinetics until adulthood. *Journal of Neurophysiology* **94**, 338-46 (2005).

33. **Vicini, S., Ferguson, C., Prybylowski, K., Kralic, J., Morrow, L., Homanics, G. . GABA(A) receptor alpha1 subunit deletion prevents developmental changes of inhibitory synaptic currents in cerebellar neurons. *The Journal of Neuroscience* 21, 3009-16 (2001).**
This paper represents the generation of the original transgenic GABA α 1 subunit knockout mouse line. We use the heterozygous animals from this line in our lab as a model for studying the pathogenesis of absence epilepsy.
34. Goldstein, P., Elsen, F., Ying, S., Ferguson, C., Homanics, G., Harrison, N. Prolongation of hippocampal miniature inhibitory postsynaptic currents in mice lacking the GABA(A) receptor alpha1 subunit. *Journal of Neurophysiology* 88, 3208-17 (2002).
35. Sur, C., Wafford, K., Reynolds, D., Hadingham, K., Bromidge, F., Macaulay, A., Collinson, N., Meara, G., Howell, O., Newman, R., Myers, J., Atack, J., Dawson, G., McKernan, R., Whiting, P., Rosahl, T. Loss of the major GABA(A) receptor subtype in the brain is not lethal in mice. *The Journal of Neuroscience* 21, 3409-18 (2001).
36. Kralic, J., Sidler, C., Parpan, F., Homanics, G., Morrow, A., Fritschy, J. Compensatory alteration of inhibitory synaptic circuits in cerebellum and thalamus of gamma-aminobutyric acid type A receptor alpha1 subunit knockout mice. *The Journal of Comparative Neurology* 495, 408-21 (2006).
37. Kralic, J., Korpi, E., Buckley, T., Homanics, G., Morrow, A. Molecular and Pharmacological Characterization of GABA A Receptor Alpha 1 Subunit Knockout Mice. *The Journal of Pharmacology and Experimental Therapeutics* 302, 1037-1045 (2002).
38. Kralic, J., O'Buckley, T., Khisti, R., Hodge, C., Homanics, G., Morrow, A. GABA(A) receptor alpha-1 subunit deletion alters receptor subtype assembly, pharmacological and behavioral responses to benzodiazepines and zolpidem. *Neuropharmacology* 43, 685-94 (2002).
39. Macdonald, R.L., Kang, J.Q., Gallagher, M.J. & Feng, H.J. GABA A Receptor Mutations Associated with Generalized Epilepsy. *Advances in Pharmacology* 54, 1497-1506 (2006).
40. Macdonald, R.L., Gallagher, M.J., Feng, H.-J. & Kang, J. GABA(A) receptor epilepsy mutations. *Biochemical Pharmacology* 68, 1497-506 (2004).
41. Macdonald, R.L., Kang, J.Q. & Gallagher, M.J. Mutations in GABAA receptor subunits associated with genetic epilepsies. *The Journal of Physiology* 588, 1861-9 (2010).
42. Lachance-Touchette, P., Brown, P., Meloche, C., Kinirons, P., Lapointe, L., Lacasse, H., Lortie, A., Carmant, L., Bedford, F., Bowie, D., Cossette, P. Novel α 1 and γ 2 GABAA receptor subunit mutations in families with idiopathic generalized epilepsy. *The European Journal of Neuroscience* 34, 237-49 (2011).
43. Cossette, P., Liu, L., Brisebois, K., Dong, H., Lortie, A., Vanasse, M., Saint-Hilaire, J., Carmant, L., Verner, A., Lu, W., Wang, Y., Rouleau, G. Mutation of GABRA1 in an autosomal dominant form of juvenile myoclonic epilepsy. *Nature Genetics* 31, 184-9 (2002).
44. **Maljevic, S., Krampff, K., Cobilanschi, J., Tilgen, N., Beyer, S., Weber, Y., Schlesinger, F., Ursu, D., Melzer, W., Cossette, P., Bufler, J., Lerche, H., Heils, A. et al. A mutation in the GABA(A) receptor alpha(1)-subunit is associated with absence epilepsy. *Annals of Neurology* 59, 983-7 (2006).**
This paper identifies the first mutation in GABA α 1 subunit associated with childhood absence epilepsy. This mutation is the basis for the mouse model our lab uses to study the pathogenesis of absence epilepsy.
45. Commission on Classification and Terminology of the International League Against Epilepsy. Proposal for Revised Classification of Epilepsies and Epileptic Syndromes. *Epilepsia* 30, 389-399 (1989).
46. Gallagher, M.J., Shen, W., Song, L. & Macdonald, R.L. Endoplasmic reticulum retention and associated degradation of a GABAA receptor epilepsy mutation that inserts an aspartate in the M3 transmembrane segment of the alpha1 subunit. *The Journal of Biological Chemistry* 280, 37995-8004 (2005).
47. Ding, L., Feng, H., Macdonald, R., Botzolakis, E., Hu, N., Gallagher, M. GABA(A) receptor alpha1 subunit mutation A322D associated with autosomal dominant juvenile myoclonic epilepsy reduces the expression and alters the composition of wild type GABA(A) receptors. *The Journal of Biological Chemistry* 285, 26390-405 (2010).
48. **Kang, J.-Q., Shen, W. & Macdonald, R.L. Two molecular pathways (NMD and ERAD) contribute to a genetic epilepsy associated with the GABA(A) receptor GABRA1 PTC mutation, 975delC, S326fs328X. *The Journal of Neuroscience* 29, 2833-44 (2009).**
This paper establishes that the mutant α 1 subunit is degraded, indicating that α 1 haploinsufficiency may be associated with the childhood absence epilepsy phenotype. These findings justify heterozygous α 1 knockout mice as a putative model for the α 1(S326fs328X) mutation identified in human epilepsy patients.
49. Macdonald, R.L., Kang, J.-Q. & Gallagher, M.J. Mutations in GABAA receptor subunits associated with genetic epilepsies. *The Journal of Physiology* 588, 1861-9 (2010).
50. **Arain, F.M., Boyd, K.L., & Gallagher, M.J. Decreased viability and absence-like epilepsy in mice lacking or deficient in the GABAA Receptor α 1 Subunit. *Epilepsia* (2012, In Press).**
This paper validates heterozygous α 1 knockout mice as a model for absence epilepsy as it shows that the mice have absence-like seizures that remit with ethosuximide, the most commonly prescribed drug for the treatment of absence seizures.

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Amygdala Developmental Consequences of Childhood Maltreatment

E. Kale Edmiston

Childhood maltreatment is a significant social problem associated with increased risk for depression and anxiety disorders. Despite this, many adults who experience childhood maltreatment do not develop psychiatric disorders and the neurobiological correlates for risk and resilience following childhood maltreatment are not well understood. Even in the absence of psychiatric diagnosis, childhood maltreatment can be associated with sub-clinical alterations in threat detection, a likely adaptive response to the early environment. Enhanced threat sensitivity following childhood maltreatment may be mediated by alterations in the developmental trajectory of the amygdala, a region often associated with threat detection. Functional MRI studies of populations with childhood maltreatment histories suggest heightened amygdala activation to aversive stimuli. Likewise, rodent models of chronic early stress exposure suggest amygdala effects, including increased dendritic arborization and decreased expression of the serotonin transporter (5-HTT). Genetic studies have also implicated 5-HTT, reporting an increased risk for depression following childhood maltreatment in carriers of the low-expressing of the 5-HTTLPR. Neuroimaging genetic studies have found increased amygdala BOLD signal in response to emotional or threatening stimuli in the low-expressing allele carriers exposed to maltreatment. Taken together, this literature suggests that the amygdala has heightened reactivity to threatening stimuli in people exposed to childhood maltreatment, and that these effects may be mediated by genetic variations in the serotonin system. Importantly, enhanced risk for depression is an end-point outcome well downstream of a host of genetic and environmental factors that interact dynamically throughout development. Future studies should address possible mediating factors by assessing for onset and duration of maltreatment, symptom severity, as well as gene x environment effects.

Keywords: *fMRI, serotonin transporter, childhood maltreatment, amygdala, anxiety, depression*

Introduction

Childhood maltreatment is a significant social problem affecting 3.7 million children annually in the United States alone¹. Childhood maltreatment, which can include physical, emotional, and sexual abuse, as well as physical and emotional neglect, is associated with a host of negative outcomes, including increased risk for psychiatric disorders such as major depressive disorder and anxiety disorders². Even in the absence of psychiatric symptoms that meet criteria for diagnosis, populations exposed to maltreatment can experience sub-clinical alterations in emotional processing, such as increased threat sensitivity and decreased emotional regulation³⁻⁵. Emotional processing changes are likely adaptive responses to early environmental exposure to threat, whereby threatening experiences have become generalized to the broader social environment as a way to avoid additional harm⁶. Observed changes in threat sensi-

tivity and emotional regulation are likely mediated by the effects of early life stress on the developmental trajectory of the amygdala, a region implicated in threat detection and fear conditioning⁷, as well as anxiety and mood disorders⁸. Despite increased risk for psychiatric disorders and behavioral dysfunction in populations exposed to childhood maltreatment, a significant proportion do not develop negative outcomes later in life. What, then, are the neurobiological correlates of enhanced risk for poor outcome following childhood maltreatment? This review will examine potential gene x environment developmental effects of childhood maltreatment on the structure and function of the amygdala, as well as how altered amygdala development may mediate increased risk for psychiatric disorder.

Amygdala Alterations in Magnetic Resonance Imaging Findings of Childhood Maltreatment

Enhanced sensitivity to potentially threatening emotional

Reference	Imaging Paradigm	Maltreated/ Total	Age Range	Psychiatric Diagnoses	Maltreatment Type	Maltreatment Assessment	Amygdala Findings
Garret et al., 2012	emotional face passive viewing	30/56	10-16	None	PN, EN, PA, EA, SA	Self report	BOLD increase, neutral and angry faces
Grant et al., 2010	emotional face flanker task	10/36	18-55	MDD	PN, EN, PA, EA, SA	Self report	BOLD increase, negative affect faces
Maheu et al., 2010	emotional face directed viewing	11/30	9-18	Anxiety d/o, n=2	PN, EN	Social services documentation	BOLD increase, angry and fear faces
McCrory et al., 2011	emotional face passive viewing	20/43	10-13	None	PN, PA, EA, SA	Social services documentation	BOLD increase, angry faces
Protopopescu et al., 2005	traumatic word passive viewing	9/14	20-55	PTSD	PA, SA	Self report	BOLD increase, traumatic words
Tottenham et al., 2011	emotional face go/no-go task	22/44	7-13	Anxiety d/o, n=2; ADHD, n=5; ODD, n=1	Previous institutionalization	Social services documentation	BOLD increase, fear and distractor faces
Van Harmelen et al., 2012	emotional face passive viewing	60/135	33-41	MDD or Anxiety d/o	EN, EA	Self report	BOLD increase for all faces
Bremner et al., 1997	sMRI, hand tracing	17/34	25-52	PTSD	PA, SA	Clinician interview	No volumetric differences
De Bellis et al., 2001	sMRI, hand tracing	9/18	8-12	PTSD	SA	Social services documentation	No volumetric differences
Driessen et al., 2000	sMRI, hand tracing	21/42	22-35	BPD	PN, EN, PA, EA, SA	Self report	Volumetric decrease
Edmiston et al., 2011	sMRI, VBM	42*	12-17	None	PN, EN, PA, EA, SA	Self report	Volumetric decrease correlated with frequency
Thomaes et al., 2010	sMRI, VBM	31/59	22-47	PTSD	PA, SA	Clinician interview	No volumetric differences
Tottenham et al., 2010	sMRI, automated segmentation	34/62	5-15	Anxiety d/o, ADHD, ODD,	Previous institutionalization	Social services documentation	Volumetric increase for late adopted relative to early adopted and control
Van Harmelen et al., 2010	sMRI, VBM	84/145	35-39	MDD or Anxiety d/o	EN, PA, EA, SA	Clinician interview	No volumetric differences

Table 1: Summary of MRI literature in populations with childhood maltreatment exposure. fMRI, functional magnetic resonance imaging; sMRI, structural MRI; VBM, voxel-based morphometry; PTSD, post-traumatic stress disorder; MDD, major depressive disorder; ADHD, attention deficit hyperactivity disorder; ODD, oppositional defiant disorder; PN, physical neglect; EN, emotional neglect; EA, emotional abuse; SA, sexual abuse.

*individual differences regression analysis

faces is a consistent finding in adults⁹, adolescents¹⁰, and children¹¹⁻¹³ exposed to childhood maltreatment. In one study, participants with maltreatment demonstrated increased sensitivity to angry faces during an emotional face-morphing paradigm. Participants viewed a set of faces that morphed along a continuum from happy to fear, happy to sad, angry to fear, or from angry to sad and were asked to identify the emotion when presented with a pair of images from each continuum. Participants with a maltreatment history overrated facial expressions as angry and identified angry facial expressions at a lower intensity than participants that did not experience maltreatment, suggesting enhanced sensitivity to potential threat, with discrimination of potential threat at a reduced sensory threshold and with decreased reaction time¹⁴. Furthermore, a study of adolescents exposed to childhood physical abuse suggests increased attentional allocation to threatening or aversive imagery compared to neutral or positive imagery during an emotional dot-probe task¹⁵. Taken together, the behavioral literature suggests that emotional stimuli are particularly salient to maltreated populations, and that social information, such as faces may be of particular importance given the social nature of early maltreatment. Although adaptive in the context of a threatening upbringing, this enhanced attention to threatening stimuli may mediate increased risk for mood and anxiety disorders later in life¹⁵⁻¹⁶.

Changes in amygdala structure and function are candidate mediators of the enhanced salience of emotional stimuli secondary to maltreatment. Previous functional magnetic resonance imaging (fMRI) studies of PTSD populations exposed to trauma, although not necessarily childhood maltreatment, have implicated the amygdala in emotional processing, and have also shown correlations between increased amygdala activation and PTSD symptom severity (for review, see 17-18). Likewise, fMRI studies of adults with childhood maltreatment exposure have found heightened amygdala activation to threatening or emotional stimuli, including negatively valenced emotional faces¹⁹⁻²⁰ and traumatic reminder words²¹ (Table 1). FMRI studies in children and adolescents report similar findings, with increased amygdala BOLD signal for negatively valenced emotional faces the most often-reported finding²²⁻²⁵. Interestingly, although all of these studies report increased BOLD signal, several have reported heightened amygdala reactivity to both neutral and emotional facial expressions^{20,25}. This discrepancy could be due to a generalized enhancement in threat detection, similar to what has been reported in the behavioral literature, such that neutral faces are perceived as potentially threatening. However, there is also evidence that in typically developing children there is no heightened amygdala activation to fear faces relative to neutral faces, as there is in adults²⁶, suggesting that height-

ened amygdala reactivity to fear faces in maltreated child and adolescents samples may reflect an altered developmental trajectory. In addition to alterations in amygdala BOLD signal magnitude, there is also evidence for changes in amygdala BOLD signal time course. Using an emotional face block design, Garret et al. report that increases in amygdala BOLD are greatest relative to control subjects in the early rather than the late phase for angry and fearful faces, suggesting that maltreated subjects may be primed for potential threat²⁵. Taken together, the fMRI literature suggests that heightened amygdala activation to emotional stimuli may underlie the increased threat sensitivity observed in behavioral studies of maltreated populations, as well as their increased risk for mood and anxiety disorders.

Volumetric MRI studies of the amygdala in childhood maltreatment have been less consistent than the fMRI literature (**Table 1**). Voxel-based morphometry studies of the amygdala in adults have found decreases²⁷ or no differences²⁸⁻³⁰ in amygdala volume, but are generally confounded by the study of participants with psychiatric diagnoses, such as PTSD and Borderline Personality Disorder, that are also associated with amygdala changes. Studies of children and adolescents have also been mixed, with reports of increases³¹, decreases³², and no differences³³ in amygdala volume. Mixed findings may in part be due to the still-developing nature of limbic-prefrontal circuits in adolescence, such that early stress exposure may trigger precocious amygdala development and sensitization, followed by volumetric decreases³⁴. Furthermore, the issue of risk vs. resilience confounds studies of adolescents or children with maltreatment exposure and no current psychiatric diagnosis. Some adolescents may still develop a mood or anxiety disorder in adulthood, but it is also possible that a study of young adults or late adolescents without psychiatric disorders has sampled the most resilient members of the maltreated population; the reported amygdala volume reductions could be a marker of resistance to later mood or anxiety symptoms, thereby making conclusions about the structure of the amygdala in at-risk populations difficult. However, a recent study using an individual differences approach found a negative correlation between reported maltreatment severity and amygdala volume in adolescents, particularly in emotional maltreatment, suggesting that part of the heterogeneity of previous findings may be due to maltreatment type and the limited sensitivity of group comparison studies to detect potential biologically significant thresholds in maltreatment severity³². An important paper by Tottenham et al. assessed a group of orphans who

experienced early childhood institutionalization in Romania, followed by adoption in the United States. In this study, there were no overall differences in amygdala volume between previously institutionalized children and controls. However, further comparison of early versus late adopted subjects revealed a significant correlation between amygdala volume and age of adoption, such that later adopted children had larger amygdala volumes than early adopted children³¹. Thus, age of maltreatment onset, duration of maltreatment, and age of assessment are all potential mediators of amygdala volumetric findings; future studies with an individual difference-based approach may prove more helpful than group comparison analyses in teasing apart the relative importance of the timing, severity and duration of maltreatment on amygdala development.

Amygdala Alterations in Rodent Models of Early Life Stress

Rodent models of early life stress effects on the amygdala have been more consistent than the human subjects literature. Both juvenile and adults rodents exposed to chronic immobilization stress have increased dendritic arborization of pyramidal neurons in the basolateral amygdala, in contrast to cells in the CA3 region of the hippocampus, which typically show dendritic atrophy. These chronically stressed rodents also show increased anxiety and depressive-like phenotypes as measured in behavioral tasks such as the elevated plus maze³⁵ and forced swim task³⁶. Other rat studies have employed maternal separation paradigms, a model of early life stress in rodents, where pups are separated from their mothers for long or short periods of time. One maternal separation study has shown that the long-separated rats exhibited down-regulation of the serotonin transporter (5-HTT) as well as of inhibitory 5-HT_{1A} receptors in the amygdala. Long-separated rats also demonstrated an anxiety phenotype compared to short-separated rats, as assessed by the open field test³⁷. Other rodent studies of chronic glucocorticoid administration have suggested down-regulation of the 5-HT_{1A} receptor subtype and up-regulation of 5-HT_{2A} receptor subtype³⁸. It is thought that in limbic regions, activation of 5-HT_{1A} receptors may be anxiolytic, while serotonergic innervation of 5-HT_{2A} receptors may be anxiogenic (for review, see 39). These region and receptor subtype specific alterations may explain the heightened amygdala reactivity common in populations exposed to early life stress.

Rodent models are better able to assess differential effects of chronic versus acute stress than human studies. In a rodent study employing both chronic and acute immobili-

zation stress, chronic stress was associated with increased dendritic arborization in the basolateral amygdala the day after stress termination. In contrast, after a single acute stress exposure, increased dendritic spine density was observed in the basolateral amygdala, but only after a delay period of ten days⁴⁰. Given the role of the amygdala in fear conditioning, this is likely an adaptive mechanism, allowing for priming for future potential stressors after early environmental uncertainty. Taken together, the rodent literature suggests increases in dendritic arborization that are specific to the amygdala after early stress; these changes may be secondary to alterations in serotonin receptor subtype density caused by excessive glucocorticoid exposure. Although caution is important when making inferences about human MRI literature on the basis of rodent models, it is possible that the heterogeneity in findings in the human literature is due to varying durations between maltreatment and assessment, or differences between chronic and acute stress effects.

5HTTLPR x Stress Effects on Depression Risk

Both the serotonin-mediated anxiety phenotypes in rodent models of early stress exposure, as well as the use of selective serotonin reuptake inhibitors (SSRIs) in depression treatment, suggest a 5-HTT mediated mechanism for observed functional and structural changes in the amygdala secondary to childhood maltreatment. The serotonin transporter is involved in the regulation of synaptic serotonin via reuptake of serotonin from the synapse into the presynaptic cell. The 5-HTT gene-linked polymorphic region (5-HTTLPR) is located on the 5' regulatory area of the serotonin transporter gene and has been of particular interest in the study of increased risk for depression in the presence of early life stress⁴¹. The 5-HTTLPR, originally thought to have a biallelic expression, has recently been shown to have a triallelic expression, with a short and long form allele. The long form allele has an L_G and L_A form, such that the L_G form is functionally similar to the low-expressing short or "S" allele. The S or L_G alleles are dominantly expressed and result in reduced transcription of SLC6A4A and decreased presence of 5-HTT at the synaptic membrane⁴².

A landmark paper by Caspi et al. employed a population-based methodology to assess for a gene x environment relationship on depression rates. Their findings suggest an increased rate of depression for adult carriers of the S allele of 5-HTTLPR who also reported exposure to stress⁴³. This study is one of the first studies to demonstrate a gene x environment effect of enhanced risk for a psychiatric disorder.

However, there has been much controversy surrounding the reported observation, as some studies have been unable to replicate the 5-HTTLPR x stress finding⁴⁴⁻⁴⁷. This may be due to differences in stress assessment between studies. For example, types of stress exposure assessments vary wildly across studies; some have employed participant interviews both with and without medical record confirmation of maltreatment, which may be subject to retrospective bias. Others have used questionnaires to assess for stress exposure, ranging from brief, four question assessments for the presence or absence of physical or sexual abuse history^{45,47}, to detailed multi-item questionnaires that assess for the severity, timing, and duration of a host of different early life stressors⁴⁸⁻⁴⁹. Importantly, many of these studies have assessed for only physical or sexual abuse, which, although associated with an increased risk for later psychopathology, may be less salient than emotional maltreatment for both increased risk of later psychopathology^{3,50} and amygdala volume alterations³². Furthermore, later studies of the 5-HTTLPR polymorphism have employed a triallelic analysis of the polymorphism, whereas other, earlier studies employed a biallelic (short or long) analysis^{48,51-53}. Although the original Caspi et al. paper employed a biallelic analysis, differences in the underlying distribution of the L_G vs. L_A allele that were not assessed may account for heterogeneity of findings in the biallelic literature. Finally, recent studies have reported a sex x genotype x environment effect on 5-HTTLPR polymorphism and increased risk for depression, with only women with the low-expressing allelic variants showing enhanced risk for depression in the presence of childhood maltreatment⁵⁴. This may explain the failure to replicate in studies that did not consider sex difference effects. Despite the heterogeneity of findings and methods in the literature, recent meta-analysis has suggested a positive finding, although with a smaller effect size than the original Caspi report⁵⁵.

5HTTLPR x Stress Effects on Amygdala Activation

Given the controversy regarding the enhanced risk for depression in short allele carriers with early life stress exposure, more recent literature has examined the effect of gene x environment on amygdala activation using fMRI. This literature has consistently found increased BOLD signal to emotional or threatening stimuli in the low-expressing allele carriers exposed to maltreatment⁵⁵. Other studies have found decreased functional coupling of the amygdala with prefrontal regions, such as the anterior cingulate, which are thought to down regulate amygdala activity during emotional processing⁵⁶. These findings suggest that the amygdala

dala has a heightened reactivity to emotional or potential threatening stimuli in low expressing carriers exposed to childhood maltreatment, and/or that there is decreased functional coupling between the amygdala and prefrontal cortex in this population. Combined genetic neuroimaging studies may be more fruitful in the study of risk for psychiatric disorder. Enhanced risk for depression is an end-point outcome well downstream of a host of genetic and environmental factors that interact dynamically throughout development; diagnostic categories are heterogeneous and depressive symptoms are likely the result of multiple potential biological causes and diagnostic categories. Therefore, by examining the association between genotype and intermediate phenotypes, such as BOLD signal or well-defined clinical subgroups, studies are better able to explore possible mechanisms for enhanced risk for depression by focusing on a single (or cluster of related) biological factors.

Concluding Remarks and Future Directions

MRI findings in amygdala volume and activation following childhood maltreatment suggest that this brain region is a significant contributor to emotional and behavioral alterations observed in maltreatment-exposed adults and children, including increased risk for mood and anxiety disorders. However, this literature is varied and complex due to a variety of factors that may moderate amygdala development, including 5-HTTLPR genetic polymorphisms and sex, as well as timing, duration, and type of maltreatment exposure. Future studies should address these possible interactive effects by assessing for severity and type of maltreatment, as well as for gene x environment interactive effects. Understanding the mechanism behind observed gene x maltreatment effects for depression risk will likely involve a close examination of 5-HT system alterations across development. Importantly, a better understanding of 5-HT system changes has the chance to improve clinical treatment of adult patients with maltreatment history and depression, as some studies have suggested that these patients, as well as patients with the low-expressing 5-HTTLPR allele, may be poor-responders to SSRI treatment⁵⁷⁻⁵⁸. Variations in treatment response may be due to underlying differences in serotonergic system function in patient subpopulations with the 5-HTTLPR risk allele and/or with childhood maltreatment. As study of brain alterations following childhood maltreatment is necessarily difficult for ethical reasons in humans, rodent models of early life stress are particularly important in dissociating the varying developmental effects of different types of maltreatment and

maltreatment duration on amygdala structure and function. Translational research to assess potential alterations in 5-HT receptor density and mechanisms underlying resultant structural and functional remodeling of limbic circuits is necessary to inform observed changes in the human behavioral, genetic, and neuroimaging maltreatment literature.

References

1. US Department of Health and Human Services, Administration for Children and Families, Children's Bureau. Child Maltreatment annual reports: reports from the states to the National Child Abuse and Neglect data systems: national statistics on child abuse and neglect. 2010. <http://www.acf.hhs.gov/programs/cb/pubs/cm09/index.htm>.
2. Spinhoven P, Elzinga BM, Hovens JG, Roelofs K, Zitman FG, van Oppen P and Penninx BW (2010). The specificity of childhood adversities and negative life events across the life span to anxiety and depressive disorders. *J Affect Disord.* 126(1-2): 103-112.
3. Briere J and Rickards S (2007). Self-awareness, affect regulation, and relatedness: differential sequels of childhood versus adult victimization experiences. *J Nerv Ment Dis.* 195(6): 497-503.
4. Lee V and Hoaken PN (2007). Cognition, emotion, and neurobiological development: mediating the relation between maltreatment and aggression. *Child Maltreat.* 12(3): 281-298.
5. Shields A and Cicchetti D (2001). Parental maltreatment and emotion dysregulation as risk factors for bullying and victimization in middle childhood. *J Clin Child Psychol.* 30(3): 349-363.
6. Pollak SD (2003). Experience-dependent affective learning and risk for psychopathology in children. *Ann N Y Acad Sci.* 1008(1): 102-111.
7. Davis M and Whalen PJ (2001). The amygdala: vigilance and emotion. *Mol Psychiatry.* 6(1): 13-34.
8. Price JL and Drevets WC (2010). Neurocircuitry of mood disorders. *Neuropsychopharmacology.* 35(1): 192-216.
9. Gibb BE, Schofield CA and Coles ME (2009). Reported history of childhood abuse and young adults' information-processing biases for facial displays of emotion. *Child Maltreat.* 14(2): 148-156.
10. Leist T and Dadds MR (2009). Adolescents' ability to read different emotional faces relates to their history of maltreatment and type of psychopathology. *Clin Child Psychol Psychiatry.* 14(2): 237-250.
11. Pollak SD, Cicchetti D, Hornung K and Reed A (2000). Recognizing emotion in faces: developmental effects of child abuse and neglect. *Dev Psychol.* 36(5): 679-688.
12. Pollak SD and Sinha P (2002). Effects of early experience on children's recognition of facial displays of emotion. *Dev Psychol.* 38(5): 784-791.

13. Pollak SD and Kistler DJ (2002). Early experience is associated with the development of categorical representations for facial expressions of emotion. *Proc Natl Acad Sci USA*. 99(13): 9072-9076.
14. Masten CL, Guyer AE, Hodgson HB, McClure EB, Charney DS, Ernst M, Kaufman J, Pine DS and Monk CS (2008). Recognition of facial emotions among maltreated children with high rates of post-traumatic stress disorder. *Child Abuse Negl*. 32(1): 138-153.
15. Kimonis ER, Frick PJ, Munoz LC and Aucoin KJ (2008). Callous-unemotional traits and the emotional processing of distress cues in detained boys: testing the moderating role of aggression, exposure to community violence, and histories of abuse. *Dev Psychopathol*. 20(2): 569-589.
16. Dalgleish T, Taghavi R, Neshat-Doost H, Moradi A, Canterbury R and Yule W (2003). Patterns of processing bias for emotional information across clinical disorders: a comparison of attention, memory, and prospective cognition in children and adolescents with depression, generalized anxiety, and posttraumatic stress disorder. *J Clin Child Adolesc Psychol*. 32(1): 10-21.
17. Wood and Hedges 2008
18. Shin LM, Rauch SL and Pitman RK (2006) Amygdala, medial prefrontal cortex, and hippocampal function in PTSD. *Ann N Y Acad Sci*. 1071: 67-69.
19. Grant MM, Cannistraci C, Hollon SD, Gore J and Shelton R (2011). Childhood trauma history differentiate amygdala response to sad faces within MDD. *J Psychiatr Res*. 45(7): 886-895.
20. Van Harmelen AL, van Tol MJ, Demenescu LR, van der Wee NJ, Veltman DJ, Aleman A, van Buchem MA, Spinhoven P, Penninx BW and Elzinga BM (2012). Enhanced amygdala reactivity to emotional faces in adults reporting childhood maltreatment. *Soc Cogn Affect Neurosci*. In Press.
21. Protopopescu X, Pan H, Tuescher O, Cloitre M, Goldstein M, Engelien W, Epstein J, Yang Y, Gorman J, LeDoux J, Silbersweig D and Stern E (2005). Differential time courses and specificity of amygdala activity in posttraumatic stress disorder subjects and normal control subjects. *Biol Psychiatry*. 57(5): 464-473.
22. Maheu FS, Dozier M, Guyer AE, Mandell D, Peloso E, Poeth K, Jenness J, Lau JY, Ackerman JP, Pine DS and Ernst M (2010). A preliminary study of medial temporal lobe function in youths with a history of caregiver deprivation and emotional neglect. *Cogn Affect Behav Neurosci*. 10(1): 34-49.
23. **McCrory EJ, De Brito SA, Sebastian CL, Mechelli A, Bird G, Kelly PA and Viding E (2011). Heightened neural reactivity to threat in child victims of family violence. *Curr Biol*. 21(23): R947-948.**
Important study showing behavioral alterations in face perception following maltreatment.
24. Tottenham N, Hare TA, Millner A, Gihooly T, Zevin JD and Casey BJ (2011). Elevated amygdala response to faces following early deprivation. *Dev Sci*. 14(2): 190-204.
25. Garret AS, Carrion V, Kletter H, Karchemskiy A, Weems CF and Reiss A (2012). Brain activation to facial expressions in youth with PTSD symptoms. *Depress Anxiety*. 29(5): 449-459.
26. Thomas KM, Drevets WC, Whalen PJ, Eccard CH, Dahl RE, Ryan ND and Casey BJ (2001). Amygdala response to facial expression in children and adults. *Biol Psychiatry*. 49(4): 309-316.
27. Driessen M, Herrmann J, Stahl K, Zwaan M, Meier S, Hill A, Osterheider M and Petersen D (2000). Magnetic resonance imaging volumes of the hippocampus and the amygdala in women with borderline personality disorder and early traumatization. *Arch Gen Psychiatry*. 57(12): 1115-1122.
28. Bremner JD, Randall P, Vermetten E, Staib L, Bronen RA, Mazure C, Capelli S, McCarthy G, Innis RB and Charney DS (1997). Magnetic resonance imaging-based measurement of hippocampal volume in posttraumatic stress disorder related to childhood physical and sexual abuse—a preliminary report. *Biol Psychiatry*. 41(1): 23-32.
29. Thomaes K, Dorrepaal E, Draijer N, de Ruiter RM, van Balkom AJ, Smit JH and Veltman DJ (2010). Reduced anterior cingulate and orbitofrontal volumes in child abuse-related complex PTSD. *J Clin Psychiatry*. 71(12): 1636-1644.
30. Van Harmelen AL, van Tol MJ, van der Wee NJ, Veltman DJ, Aleman A, Spinhoven P, van Buchem MA, Zitman FG, Penninx BW and Elzinga BM (2010). Reduced medial prefrontal cortex volume in adults reporting childhood emotional maltreatment. *Biol Psychiatry*. 68(9): 832-38.
31. **Tottenham N, Hare TA, Quinn BT, McCarry TW, Nurse M, Gilhooly T, Millner A, Galvan A, Davidson MC, Elgsti IM, Thomas KM, Freed PJ, Booma ES, Gunnar MR, Altemus M, Aronson J and Casey BJ (2010). Prolonged institutional rearing is associated with atypically large amygdala volume and difficulties in emotion regulation. *Dev Sci*. 13(1): 46-61.**
First study to show effects of maltreatment duration on amygdala volume.
32. Edmiston EE, Wang F, Mazure CM, Guiney J, Sinha R, Mayes LC and Blumberg HP (2011). Corticostriatal-limbic gray matter morphology in adolescents with self-reported exposure to childhood maltreatment. *Arch Pediatr Adolesc Med*. 165(12): 1069-1077.
33. De Bellis MD, Hall J, Boring AM, Frustaci K and Moritz G (2001). A pilot longitudinal study of hippocampal volumes in pediatric maltreatment-related posttraumatic stress disorder. *Biol Psychiatry*. 50(4): 305-309.
34. Tottenham N and Sheridan MA (2010). A review of adversity, the amygdala and the hippocampus: a consideration of developmental timing. *Front Hum Neurosci*. 8(3): 68.
35. **Vyas A, Mitra R, Shankaranarayana R and Chattarji S (2002). Chronic stress induces contrasting patterns of dendritic remodeling in hippocampus and amygdaloid neurons. *J Neurosci*. 22(15): 6810-6818.**
This paper shows important differences in morphological effects of chronic stress on amygdala versus hippocampal neurons.
36. Elland L, Ramroop J, Hill MN, Manley J and McEwen BS (2012). Chronic juvenile stress produces corticolimbic dendritic architectural remodeling and modulates emotional behavior in male and female rats. *Psychoneuroendocrinology*. 37(1): 39-47.
37. Vicentic A, Francis D, Moffett M, Lakatos A, Rogge G, Hubert GW, Harley J and Kuhar MJ (2006). Maternal separation alters serotonergic transporter densities and serotonergic 1A receptors in the rat brain. *Neuroscience*. 140(1): 355-365.
38. Watanabe Y, Sakai RR, McEwen BS and Mendelson S (1993). Stress and antidepressant effects on hippocampal and cortical 5-HT1a and 5-HT2 receptors and transport sites for serotonin.

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Brain Res. 615(1): 87-94.

39. Leonard BE (2005). The HPA and immune axes in stress: the involvement of the serotonergic system. *Eur Psychiatry. Suppl3*: S302-S306.
40. Mitra R, Jadhav S, McEwen BS, Vyas A and Chattarji S (2005). Stress duration modulates the spatiotemporal patterns of spine formation in the basolateral amygdala. *Proc Natl Acad Sci U S A*. 102(26): 9371-9376.
41. Lesch KP, Bengel D, Heils A, Sabol SZ, Greenberg BD, Petri S, Benjamin J, Muller CR, Hamer DH and Murphy DL (1996). Association of anxiety-related traits with a polymorphism in the serotonin transporter gene regulatory region. *Science*. 274(5292): 1527-1531.
42. Nakamura N, Ueno S, Sano A and Tanabe H (2000). The human serotonin transporter gene linked polymorphism (5-HTTLPR) shows ten novel allelic variants. *Mol Psychiatry*. 5(1): 32-38.
43. **Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, McClay J, Mill J, Martin J, Braithwaite A and Poulton R (2003). Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science*. 301(386): 386-389.**
Important study showing interactive effect between environment and genotype on risk for depression.
44. Surtees PG, Wainwright NW, Willis-Owens SA, Luben R, Day NE and Flint J (2005). Social adversity, the serotonin transporter, (5-HTTLPR) polymorphism and major depressive disorder. *Biol Psychiatry*. 59(3): 224-229.
45. Sakai JT, Lessem JM, Haberstick BC, Hopfer CJ, Smolen A, Ehlinger MA, Timberlake D and Hewitt JK (2006). Case-control and within-family tests for association between 5HTTLPR and conduct problems in a longitudinal adolescent sample. *Psychiatric Genetics*. 17: 207-214.
46. Chipman P, Jorm AF, Prior F, Sanson A, Smart D, Tan X and Eastaugh S (2007). No interaction between the serotonin transporter polymorphism (5-HTTLPR) and childhood adversity or recent stressful life events on symptoms of depression: results from two community surveys. *Am J Med Genet Part B*. 144(B): 561-565.
47. Aslund C, Leppert Jerzy, Comasco E, Nordquist N, Orelund L and Nilsson KW (2009). Impact of the interaction between the 5HTTLPR polymorphism and maltreatment on adolescent depression: a population-based study. *Behav Genet*. 39: 534-531.
48. Frodl T, Reinhold E, Koutsouleris N, Donohoe G, Bondy B, Reiser M, Moller HJ and Meisenzahl EM (2010). Childhood stress, serotonin transporter gene, and brain structures in major depression. *Neuropsychopharmacology*. 35: 1383-1390.
49. Klauke B, Deckert J, Reif A, Pauli P, Zwanzger P, Baumann C, Arolt V, Glockner-Rist A and Domschke K (2011). Serotonin transport gene and childhood trauma - a G x E effect on anxiety sensitivity. *Depression and Anxiety*. 28: 1048-1057.
50. Iffland B, Sansen LM, Catani C and Neuner F (2012). Emotional but not physical maltreatment is independently related to psychopathology in subjects with various degrees of social anxiety: a web-based internet survey. *BMC Psychiatry*. 12(49).
51. Steiger H, Richardson J, Joober R, Gauvin L, Israel M, Bruce KR, Kin NY, Howard and Young SN (2007). The 5HTTLPR polymorphism, prior maltreatment and dramatic-erratic personality manifestations in women with bulimic syndromes. *Rev Psychiatr Neurosci*. 32(5): 354-362.
52. Steiger H, Richardson J, Joober R, Israel M, Bruce KR, Kin NY, Howard H, Anestin A, Dandurand C and Gauvin L (2008). Dissocial behavior, the 5HTTLPR polymorphism, and maltreatment in women with bulimic syndromes. *Am J Med Genet Part B*. 147(B): 128-130.
53. Wiggins JL, Bedoyan JK, Peltier SJ, Ashinoff S, Carrasco M, Weng SJ, Welsh RC, Martin DM and Monsk CS (2012). The impact of serotonin transporter (5-HTTLPR) genotype on the development of resting-state functional connectivity in children and adolescents: a preliminary report. *Neuroimage*. 59: 2760-2770.
54. Eley TC, Sugden K, Corsico A, Gregory AM, Sham P, McGuffin P, Plomin R and Craig IW (2004). Gene-environment interaction analysis of serotonin system markers with adolescent depression. *Mol Psychiatry*. 9(10): 908-915.
55. Munafo MR, Brown SM and Hariri AR (2008). Serotonin transporter (5-HTTLPR) genotype and amygdala activation: a meta-analysis. *Biol Psychiatry*. 63(9): 852-857.
56. Pezawas L, Meyer-Lindenberg A, Drabant EM, Verchinski BA, Munoz KE, Kolachana BS, Egan MF, Mattay VS, Hariri AR and Weinberger DR (2005). 5-HTTLPR polymorphism impacts human cingulate-amygdala interactions: a genetic susceptibility mechanism for depression. *Nat Neurosci*. 8(6): 828-834.
57. Miniati M, Rucci P, Benvenuti A, Frank E, Buitendijk J, Giogi G and Cassano GB (2010). Clinical characteristics and treatment outcome of depression in patients with and without a history of emotional and physical abuse. *J Psychiatr Res*. 44(5): 302-309.
58. Lee SH, Choi TK, Lee E, Seok JH, Lee SH, Lee HS and Kim SJ (2010). Serotonin transporter gene polymorphism associated with short-term treatment response to venlafaxine. *Neuropsychobiology*. 62(3): 198-206.

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G Protein-Coupled Receptor Kinases (GRKs) and G Protein-Coupled Receptors (GPCRs): GRK6 as a Potential Drug Target for CNS Disorders

Mika Garrett

G protein-coupled receptor kinases (GRKs) specifically interact with G protein-coupled receptors (GPCRs) and play an important role in terminating agonist-induced GPCR signaling. Activated by an agonist-stimulated GPCR, GRKs in turn phosphorylate the GPCR, which leads to recruitment and activation of β -arrestin and subsequent receptor desensitization. Through activation of β -arrestins, GRK can also participate in downstream signaling events. GRK6, one of the GRK isoforms, is expressed abundantly in the brain. In recent years, potential links between GRK6 and central nervous system (CNS) disorders, in particular Parkinson's disease (PD), have been suggested. Specifically, GRK6 appears to play a significant role in dopamine-mediated locomotor activity and dopamine agonist-induced dyskinesia. Therefore, GRK6 should be considered as a potential drug target for CNS disorders.

Keywords: *GPCR, GRK, GRK6, Parkinson's disease, dopamine, L-DOPA, dyskinesia*

Mechanism and regulation of GPCR signaling

GPCRs, such as rhodopsin, prostaglandin E_2 receptors and β adrenergic receptors, are members of the seven transmembrane receptor family. G proteins are heterotrimeric complexes comprised of G_α and $G_{\beta\gamma}$ subunits. Upon agonist binding to a GPCR, the G_α subunit exchanges its bound GDP with GTP, which triggers release of G_α and its subsequent association with effectors. The released $G_{\beta\gamma}$ subunit can also stimulate downstream signaling¹. GPCR signaling is controlled by receptor desensitization, a process that involves receptor phosphorylation by G protein-coupled receptor kinases (GRKs) and subsequent β -arrestin binding². β -arrestin binding to the receptor sterically blocks the interaction between GPCR and G proteins, preventing further signaling³. β -arrestins also promote endocytosis by functioning as adaptors between endocytotic elements and the receptor², promoting internalization of GPCRs through clathrin-coated pits⁴. In addition to receptor desensitization, β -arrestins can coordinate the process of signaling termination through degradation of second messengers⁵ such as cAMP⁶. In other instances, arrestins can also act as signaling molecules independent of G-proteins by, for example, binding directly to mitogen activated protein kinases (MAPKs)³. Furthermore, upon GPCR activation, β -arrestin1 can translocate into the nucleus, where it facili-

tates the recruitment of histone acetyltransferase, leading to transcription of genes encoding proteins such as *c-fos*⁷.

Activation of arrestins depends on the phosphorylation of agonist-stimulated GPCRs. Importantly, phosphorylation of GPCRs specifically by GRK enhances the inhibitory effect of arrestins⁴. Furthermore, the pattern of GRK-mediated receptor phosphorylation determines how tightly β -arrestins bind to the activated and phosphorylated receptor². In summary, GPCR activity and signaling are positively and negatively regulated through β -arrestins. Activity of β -arrestins, in turn, depends on the phosphorylation state of the activated GPCR, which is controlled by GRKs.

Overview of G protein-coupled receptor kinases

The seven vertebrate GRKs (GRK1-7) are grouped into three subfamilies: GRK1 (GRK1 and 7), GRK2 (GRK2 and 3), and GRK4 (GRK4, 5 and 6)^{8,9}. GRK1 and 7 are exclusively expressed in photoreceptors, where they phosphorylate rhodopsin^{10,11}. GRK2 and 3 are broadly distributed in the CNS^{12,13}, while expression of GRK4 is mostly found in testis¹⁴. GRK5 is most abundantly expressed in lung, heart, retina, and lingual epithelium, with moderate expression in brain^{50,53,54}. The highest expression of GRK6 is found in brain and skeletal muscle, followed by pancreas, and much

lower levels in other organs¹⁵.

All GRKs have in common a multi-domained structure consisting of (1) the N-terminal region, (2) the regulator of G-protein signaling homology domain, (3) a protein kinase domain, and (4) a variable C-terminal domain¹⁶. In general, the N-terminal region of GRK is believed to recognize the activated form of GPCRs¹⁷. Lipid modification of the C-terminus, as seen in GRK2, is also involved in this process^{18,19}. Binding of a lipid-modified G_{βγ} subunit to the C-terminus of GRK2 facilitates its translocation from cytoplasm to membrane, where the target GPCR is located. In addition, binding of lipids to the C-terminal pleckstrin homology domain can directly regulate GRK2 activity²⁰. In effect, these two mechanisms can synergistically enhance the activity of GRK2^{21,22}. Increases in activity of GRKs by lipid binding have been observed for all members of the GRK4 subfamily²³. In addition to lipid modifications, kinase activity of GRKs can be regulated by other kinases²⁴. For example, GRK5 activity can be inhibited by autophosphorylation promoted by calmodulin (CaM)²⁵⁻²⁷ or by protein kinase C (PKC)-mediated phosphorylation²⁸. Interestingly, GRK2 activity is inhibited by CaM, but this effect is reversed by PKC^{29,30}. Other kinases such as Src and MAPK have also been found to have regulatory effects on GRKs^{31,32}. In another case, Raf kinase inhibitor protein (RKIP), upon its phosphorylation by PKC, releases from its normal target of Raf1 and binds to GRK2, inhibiting its activity. This change in RKIP function from Raf-1 inhibition to GRK2 inhibition results in enhanced receptor signaling³³. Taken together, these findings indicate that the activity of GRKs, and therefore subsequent changes in GPCR-mediated signaling, are regulated by many different mechanisms. The effects of CaM and PKC, for example, suggest a model of coordinated regulation of GPCRs¹⁶. Therefore, any alterations in GRK activity may have a critical impact on cellular functions.

Molecular properties of GRK6

The GRK of most relevance to CNS disorders is GRK6, which is the GRK with highest expression in the brain. Alternative splicing of the C-terminal end of GRK6 yields three variants: GRK6A, B and C, with the sizes of 576, 589, and 560 amino acid residues, respectively³⁴. Only GRK6A has the palmitoylation site within the C-terminal domain, which allows membrane localization³⁵. The C-terminal region of GRK6B contains consensus phosphorylation sites for PKC and cAMP/cGMP-dependent protein kinases that

may contribute to phosphorylation-dependent membrane association. GRK6C, on the other hand, has a truncated C terminus and lacks both the palmitoylation and phosphorylation sites; therefore, it is suspected to have poor membrane association^{34,36}. The existence of a fourth variant, GRK6D, has also been reported. This variant, however, lacks a functional catalytic domain and is speculated to act as an inhibitor of other GRK6 isoforms³⁶.

Palmitoylation of GRK6A has important consequences. Palmitoylated GRK6 is membrane-associated, localized in close proximity to target GPCRs and thus has increased activity^{37,38}. However, palmitoylation is not the only mechanism leading to membrane localization for GRK6 variants. When overexpressed in COS-7 cells, the three variants of GRK6 (A, B and C) are membrane-associated despite the lack of putative C-terminal palmitoylation sites in both GRK6B and C³⁹. Furthermore, the GRK6C isoform has the highest catalytic activity, suggesting that the C-terminal extensions found in variants A and B, but not in variant C, result in decreased activity³⁹. According to the crystal structure, there are two sets of complementary interactions that bring GRK6 into close proximity of a GPCR. First, two regions of GRK6, the α helix region in the N-terminus and the C-terminal tail, directly bind to GPCRs. Second, GRK6 associates with membrane through the phospholipid-binding surface⁴⁰. It has been suggested that the complementary receptor binding provides the free energy needed to induce and stabilize the active form of GRK6^{40,41}.

While the molecular mechanisms of interactions between GRK6 and GPCRs are becoming better understood, studies attempting to identify the native receptor substrate of GRK6 have yielded inconclusive results. Nevertheless, some candidate receptors have emerged. For example, despite the negative results of earlier studies^{42,43}, several, more recent studies verified the M3 muscarinic acetylcholine receptor as a target of GRK6⁴⁴⁻⁴⁶. Other receptors identified as potential targets of GRK6 are the β 2 adrenergic receptor⁴⁷ and the insulin-like growth factor 1 receptor⁴⁸. In the CNS, one study demonstrated that the D₂-like dopamine receptor was a target of GRK6⁴⁹. However, more studies are needed to verify the identity of receptor substrates for GRK6 *in vivo*.

GRK6 involvement in PD and animal models of PD

As mentioned earlier, GRK6 is highly expressed in the brain¹⁵. It reaches its highest expression in the striatum^{13,50},

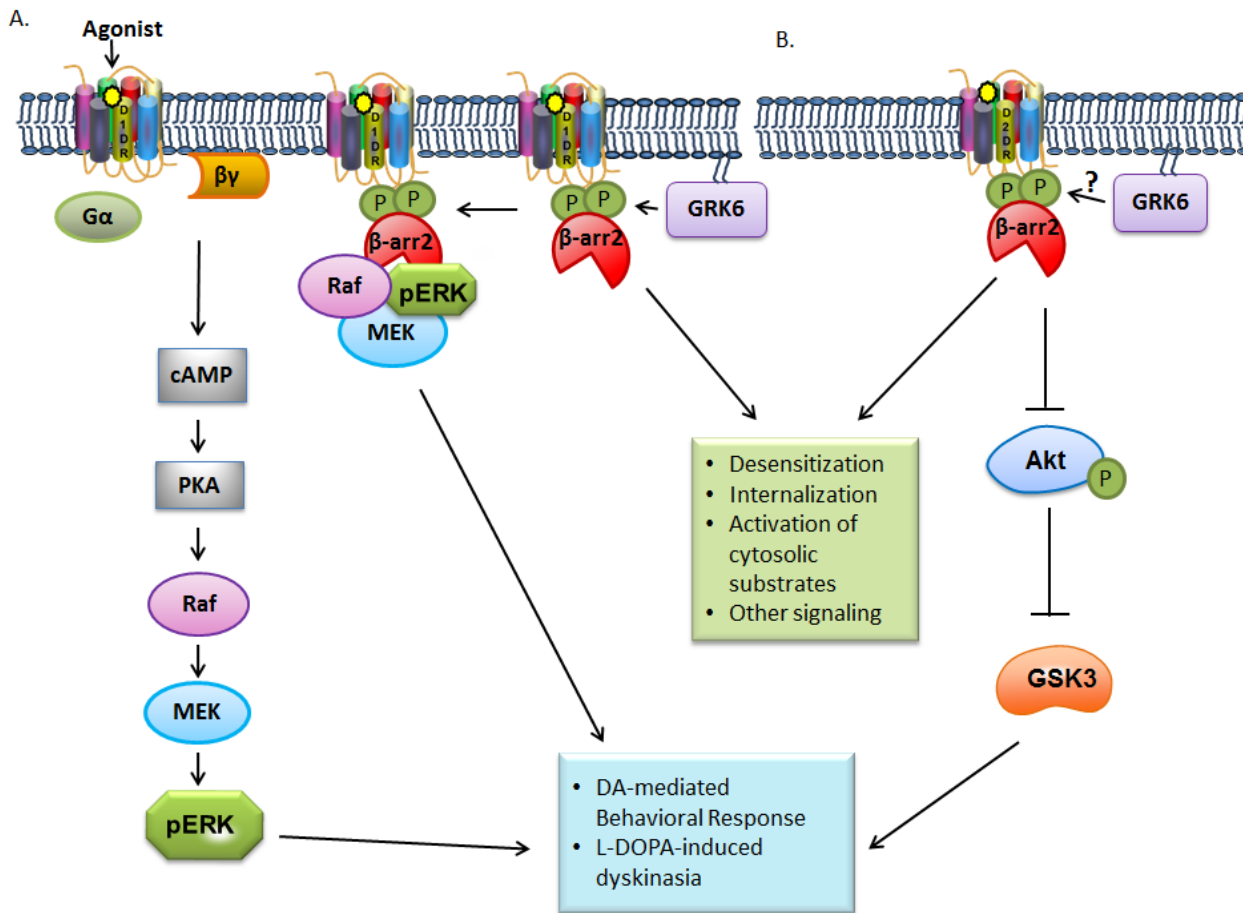


Figure 1: Signal transduction mediated by dopamine receptors that may lead to Parkinson's disease-related motor dysfunction. (A) D1 receptor-mediated signal transduction. G protein-mediated signal transduction (left). Activation of trimeric G protein results in increased level of cAMP and activation of PKA, which leads to activation of MAPK cascade and increased Erk phosphorylation. β-arrestin-dependent activation of MAPK (middle). β-arrestin binds to agonist-activated, GRK-phosphorylated receptor, and acts as signal transducer independent of G proteins. Increased activation of ERK by β-arrestin in this manner may contribute to L-DOPA-induced dyskinesia. β-arrestin binding to the activated receptor also leads to receptor internalization (right), followed by receptor desensitization or activation of other signaling cascades. (B) D2 receptor-mediated β-arrestin dependent signal transduction. β-arrestin inactivates Akt through dephosphorylation, and Akt inactivates GSK3 through phosphorylation. Therefore, β-arrestin contributes to dopamine (DA)-mediated signal activation as well as receptor desensitization and internalization. Increased activity of GSK3 has been linked to L-DOPA-induced dyskinesia.

and changes in GRK6 expression levels have been observed in human clinical conditions as well as in animal models of neurological disorders. Therefore, it has been speculated that alterations in GRK6 activity may contribute to pathophysiology of CNS disorders. One of the CNS disorders in which changes in GRK6 have been reported is Parkinson's disease (PD). For example, in postmortem human brain samples, increased GRK6 mRNA levels were detected among the PD patients who also exhibited dementia compared to PD patients without dementia or the control group⁵⁰.

PD is a neurological disorder characterized by bradykinesia (slowness of movement), rigidity, and tremor. PD

symptoms are the result of a loss of the striatal innervation by dopaminergic neurons in the substantia nigra pars compacta. Currently, L-3,4-dihydroxyphenylalanine (L-DOPA), a precursor of dopamine, is the most effective pharmacological therapy for the treatment of motor symptoms of PD⁵¹. Unfortunately, after 4-6 years of L-DOPA treatment, about 40% of PD patients develop increased involuntary movements known as dyskinesias. After prolonged L-DOPA therapy, almost 90% of patients develop dyskinesia⁵². The loss of striatal dopamine in PD can be modeled in rodents and non-human primates by injection of dopaminergic neurotoxins. In rats, the neurotoxin 6-hydroxydopamine (6-OHDA) is directly injected into the nigrostriatal pathway or the striatum, whereas dopamine depletion in

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non-human primates is achieved by systemic injection of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). Like in humans, dopamine-depleted animals develop dyskinesias following repeated treatment with L-DOPA.

There are a series of studies investigating the role of GRK6 in animal models of PD that were aimed at gaining valuable insight into the molecular mechanisms underlying the altered dopamine signaling in this disorder and, in particular, L-DOPA-induced dyskinesia. Biochemical studies in both MPTP-lesioned monkeys and 6-OHDA-lesioned rats revealed increased GRK6 expression levels in the dorsal and ventral striatum^{53,54}. Behavioral studies have also provided insights into the connection between GRK6 and motor function. A study by Ahmed et al. (2010)⁵⁵ demonstrated that L-DOPA- or dopamine receptor agonist-induced rotation in 6-OHDA-lesioned rats produces a progression of adverse involuntary movements (AIMs). Similar results were seen in MPTP-lesioned monkeys. It was shown that overexpression of GRK6 alleviated AIMs, and down-regulation exacerbated them. However, in another study, 6-OHDA-lesioned GRK6 knockout (KO) mice showed significantly less dyskinesia as measured by AIM score compared to lesioned wild-type (WT) mice⁴⁹. Therefore, while the former study demonstrated that an increase in GRK6 availability alleviated dyskinesias, the latter study showed GRK6 depletion suppressed dyskinesia. Although these results seem conflicting, it should be noted that there is a caveat in using knockout animals. For example, since GRK2 is also expressed in striatum⁵⁴, although at lower levels than GRK6, there may be a compensatory expression of GRK2 or other GRK isoforms in GRK6 KO animals. If that is the case, decreased dyskinesias in GRK6 KO animals may be a result of other GRK isoforms being upregulated. Further studies are needed to address these possibilities.

To further understand the role of GRK6 in dopamine receptor-mediated locomotor activity, Gainetdinov et al. (2003)⁵⁶, using GRK6 KO mice, took a pharmacological approach in the GRK6 KO mice. This study showed that GRK6 KO mice had an increased locomotor response to the D₂ agonist quinpirole. *In vitro*, GRK6-mediated desensitization of D₂-like, but not D₁-like dopamine receptors. In mouse striatum, one possible signaling mechanism downstream of the D₂ dopamine receptor in which GRK6 plays a role involves Akt and glycogen synthase kinase 3 (GSK3)⁵⁷. Activation of β -arrestin 2 via D₂ receptor stimulation leads to dephosphorylation and inactivation of Akt which, through phosphorylation, has an inhibitory effect

on GSK3. In support of GRK6's involvement in this signaling cascade, GRK6 KO mice had significantly higher basal levels of pGSK3 β compared to WT mice⁴⁹. Interestingly, there are also some data suggesting that GRK6 may exert its effect on dyskinesia via the MAPK signaling cascade. For example, after continuing L-DOPA treatment on 6-OHDA-lesioned mice, expression of phosphorylated extracellular signal-regulated kinase 2 (pErk2), one member of the MAPK family, was significantly increased compared to sham-operated control. There were no changes in pErk expression, however, between controls and 6-OHDA-lesioned GRK6-KO mice⁴⁹. In a separate study, Westin et al. (2007)⁵⁸ showed that there was a positive correlation between the severity of L-DOPA-induced dyskinesia and the amount of pErk expression in 6-OHDA-lesioned rat striatum. In addition, in those animals, application of a D₁ antagonist reduced the development of dyskinesia as well as the amount of pErk. Together, these results suggest that dyskinesia induced by the action of L-DOPA on D₁ receptors may involve GRK6 through its activation of the MAPK cascade.

GRK6 as a drug target

Development of drugs for CNS disorders has been less successful than in other areas of disorders because of limited knowledge of the underlying pathophysiology, limitations in current preclinical models of CNS disorders, and a lack of good biomarkers⁵⁹. For treatment of PD, there are many available drugs that include, for example, L-DOPA, D₂ dopamine receptor agonists, catechol-O-methyltransferase inhibitors, and monoamine oxidase inhibitors. Despite the fact that L-DOPA treatment loses efficacy with time and produces adverse side effects, it remains the most widely used therapy today due to the limited efficacy of other drugs⁶⁰. Currently, amantadine, a non-competitive N-methyl-D-aspartate receptor antagonist is the only drug used to alleviate the symptoms of L-DOPA-induced dyskinesia, but prolonged use of amantadine leads to the development of adverse side effects such as dizziness, confusion and hallucinations⁵¹. Another, non-pharmacological approach to symptomatic treatment of PD that can reduce L-DOPA induced dyskinesia is deep brain stimulation (DBS) of the subthalamic nucleus. DBS, however, involves neurosurgeries that are expensive, come with potential risks, and require regular maintenance of the stimulator⁵¹. Because of the limitations associated with all the current therapies, there is a need for continuing efforts to develop novel therapies aimed at new potential drug targets. As an

alternative to drugs that directly activate or inhibit GPCRs, there is the potential for therapeutic value in regulators of GPCR activities¹. Although lagging behind, continuing efforts have been made for development of drugs targeting CNS protein kinases such as GSK3 for Alzheimer's disease and neuropsychiatric disorders, PKC for bipolar disorder, and mammalian target of rapamycin for autism⁶¹. Some interesting properties of GRK-mediated GPCR regulation make GRK an attractive drug target. Probably one of the most intriguing discoveries is the ability of GRK-produced phosphorylation patterns to stimulate distinct downstream signaling through β -arrestin in a GRK variant-specific manner⁶². The mechanism of this process was unlocked in a study by Nobles et al. (2011)⁶³. In this experiment, the authors demonstrated that, although both GRK2 and GRK6 participated in the phosphorylation of the β_2 adrenoreceptor, the functional outcome was different depending on the distinct phosphorylation patterns that were produced by each kinase. Furthermore, binding of different ligands to GPCRs could determine which GRK would be activated.

Conclusions and future directions

As discussed in the previous sections, several lines of evidence suggest that GRK6 has a critical role in PD. Its tissue-specific expression and intricate network of regulatory functions make GRK6 a good drug target for PD treatment. The future of successful drug development may lie in the discovery of selective ligands for GRK6, which may enable precisely targeted treatment for CNS disorders and/or alleviate undesired side effects when used in combination with other GPCR ligands. By manipulating the activity of GRK6, it is possible to selectively target and fine-tune the cellular signaling events that contribute to CNS disease phenotypes downstream of GPCRs.

One of the most prominent drug-discovery techniques, high-throughput screening (HTS), is a promising route to identifying possible GRK6-targeted drugs. HTS of compound libraries has evolved into a successful approach for discovering novel drugs for both CNS disorders and other indications. Examples of successful HTS efforts include the discovery of drugs that target tyrosine kinase for cancer treatment (e.g. Gefitinib, Lapatinib), proteases for HIV (Tipranavir) and GPCRs for hypertension (Ambrisentan)⁶⁴. It should be noted, however, that screening for drugs that target protein kinases is in its infancy, and there are undoubtedly many challenges in the process. Some of the difficulties include assay design, finding compounds with

sufficient affinity and selectivity for the target as well as appropriate chemical properties for CNS penetrance⁶¹. Another inherent challenge is that most protein kinase inhibitors identified by HTS are ATP derivatives that compete for the binding site with endogenous ATP⁶⁵. Therefore, it will require careful planning and optimization of the assay conditions before screening attempts should be made. With utilization of proper conditions and technique, the HTS approach may lead to the discovery of compounds that selectively act on GRK6 and could be used to further define the role of GRK6 in PD.

References

1. Neubig RR and Siderovski DP (2002). Regulators of G-protein signaling as new central nervous system drug targets. *Nat. Rev. Drug. Disc.* **1**: 187-197.
2. Lefkowitz RJ and Shenoy SK (2005). Transduction of receptor signals by β -arrestins. *Science*, **308**: 512-517.
3. Lefkowitz RJ and Whalen EJ (2004). β -arrestins: traffic cops of cell signaling. *Curr. Opin. Cell. Biol.* **16**:162-168.
4. Krupnick JG and Benovic JL (1998). The role of receptor kinases and arrestins in G protein-coupled receptor regulation. *Ann. Rev. Pharmacol. Toxicol.* **38**: 289-319.
5. Nelson CD, Perry SJ, Regier DS, Prescott SM, Topham MK and Lefkowitz RJ (2007). Targeting of diacylglycerol degradation to M1 muscarinic receptors by β -arrestins. *Science*. **315**: 663-666.
6. Perry SJ, Baillie GS, Kohout TA, McPhee I, Magiera MM, Ang KL, Miller WE, McLean AJ, Conti M, Houslay MD and Lefkowitz RJ (2002). Targeting of cyclic AMP degradation to β_2 -adrenergic receptors by β -arrestins. *Science*. **298**: 834-836.
7. Kan J, Shi Y, Xiang B, Qu B, Su W, Zhu M, Zhang M, Bao G, Wang F, Zhang X, Yang R, Fan F, Chen X, Pei G and Ma, L (2005). A nuclear function of β -arrestin 1 in GPCR signaling: Regulation of histone acetylation and gene transcription. *Cell*. **123**: 833-847.
8. Premont RT, Koch WJ, Inglese J and Lefkowitz RJ (1994). Identification, purification, and characterization of GRK5, a member of the family of G protein-coupled receptor kinases. *J. Biol. Chem.* **269**(9): 6832-6841.
9. Gurevich EV, Tesmer JJ, Mushegian A and Gurevich VV (2012) G protein-coupled receptor kinases: More than just kinases and not only for GPCRs. *Pharm. Ther.* **133**: 40-69.
10. Zhao X, Huang J, Khani SC and Palczewski KJ (1998). Molecular forms of human rhodopsin kinase (GRK1). *Biol. Chem.* **273**(9): 5124-5131.
11. Rinner O, Makhankov YV, Biehlmaier O and Neuhaus SC (2005). Knockdown of cone-specific kinase GRK7 in larval zebrafish leads to impaired cone response recovery and delayed dark adaptation. *Neuron*. **47**(2): 231-242.
12. Arriza JL, Dawson TM, Simerly RB, Martin LJ, Caron MG,

CANDIDATE REVIEWS

- Snyder SH and Lefkowitz RJ (1992). The G-protein-coupled receptor kinases β ARK1 and β ARK2 are widely distributed at synapses in rat brain. *J. Neurosci.* **12**(10): 4045-4055.
13. Erdtmann-Vourliotis M, Mayer P, Ammon S, Riechert U and Höllt V (2001). Distribution of G-protein-coupled receptor kinase (GRK) isoforms 2, 3, 5 and 6 mRNA in the rat brain. *Mol. Brain. Res.* **95**: 129-137
 14. Premont RT, Macrae AD, Stoffel RH, Chung N, Pitcher JA, Ambrose C, Inglese J, MacDonald ME, Lefkowitz RJ (1996). Characterization of the G protein-coupled receptor kinase GRK4. Identification of four splice variants. *J. Biol. Chem.* **271**(11): 6403-6410.
 15. Benovic JL and Gomez J (1993). Molecular cloning and expression of GRK6: A new member of the G protein-coupled receptor kinase family. *J. Biol. Chem.* **268**(26):19521-19527.
 16. Penn RB, Pronin AN and Benovic J (2000). Regulation of G protein-coupled receptor kinases. *Trends Cardiovasc. Med.* **10**:81-89.
 17. Palczewski K, Buczylo J, Lebioda L, Crabb JW and Plans AS (1993). Identification of the N-terminal region in rhodopsin kinase involved in its interaction with rhodopsin. *J. Biol. Chem.* **268**(8): 6004-6013.
 18. Inglese J, Koch WJ, Caron MG and Lefkowitz RJ (1992). Isoprenylation in regulation of signal transduction by G-protein-coupled receptor kinases. *Nature.* **359**: 147-150.
 19. Pitcher JA, Inglese J, Higgins JB, Arriza JL, Casey PJ, Kim C, Benovic JL, Kwatra MM, Caron MG and Lefkowitz RJ (1992). Role of β subunits of G proteins in targeting the β -adrenergic receptor kinase to membrane-bound receptors. *Science.* **257**: 1264-1267.
 20. Onorato JJ, Gillis ME, Liu Y, Benovic JL and Ruoho AE (1995). The β -adrenergic receptor kinase (GRK2) is regulated by phospholipids. *J. Biol. Chem.* **270**(36): 21346-21353.
 21. DebBurman SK, Ptasinski J, Boetticher E, Lomasney JW, Benovic JL and Hosey MM (1995). Lipid-mediated regulation of G protein-coupled receptor kinases 2 and 3. *J. Biol. Chem.* **270**: 5742-5747.
 22. Pitcher JA, Touhara K, Payne ES and Lefkowitz RJ (1995). Pleckstrin homology domain-mediated membrane association and activation of the β -adrenergic receptor kinase requires coordinate interaction with G β subunits and lipid. *J. Biol. Chem.* **270**(20): 11707-11710.
 23. Pitcher JA, Fredericks ZL, Stone WC, Premont RT, Stoffel RH, Koch WJ and Lefkowitz RJ (1996). Phosphatidylinositol 4, 5-bisphosphate (PIP₂)-enhanced G protein-coupled receptor kinase (GRK) activity. *J. Biol. Chem.* **271**(40): 24907-24913.
 24. Levay K, Satpaev DK, Pronin AN, Benovic JL and Slepak VZ (1998). Localization of the sites for Ca²⁺-binding proteins on G protein-coupled receptor kinases. *Biochemistry.* **37**: 13650-13659.
 25. Chuang TT, Paolucci L and Blasi AD (1996). Inhibition of G protein-coupled receptor kinase subtypes by Ca²⁺/Calmodulin. *J. Biol. Chem.* **271**(45): 28691-28696.
 26. Pronin AN, Carman CV and Benovic JL (1998). Structure-function analysis of G protein-coupled receptor kinase-5. *J. Biol. Chem.* **273**(47): 31510-31518.
 27. Pronin AN, Satpaev DK, Slepak VZ and Benovic JL (1997). Regulation of G protein-coupled receptor kinases by calmodulin and localization of the calmodulin binding domain. *J. Biol. Chem.* **272**(29): 18273-18280.
 28. Pronin AN and Benovic JL (1997). Regulation of the G protein-coupled receptor kinase GRK5 by protein kinase C. *J. Biol. Chem.* **272**(6): 3806-3812.
 29. Winstel R, Freund S, Krasel C, Hoppe E and Lohse M (1995). Protein kinase cross-talk: Membrane targeting of the β -adrenergic receptor kinase by protein kinase C. *Proc. Natl. Acad. Sci. USA.* **93**:2105-2109.
 30. Krasel C, Dammeier S, Winstel R, Brockmann J, Mischak H and Lohse MJ (2001). Phosphorylation of GRK2 by protein kinase C abolishes its inhibition by calmodulin. *J. Biol. Chem.* **276**(3): 1911-1915.
 31. Fan G, Shumay E, Malbon CC and Wang H (2001). C-Src tyrosine kinase binds the β 2-adrenergic receptor via phosphor-Tyr-350, phosphorylates G-protein-linked receptor kinase 2, and mediates agonist-induced receptor desensitization. *J. Biol. Chem.* **276**(16): 13240-13247.
 32. Elorza A, Sarnago S and Mayor Jr F (2000). Agonist-dependent modulation of G protein-coupled receptor kinase 2 by mitogen-activated protein kinases. *Mol. Pharmacol.* **57**: 778-783.
 33. Lorenz K, Lohse MJ and Quitterer U (2003). Protein kinase C switches the Raf kinase inhibitor from Raf-1 to GRK-2. *Nature.* **426**: 574-579.
 34. Premont RT, Macrae AD, Aparicio SAJR, Kendall HE, Welch JE and Lefkowitz RJ (1999). The GRK4 subfamily of G protein-coupled receptor kinases: Alternative splicing, gene organization, and sequence conservation. *J. Biol. Chem.* **274**(41): 29381-29389.
 35. Stoffel RH, Randall RR, Premont RT, Lefkowitz RJ and Inglese J (1994). Palmitoylation of G protein-coupled receptor kinase, GRK6. *J. Biol. Chem.* **269**(45): 27791-27794.
 36. Moepps B, Vatter P, Frodl R, Waechter F, Dixkens C, Hameister H and Gierschik P (1999). Alternative splicing produces transcripts encoding four variants of mouse G-protein-coupled receptor kinase 6. *Genomics*, **60**: 199-209.
 37. Loudon RP and Benovic JL (1997). Altered activity of palmitoylation-deficient and isoprenylated forms of the G protein-coupled receptor kinase GRK6. *J. Biol. Chem.* **272**(43): 27422-27427.
 38. Stoffel RH, Inglese J, Macrae AD, Lefkowitz RJ and Premont RT (1998). Palmitoylation increases the kinase activity of the G protein-coupled receptor kinase, GRK6. *Biochemistry.* **37**: 16053-16059.
 39. Vatter P, Stoesser C, Samel I, Gierschik P and Moepps B (2005). The variable C-terminal extension of G-protein-coupled receptor kinase 6 constitutes an accessory autoregulatory domain. *FEBS Journal.* **272**: 6039-6051.
 40. Boguth CA, Singh P, Huang C and Tesmer JJ (2010). Molecular basis for activation of G protein-coupled receptor kinases. *EMBO.* **29**: 3249-3259.
 41. Lodowski DT, Tesmer VM, Benovic JL and Tesmer JJ (2006). The structure of g protein-coupled receptor kinase (GRK)-6 defines a second lineage of GRKs. *J. Biol. Chem.* **281**(24): 16785-16793.

42. Debburman SK, Kunapuli P, Benovic JL and Hosey MM (1995). Agonist-dependent phosphorylation of human muscarinic receptors in *Spodoptera frugiperda* insect cell membranes by G protein-coupled receptor kinases. *Mol. Pharmacology*. **47**:224-233.
43. Tsuga H, Okuno E, Kameyama K and Haga T. (1998). Sequestration of human muscarinic acetylcholine receptor hm1-hm5 subtypes: Effect of G protein-coupled receptor kinases GRK2, GRK4, GRK5 and GRK6. *J. Pharmacol. Exp. Therapeutics*. **284**(3): 1218-1226.
44. Willets JM, Challiss RAJ, Kelly E and Nahorski SR (2001). G protein-coupled receptor kinase 3 and 6 use different pathways to desensitize the endogenous M3 muscarinic acetylcholine receptor in human SH-SY5Y cells. *Mol. Pharmacol.* **60**(2): 321-330.
45. Willets JM, Challiss RAJ and Nahorski SR (2002). Endogenous G protein-coupled receptor kinase 6 regulates M3 muscarinic acetylcholine receptor phosphorylation and desensitization in human SH-SY5Y neuroblastoma cells. *J. Biol. Chem.* **277**(18): 15523-15529.
46. Luo J, Busillo JM and Benovic JL (2008). M3 muscarinic acetylcholine receptor-mediated signaling is regulated by distinct mechanisms. *Mol. Pharmacol.* **74**(2): 338-347.
47. Violin JD, DiPilato LM, Yildirim N, Elston TC, Zhang J and Lefkowitz RJ (2008). β 2-adrenergic receptor signaling and desensitization elucidated by quantitative modeling of real time cAMP dynamics. *J. Biol. Chem.* **283**(5): 2949-2961.
48. Zheng H, Worrall C, Shen H, Issad T, Seregard S, Girnita A and Girnita L (2012). Selective recruitment of G protein-coupled receptor kinases (GRKs) controls signaling of the insulin-like growth factor 1 receptor. *Proc. Natl. Acad. Sci.* **109**(18): 7055-7060.
49. **Manago F, Espinoza S, Salahpour A, Sotnikova TD, Caron MG, Premont RT and Gainetdinov RR. The role of GRK6 in animal models of Parkinson's disease and L-DOPA treatment. *Sci. Rep.* **2**, 301; DOI:10.1038/srep00301 (2012).**
- This paper demonstrates how behavioral and cellular responses are altered upon administration of dopamine receptor agonists or L-DOPA in mouse models of PD using GRK6 KO animals.*
50. Bychkov ER, Gurevich VV, Joyce JN, Benovic JL and Gurevich EV (2008). Arrestins and two receptor kinases are upregulated in Parkinson's disease with dementia. *Neurobiol. Aging*. **29**:379-396.
51. Buck K and Ferger B (2010). L-DOPA-induced dyskinesia in Parkinson's disease: a drug discovery perspective. *Drug. Disc. Today*. **15**: 867-875.
52. Ahlskog JE & Muentner MD (2001). Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature. *Movement Disorders*. **16**(3): 448-158.
53. Bezaud E, Gross CE, Qin L, Gurevich VV, Benovic JL and Gurevich EV (2005). L-DOPA reverses the MPTP-induced elevation of the arrestin2 and GRK6 expression and enhanced ERK activation in monkey brain. *Neurobiol. Disease*. **18**: 323-335.
54. Ahmed MR, Bychkov E, Gurevich VV, Benovic JL & Gurevich EV (2008). Altered expression and subcellular distribution of GRK subtypes in the dopamine-depleted rat basal ganglia is not normalized by L-DOPA treatment. *J. Neurochem.* **104**: 1622-1636.
55. **Ahmed MR, Berthet A, Bychkov E, Porras G, Li Q, Bioulac BH, Carl YT, Bloch B, Kook, S, Aubert I, Dovero S, Doudnikoff E, Gurevich VV, Gurevich E and Bezaud E (2010). Lentiviral overexpression of GRK6 alleviates L-DOPA-induced dyskinesia in experimental Parkinson's disease. *Sci. Transl. Med.* **2**(28): 1-9.**
- This paper demonstrates that the symptoms of L-DOPA-induced dyskinesia may be alleviated by manipulating the level of GRK6 in vivo.*
56. **Gainetdinov RR, Bohn LM, Sotnikova TD, Cyr, M, Laakso A, Macrae AD, Torres GE, Kim KM, Lefkowitz RJ, Caron MG and Premont RT (2003). Dopaminergic supersensitivity in G protein-coupled receptor kinase 6-deficient mice. *Neuron*. **38**: 291-303.**
- This is the first paper investigating the effects of GRK6 depletion on dopamine-mediated locomotor activities using GRK6 KO mice.*
57. Beaulieu J, Sotnikova TD, Marion S, Lefkowitz RJ, Gainetdinov RR and Caron MG (2005). An Akt/ β -arrestin 2/PP2A signaling complex mediates dopaminergic neurotransmission and behavior. *Cell*. **122**: 261-273.
58. **Westin JE, Vercammen L, Strome EM, Konradi C and Cenci MA (2007). Spatiotemporal pattern of striatal ERK1/2 phosphorylation in a rat model of L-DOPA-induced dyskinesia and the role of dopamine D1 receptors. *Biol. Psychiatry*. **62**: 800-810.**
- This paper shows that there is a positive correlation between the severity of L-DOPA induced dyskinesia and phosphor-ERK1/2 levels in a rat PD model, and that this effect is D1-receptor dependent.*
59. Schoepp DD (2011). Where will new neuroscience therapies come from? *Nat. Drug. Discovery*. **10**:715-716.
60. Huynh T (2011). The Parkinson's disease market. *Nat. Rev. Drug. Disc.* **10**: 571-572.
61. Chico LK, Van Eldik LJ and Watterson M (2009). Targeting protein kinases in central nervous system disorders. *Nat. Rev. Drug Disc.* **8**: 892-909.
62. Liggett SB (2011). Phosphorylation barcoding as a mechanism of directing GPCR signaling. *Sci. Signaling*. **4**(185): 1-3.
63. Nobles KN, Xiao K, Ahn S, Shukla AK, Lam CM, Rajagopal S, Strachan RT, Huang TY, Bressler EA, Hara MR, Shenoy SK, Gygi SP and Lefkowitz RJ. (2011). Distinct phosphorylation sites on the β 2-adrenergic receptor establish a barcode that encodes differential functions of β -arrestin. *Sci. Signaling*. **4**(185): 1-10.
64. Macarron R, Banks MN, Bojanic D, Burns DJ, Cirovic DA, Garyantes T, Green DV, Hertzberg RP, Janzen WP, Paslay JW, Schopfer U and Sittampalam S (2011). Impact of high-throughput screening in biomedical research. *Nat. Rev. Drugdisc.* **10**: 188-195.
65. Cohen P (2002). Protein kinases – the major drug targets of the twenty-first century? *Nat. Drug. Disc.* **1**, 309-315.

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Using Context to Search Memory: Functional Roles of the Medial Temporal Lobe and Prefrontal Cortex

James Kragel

The ability to search one's past for a specific memory amongst similar occurrences is a hallmark feature of the episodic memory system. Context-based theories of episodic memory retrieval propose that a slowly changing contextual representation allows for the construction of discrete, separable episodes. A review of recent neuroimaging, neuropsychological, and electrophysiological evidence suggests that activity within the medial temporal lobe (MTL) may support such a contextual representation, while control processes within the prefrontal cortex may influence the maintenance and updating of representations within the MTL.

Keywords: *Episodic memory, medial temporal lobe, prefrontal cortex*

Episodic memory:

Memory of events, incorporating contextual information such as the time, place, and emotional state of the memory.

Item recognition:

A memory task in which participants must determine whether a stimulus has been presented previously.

Source memory:

Memory for contextual details associated with a particular episode.

Introduction

Cognitive theories of episodic memory posit that memory retrieval involves patterns of neural activity returning to a previous state^{1,2}, resulting in the subjective experience of 'mental time travel'³. These cognitive models rely on a contextual representation that integrates information over long time scales, representing information about both the environment and internal states. When an item is encoded into memory, associations are formed between the item representation and the current state of context. This associative link allows for two different processes to occur: 1) a state of context can be used to retrieve particular items from memory; and 2) particular items can be used to retrieve contextual information, such as the location or time in which they were studied. When an item is retrieved, the context in which it was studied may also be retrieved, causing the contextual representation to return to its prior state. A growing body of evidence supporting the representation of item information within posterior cortical regions, as well as their reactivation during memory retrieval, will be reviewed. While there is ample support for the neural representation of item-specific information, only recent investigations have begun to search for neural mechanisms that may mediate a contextual representation.

Findings from the neuroimaging and neuropsychological literature that support the role of MTL and prefrontal regions in the processing of contextual information will be reviewed.

Reactivation of cortical activity during retrieval

Recent neuroimaging studies provide evidence that retrieval of a previously bound episode occurs when the cortical activity present when the memory is constructed is reactivated. Studies examining cortical reactivation typically have similar experimental form, in which a particular cue is presented during the encoding of an episodic memory. Neural recording during retrieval, when participants typically perform an item recognition or source memory task associated with a specific cue, has provided evidence that cortical activity related to modality (auditory or visual)^{4,5}, encoding task (imagine or read)⁶, or type of odor⁷ reactivates during these retrieval tasks. Critically, the reactivation of contextual details during these tests is driven by associative retrieval mechanisms, as only the cue is presented during test. Studies of this nature have shown the reactivation in visual and auditory cortex during the associative retrieval of visual^{4,8-11} and auditory information^{4,5}. These findings of reactivation in sensory association cortex are supportive

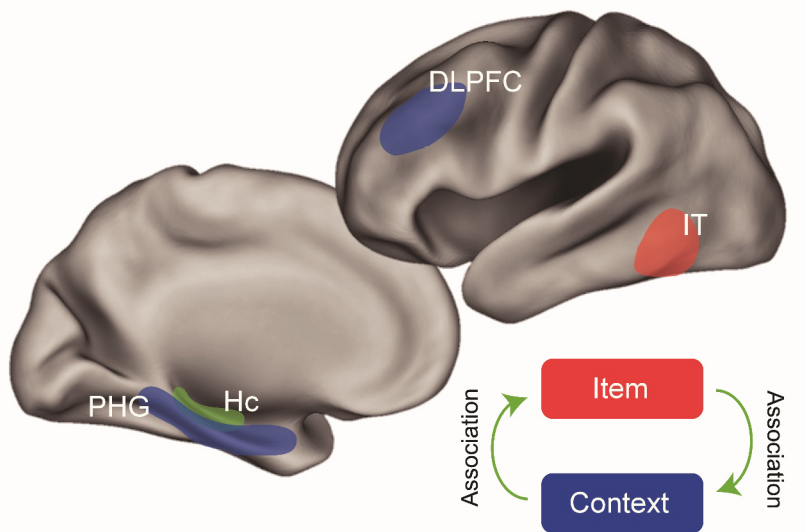


Figure 1: Schematic diagram mapping cortical regions implicated in maintaining and updating context (blue), representing the features of items (red), and forming associations between items and the context in which they are encountered (green). PHG, parahippocampal gyrus. Hc, hippocampus. DLPFC, dorsolateral prefrontal cortex. IT, inferior temporal gyrus.

of context-based theories of episodic memory, which rely on a representation of item information that activates when a specific item is presented during study, or retrieved through mnemonic processing.

In addition to the activation of associated perceptual information, studies have revealed that activity specific to the type of processes engaged during encoding may also be reactivated during retrieval. Work by Johnson and Rugg¹² demonstrated cortical reactivation in left occipital cortex and anterior fusiform gyrus when participants imagined a verbally presented object in a visual scene, while activity in ventromedial prefrontal cortex was reinstated when the same stimulus was incorporated into a sentence. Work by Kahn and colleagues⁶ showed cortical reactivation in the parahippocampal place area (implicated in processing scene information¹³) for imagined items, while items that were read reactivated posterior ventrolateral prefrontal cortex (implicated in lexical tasks). These findings suggest that contextual information beyond sensory perception is bound into episodic memories.

The above neuroimaging studies rely upon the presentation of a cue during test in order to test for the associative retrieval of contextual details. Context-based theories of episodic memory propose that retrieval may occur in the absence of an external cue, as memory

search is guided by an internally maintained contextual cue. In support of the contextual reinstatement hypothesis, Polyn and colleagues¹⁴ found cortical reactivation in the visual processing stream during a free recall task. In this study, fMRI data was collected while participants studied images of famous celebrities, locations, or objects, and later retrieved them during a free-recall task. Using multivariate pattern analysis techniques, activity in fusiform gyrus, parahippocampal cortex, and inferior temporal gyrus was shown to reactivate prior to the recall of celebrities, locations, and objects, respectively. These findings are consistent with category-specific activity cuing memory retrieval. In line with the reactivation of task-specific activity in source memory studies, reactivation of task specific activity in occipital cortex, parietal cortex, and lingual gyrus was observed during a similar free-recall task¹⁵.

Perhaps the most compelling evidence of reactivation of neural activity, driven through the hippocampus, comes from electrophysiological recordings of hippocampal neurons and their surrounding local field potentials from patients with intractable epilepsy. Gelbard-Sagiv and colleagues¹⁶ recorded activity in single neurons of the human hippocampus while patients first viewed cinematic episodes, and later recalled these episodes. A subset of recorded neurons exhibited selective firing throughout

Free-recall task:

A memory paradigm in which participants study a list of items on each trial, and are prompted to recall the items in any order, without a retrieval cue.

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Oscillatory activity:

Synchronized activity of a large number of neurons firing at a specific frequency.

Hebbian learning:

An associative learning process in which connected neurons that exhibit simultaneous firing strengthen their synaptic connections to one another.

the presentation of the encoded materials. Reactivation of these neurons preceded recall of the encoded material, directly linking hippocampal firing and human recollection during memory search. Reactivation of oscillatory activity in the gamma band has also been observed during successful encoding during a free-recall paradigm¹⁷. This activity, localized to prefrontal, hippocampal, and left temporal lobe electrodes was shown to reactivate during the recall of studied items. Reactivation occurred first within the hippocampus, before the pattern of activity in cortex was reinstated. Findings of this nature are suggestive of recall of past experience originating in the MTL, and propagating to association cortex, where activity reflects the subjective experience of the recollected memory.

Bridging cognitive models and neural mechanisms

In the following section, evidence for the neural substrates mediating the major cognitive processes proposed by context-based theories will be briefly reviewed. These theories propose that activity in neocortical association cortex reflects information about individual items recently encountered in the environment. In addition, a context representation contains information about the recent history of items that have been active in association cortex; these theories propose this activity is represented in either prefrontal¹⁸ or parahippocampal¹⁹ regions. Finally, these theories propose that the hippocampus mediates the binding of item and context representations, in the formation of distinct episodes that may be later retrieved. The recall of a specific item, coincident with cortical reactivation of the pattern of neural activity representing said item, is enacted by similar hippocampally mediated Hebbian learning. In this manner, these models provide a means to explain the phenomenon of cortical reactivation during episodic remembering (depicted in **Figure 1**).

Extant neuroimaging evidence speaks to the role of the hippocampus in the successful

encoding^{12, 20-23} and retrieval²³⁻²⁶ of episodic memories. There is also a consensus in terms of object representations in association cortex based on modality: superior temporal gyrus for auditory representations, and inferior temporal gyrus for visual representations. Within the ventral visual pathway, specific cortical regions encode specific stimuli, such as the fusiform face area²⁷, the parahippocampal place area¹³, and object responsive areas in lateral occipital cortex²⁸. Given the recent advent of context-based theories of episodic memory, limited but promising progress has been made towards identifying potential substrates that mediate a contextual representation. In the following section, evidence supporting parahippocampal cortex activity mediating a temporal context representation will be reviewed. Evidence for the role of the dorsolateral prefrontal cortex (DLPFC) in manipulating, or shaping the information stored in this representation will also be considered.

Contextual maintenance and integration

The existence of a slowly changing contextual representation is essential to models of episodic memory. According to the temporal context model¹, changes in context are driven by perceptual inputs to the system, with the similarity of context to prior states decaying in an exponential manner with additional inputs. In order for a region to represent context, patterns of neuronal firing must be capable of changing firing rates based on external inputs (proposed here to reflect perceptual inputs due to feature information), as well as maintain persistent firing over delay periods. Persistent firing enables the maintenance of information in the absence of external stimuli, a critical property of context representations. Persistent firing of cortical assemblies has been recorded in prefrontal cortex^{29, 30}, limbic regions, including entorhinal cortex^{31, 32}, perirhinal cortex^{33, 34}, and lateral amygdala³⁵ of nonhuman primates and rodents (for a review, see work by Wang³⁶).

Parahippocampal cortex

Given the firing patterns observed in parahippocampal regions, their afferent sensory inputs, and extensive reciprocal connections to the hippocampus, activity in parahippocampal cortex is well suited to mediate a contextual representation^{37,38}. Studies examining working memory in humans have identified a dissociation between the maintenance of novel information, supported through MTL function, in contrast to familiar stimuli, supported in part by increased prefrontal activation³⁹. Similar work has identified that successful encoding of novel visual information is supported by the parahippocampal gyrus⁴⁰, which can be reduced through the application of the muscarinic cholinergic antagonist, scopolamine, prior to scanning⁴¹. Loss of cholinergic modulation, and consequently recurrent firing, to entorhinal cortex also produces reduced encoding of novel, but not familiar odors, in rats⁴².

The most compelling evidence for a temporal context representation supported through activity within the MTL comes from recent electrophysiological studies recording from the MTL in humans. Work by Manning and colleagues⁴³ found direct support for contextual reinstatement, evidenced by the reactivation of oscillatory patterns of brain activity recorded in local field potentials, while patients with intractable epilepsy studied a list of nouns and performed free recall. In their analysis, they identified a slowly changing, autocorrelated signal while patients studied specific items- a contextual representation. Critically, just prior to the recall of individual items, the pattern of contextual activity recorded from temporal lobe electrodes, including the hippocampus, returned to a prior state. Recent recordings from within the MTL have also revealed autocorrelated patterns of neuronal firing, which return to a previous state of firing during a continuous recognition task⁴⁴. These studies build on a growing body of evidence from rodent^{45,46}, nonhuman primates⁴⁷, and humans^{48,49} for the encoding and maintenance of temporal context within the MTL.

Dorsolateral prefrontal cortex

The DLPFC has been proposed to act as central locus for a contextual representation^{18,50}, due to its sustained delay period activity in working memory tasks, as well as deficits in source memory⁵¹⁻⁵³, and recency judgments^{54,55} with frontal lesion pathology (although explicit contextual encoding remains intact in frontal amnesics^{56,57}). Furthermore, patients with frontal lobe lesions have been shown to have spared automatic encoding of temporal information,

but deficits in the processing of this information⁵⁸. Functional neuroimaging evidence also points to the role of DLPFC in the selection of action from memory^{59,60} rather than maintenance of information (reviewed by Curtis and D'Esposito⁶¹). Recent evidence from neural decoding of a prospective memory tasks supports the role of prefrontal cortex in modifying the contents of working memory, which could be decoded in posterior cortical regions (but not task-sensitive regions in anterior and lateral prefrontal cortex)⁶². This top-down modulation of contextual representations fits in line with the temporal patterns of firing in electrophysiological recordings of DLPFC⁶³, as well as lesions studies of DLPFC and inferior temporal cortex⁶⁴. This evidence speaks to an interactive role of the DLPFC, with a potential role in the organization of information stored in context.

Recent neuroimaging work investigating the role of DLPFC provides evidence for its role in organizing currently active representations held in working memory⁶⁵. In this study, participants encoded a list of three items, and either performed rote rehearsal or reordered the items according to their weight, during a delay period. This reordering task requires additional manipulation of the contents of working memory, as compared to the rehearsal control task. Consistent with the theorized role of DLPFC in the organization of currently maintained representations, activity in DLPFC was greater for reordering, relative to rehearsal, of items in working memory. Increased activity within DLPFC has also been linked to subsequent relational memory effects^{66,67}, and subsequent clustering⁶⁸ and memory⁶⁹ of studied materials during free recall. These findings support a role for the DLPFC in the encoding of associations between currently active item representations; activity in this region may directly interact with inputs to a contextual representation, allowing individual items presented across longer time scales to be associate to similar contextual states. Future studies investigating how activity within DLPFC during working memory tasks influences item representation in posterior cortical regions may provide further insight into the functional role of the DLPFC.

In addition to supporting relational encoding of items, neuroimaging studies of lateral prefrontal cortex function suggest it may mediate temporal context encoding. As previously mentioned, lesions to prefrontal cortex often cause deficits in recency discrimination^{54,70-72}. Motivated by evidence from the neuropsychological domain, Jenkins and

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Ranganath⁴⁸ examined prefrontal and MTL contributions to the encoding of temporal-order memory. In this study, participants encoded a series of four stimuli, and after an eight second delay, were required to indicate from which position a test probe was encoded in the list. Activity during trials in which subjects correctly assessed the position of the probe was compared to activity during incorrect trials. This contrast of fine temporal memory encoding revealed activity in parahippocampal cortex. In addition to this within-trial measure of temporal context encoding, participants were given a post-scan test in which they indicated the temporal position an item was presented in on a time line representing the duration of the experiment. Activity in rostralateral prefrontal cortex, DLPFC, and anterior hippocampus promoted the encoding of coarse temporal memory. To test if the pattern of activity in any of these regions represented a gradually changing contextual representation, a multivariate pattern analysis of activity in regions showing temporal memory effects was conducted. In this analysis, patterns of activity were constructed in each region of interest, and the similarity of these patterns was compared to adjacent trials. Trials with accurate coarse memory were associated with a dissimilar pattern of activity, relative to neighboring trials. If this pattern of activity in fact represents the encoding of temporal context, the distinct temporal context may facilitate subsequent temporal order judgments. Alternatively, if items are encoded with similar contextual states, it may be more difficult to distinguish when they occurred, relative to other items in the list. These findings, as well as other reports of DLPFC activity supporting subsequent temporal order memory in similar studies⁴⁹ supports the role of prefrontal processes in supporting the encoding of temporal context.

Conclusion

Recent findings suggest that activity in the MTL may mediate a contextual representation, critical for episodic memory. The manner in which information is gated into the medial temporal lobe may be controlled by activity in prefrontal frontal cortex, allowing for control over the current contextual representation. Future studies should emphasize how interactions between prefrontal cortex and the medial temporal lobe shape the current state of context.

References

1. Howard, M. W. and Kahana, M. J. A distributed representation of temporal context. *Journal of Mathematical Psychology* 46, 269–299 (2002).
2. Polyn, S. M., Norman, K. A., and Kahana, M. J. A context maintenance and retrieval model of organizational processes in free recall. *Psychological Review* 116(1), 129–156 (2009).
3. Tulving, E. What is episodic memory? *Current Directions in Psychological Science* 2(3), 67–70, June (1993). ArticleType: research-article / Full publication date: Jun., 1993 / Copyright 1993 Association for Psychological Science.
4. Wheeler, M. E., Petersen, S. E., and Buckner, R. L. Memory's echo: Vivid remembering reactivates Sensory-Specific cortex. *Proceedings of the National Academy of Sciences* 97(20), 11125–11129, September (2000).
5. Nyberg, L., Habib, R., McIntosh, A. R., and Tulving, E. Reactivation of Encoding-Related brain activity during memory retrieval. *Proceedings of the National Academy of Sciences* 97(20), 11120–11124, September (2000).
6. Kahn, I., Davachi, L., and Wagner, A. D. Functional-neuroanatomic correlates of recollection: implications for models of recognition memory. *The Journal of neuroscience: the official journal of the Society for Neuroscience* 24(17), 4172–4180, April (2004). PMID: 15115812.
7. Gottfried, J. A., Smith, A. P. R., Rugg, M. D., and Dolan, R. J. Remembrance of odors past: human olfactory cortex in cross-modal recognition memory. *Neuron* 42(4), 687–695, May (2004). PMID: 15157428.
8. Vaidya, C. J., Zhao, M., Desmond, J. E., and Gabrieli, J. D. E. Evidence for cortical encoding specificity in episodic memory: memory-induced re-activation of picture processing areas. *Neuropsychologia* 40(12), 2136–2143 (2002). PMID: 12208009.
9. Wheeler, M. E. and Buckner, R. L. Functional dissociation among components of remembering: control, perceived oldness, and content. *The Journal of neuroscience: the official journal of the Society for Neuroscience* 23(9), 3869–3880, May (2003). PMID: 12736357.
10. Wheeler, M. E. and Buckner, R. L. Functional-anatomic correlates of remembering and knowing. *NeuroImage* 21(4), 1337–1349, April (2004). PMID: 15050559.
11. Wheeler, M. E., Shulman, G. L., Buckner, R. L., Miezin, F. M., Velanova, K., and Petersen, S. E. Evidence for separate perceptual reactivation and search processes during remembering. *Cerebral cortex (New York, N.Y.: 1991)* 16(7), 949–959, July (2006). PMID: 16162854.
12. Johnson, J. D. and Rugg, M. D. Recollection and the reinstatement of encoding-related cortical activity. *Cerebral cortex (New York, N.Y.: 1991)* 17(11), 2507–2515, November (2007). PMID: 17204822.
13. Epstein, R. and Kanwisher, N. A cortical representation of the local visual environment. *Nature* 392(6676), 598–601, April (1998).
14. Polyn, S. M., Natu, V. S., Cohen, J. D., and Norman, K. A. **Category-Specific cortical activity precedes retrieval during memory search. *Science* 310(5756), 1963–1966, December (2005).**
15. Polyn, S. M., Kragel, J. E., Morton, N. W., McCluey, J. D., and Cohen, Z. D. The neural dynamics of task context in free recall. *Neuropsychologia* (2012).

16. **Gelbard-Sagiv, H., Mukamel, R., Harel, M., Malach, R., and Fried, I. Internally generated reactivation of single neurons in human hippocampus during free recall. *Science* 322(5898), 96–101, October (2008).**
17. Sederberg, P. B., Schulze-Bonhage, A., Madsen, J. R., Bromfield, E. B., Litt, B., Brandt, A., and Kahana, M. J. Gamma oscillations distinguish true from false memories. *Psychological Science* 18(11), 927–932, November (2007).
18. Polyn, S. M. and Kahana, M. J. Memory search and the neural representation of context. *Trends in Cognitive Sciences* 12(1), 24–30, January (2008).
19. Howard, M. W., Fotedar, M. S., Datey, A. V., and Hasselmo, M. E. The temporal context model in spatial navigation and relational learning: Toward a common explanation of medial temporal lobe function across domains. *Psychological Review* 112(1), 75–116 (2005).
20. Davachi, L., Mitchell, J. P., and Wagner, A. D. Multiple routes to memory: Distinct medial temporal lobe processes build item and source memories. *Proceedings of the National Academy of Sciences* 100(4), 2157–2162, February (2003).
21. Ranganath, C., Johnson, M. K., and D’Esposito, M. Prefrontal activity associated with working memory and episodic long-term memory. *Neuropsychologia* 41(3), 378–389 (2003).
22. Kirwan, C. B. and Stark, C. E. L. Medial temporal lobe activation during encoding and retrieval of novel face-name pairs. *Hippocampus* 14(7), 919–930, January (2004).
23. Prince, S. E., Daselaar, S. M., and Cabeza, R. Neural correlates of relational memory: Successful encoding and retrieval of semantic and perceptual associations. *The Journal of Neuroscience* 25(5), 1203–1210, February (2005).
24. Buckner, R. L., Koutstaal, W., Schacter, D. L., Dale, A. M., Rotte, M., and Rosen, B. R. Functional–Anatomic study of episodic retrieval: II. selective averaging of Event-Related fMRI trials to test the retrieval success hypothesis. *NeuroImage* 7(3), 163–175, April (1998).
25. Nyberg, L., McIntosh, A. R., Houle, S., Nilsson, L., and Tulving, E. Activation of medial temporal structures during episodic memory retrieval. , Published online: 25 April 1996; doi:10.1038/380715a0 380(6576), 715–717, April (1996).
26. Konishi, S., Wheeler, M. E., Donaldson, D. I., and Buckner, R. L. Neural correlates of episodic retrieval success. *NeuroImage* 12(3), 276–286, September (2000).
27. Kanwisher, N., McDermott, J., and Chun, M. M. The fusiform face area: A module in human extrastriate cortex specialized for face perception. *The Journal of Neuroscience* 17(11), 4302–4311, June (1997).
28. Malach, R., Reppas, J. B., Benson, R. R., Kwong, K. K., Jiang, H., Kennedy, W. A., Ledden, P. J., Brady, T. J., Rosen, B. R., and Tootell, R. B. Object-Related activity revealed by functional magnetic resonance imaging in human occipital cortex. *Proceedings of the National Academy of Sciences* 92(18), 8135–8139, August (1995).
29. Fuster, J. M. and Alexander, G. E. Neuron activity related to Short-Term memory. *Science* 173(3997), 652–654, August (1971).
30. Romo, R., Brody, C. D., Hernaacuteandez, A., and Lemus, L. Neuronal correlates of parametric working memory in the prefrontal cortex. *Nature* 399(6735), 470–473, June (1999).
31. Egorov, A. V., Hamam, B. N., Franseacuten, E., Hasselmo, M. E., and Alonso, A. A. Graded persistent activity in entorhinal cortex neurons. *Nature* 420(6912), 173–178, November (2002).
32. Yoshida, M., Fransén, E., and Hasselmo, M. E. mGluR-dependent persistent firing in entorhinal cortex layer III neurons. *European Journal of Neuroscience* 28(6), 1116–1126, September (2008).
33. Navaroli, V. L., Zhao, Y., Boguszewski, P., and Brown, T. H. Muscarinic receptor activation enables persistent firing in pyramidal neurons from superficial layers of dorsal perirhinal cortex. *Hippocampus*, September (2011).
34. Kholodar-Smith, D., Boguszewski, P., and Brown, T. Auditory trace fear conditioning requires perirhinal cortex. *Neurobiology of Learning and Memory* 90(3), 537–543, October (2008).
35. Egorov, A. V., Unsicker, K., and Von Bohlen und Halbach, O. Muscarinic control of graded persistent activity in lateral amygdala neurons. *European Journal of Neuroscience* 24(11), 3183–3194, December (2006).
36. Wang, X. Synaptic reverberation underlying mnemonic persistent activity. *Trends in Neurosciences* 24(8), 455–463, August (2001).
37. Burwell, R. D., Witter, M. P., and Amaral, D. G. Perirhinal and postrhinal cortices of the rat: A review of the neuroanatomical literature and comparison with findings from the monkey brain. *Hippocampus* 5(5), 390–408, October (2004).
38. Suzuki, W. A. The anatomy, physiology and functions of the perirhinal cortex. *Current Opinion in Neurobiology* 6(2), 179–186, April (1996).
39. Stern, C. E., Sherman, S. J., Kirchhoff, B. A., and Hasselmo, M. E. Medial temporal and prefrontal contributions to working memory tasks with novel and familiar stimuli. *Hippocampus* 11(4), 337–346, August (2001).
40. Schon, K., Hasselmo, M. E., LoPresti, M. L., Tricarico, M. D., and Stern, C. E. Persistence of parahippocampal representation in the absence of stimulus input enhances Long-Term encoding: A functional magnetic resonance imaging study of subsequent memory after a delayed Match-to-Sample task. *The Journal of Neuroscience* 24(49), 11088–11097, December (2004).
41. Schon, K., Atri, A., Hasselmo, M. E., Tricarico, M. D., LoPresti, M. L., and Stern, C. E. Scopolamine reduces persistent activity related to Long-Term encoding in the parahippocampal gyrus during delayed matching in humans. *The Journal of Neuroscience* 25(40), 9112–9123, October (2005).
42. McGaughy, J., Koene, R. A., Eichenbaum, H., and Hasselmo, M. E. Cholinergic deafferentation of the entorhinal cortex in rats impairs encoding of novel but not familiar stimuli in a delayed Nonmatch-to-Sample task. *The Journal of Neuroscience* 25(44), 10273–10281, November (2005).
43. **Manning, J. R., Polyn, S. M., Baltuch, G. H., Litt, B., and Kahana, M. J. Oscillatory patterns in temporal lobe reveal context reinstatement during memory search. *Proceedings of the National Academy of Sciences* 108(31), 12893–12897, August (2011).**
44. Howard, M. W., Viskontas, I. V., Shankar, K. H., and Fried, I. Ensembles of human MTL neurons “jump back in time” in response to a repeated stimulus. *Hippocampus*, April (2012).
45. Fortin, N. J., Agster, K. L., and Eichenbaum, H. B. Critical role of the hippocampus in memory for sequences of events. *Nature Neuroscience*, March (2002).
46. Manns, J. R., Howard, M. W., and Eichenbaum, H. Gradual

CANDIDATE REVIEWS

- changes in hippocampal activity support remembering the order of events. *Neuron* 56(3), 530–540 (2007).
47. Naya, Y. and Suzuki, W. A. Integrating what and when across the primate medial temporal lobe. *Science* 333(6043), 773–776, August (2011).
48. **Jenkins, L. J. and Ranganath, C. Prefrontal and medial temporal lobe activity at encoding predicts temporal context memory. *The Journal of Neuroscience* 30(46), 15558–15565, November (2010).**
49. Tubridy, S. and Davachi, L. Medial temporal lobe contributions to episodic sequence encoding. *Cerebral Cortex* 21(2), 272–280, February (2011).
50. Braver, T. S., Barch, D. M., Keys, B. A., Carter, C. S., Cohen, J. D., Kaye, J. A., Janowsky, J. S., Taylor, S. F., Yesavage, J. A., Mumenthaler, M. S., Jagust, W. J., and Reed, B. R. Context processing in older adults: Evidence for a theory relating cognitive control to neurobiology in healthy aging. *Journal of Experimental Psychology: General* 130(4), 746–763 (2001).
51. Baldo, J. V., Delis, D., Kramer, J., and Shimamura, A. P. Memory performance on the california verbal learning Test-II: findings from patients with focal frontal lesions. *Journal of the International Neuropsychological Society* 8(04), 539–546 (2002).
52. Janowsky, J. S., Shimamura, A. P., and Squire, L. R. Source memory impairment in patients with frontal lobe lesions. *Neuropsychologia* 27(8), 1043–1056 (1989).
53. Duarte, A., Ranganath, C., and Knight, R. T. Effects of unilateral prefrontal lesions on familiarity, recollection, and source memory. *The Journal of Neuroscience* 25(36), 8333–8337, September (2005).
54. Milner, B., Corsi, P., and Leonard, G. Frontal-lobe contribution to recency judgements. *Neuropsychologia* 29(6), 601–618 (1991).
55. Shimamura, A. P., Janowsky, J. S., and Squire, L. R. Memory for the temporal order of events in patients with frontal lobe lesions and amnesic patients. *Neuropsychologia* 28(8), 803–813 (1990).
56. Thaiss, L. and Petrides, M. Source versus content memory in patients with a unilateral frontal cortex or a temporal lobe excision. *Brain* 126(5), 1112–1126, May (2003).
57. Diana, R. A., Yonelinas, A. P., and Ranganath, C. Medial temporal lobe activity during source retrieval reflects information type, not memory strength. *Journal of Cognitive Neuroscience* 22(8), 1808–1818 (2009).
58. Mangels, J. A. Strategic processing and memory for temporal order in patients with frontal lobe lesions. *Neuropsychology* 11(2), 207–221 (1997).
59. Rowe, J. B., Toni, I., Josephs, O., Frackowiak, R. S. J., and Passingham, R. E. The prefrontal cortex: Response selection or maintenance within working memory? *Science* 288(5471), 1656–1660, June (2000).
60. Pochon, J., Levy, R., Poline, J., Crozier, S., Lehericy, S., Pillon, B., Deweer, B., Le Bihan, D., and Dubois, B. The role of dorsolateral prefrontal cortex in the preparation of forthcoming actions: An fMRI study. *Cerebral Cortex* 11(3), 260–266, March (2001).
61. Curtis, C. E. and D'Esposito, M. Persistent activity in the prefrontal cortex during working memory. *Trends in Cognitive Sciences* 7(9), 415–423, September (2003).
62. Gilbert, S. J. Decoding the content of delayed intentions. *The Journal of Neuroscience* 31(8), 2888–2894, February (2011).
63. Goldman-Rakic, P. Handbook of Physiology, chapter Circuitry of primate prefrontal cortex and regulation of behavior by representational memory, 373–517. American Physiological Society, Washington, DC (1987).
64. Petrides, M. Without title. *Experimental Brain Research* 133(1), 44–54 (2000).
65. Blumenfeld, R. S. and Ranganath, C. Dorsolateral prefrontal cortex promotes Long-Term memory formation through its role in working memory organization. *The Journal of Neuroscience* 26(3), 916–925, January (2006).
66. Blumenfeld, R. S. and Ranganath, C. Prefrontal cortex and Long-Term memory encoding: An integrative review of findings from neuropsychology and neuroimaging. *The Neuroscientist* 13(3), 280–291, June (2007).
67. Murray, L. J. and Ranganath, C. The dorsolateral prefrontal cortex contributes to successful relational memory encoding. *The Journal of Neuroscience* 27(20), 5515–5522, May (2007).
68. Long, N. M., Öztekin, I., and Badre, D. Separable prefrontal cortex contributions to free recall. *The Journal of Neuroscience* 30(33), 10967–10976, August (2010).
69. Staresina, B. P. and Davachi, L. Differential encoding mechanisms for subsequent associative recognition and free recall. *The Journal of Neuroscience* 26(36), 9162–9172, September (2006).
70. McAndrews, M. P. and Milner, B. The frontal cortex and memory for temporal order. *Neuropsychologia* 29(9), 849–859 (1991).
71. Butters, M. A., Kaszniak, A. W., Glisky, E. L., Eslinger, P. J., and Schacter, D. L. Recency discrimination deficits in frontal lobe patients. *Neuropsychology* 8(3), 343–354 (1994).
72. Duarte, A., Henson, R. N., Knight, R. T., Emery, T., and Graham, K. S. Orbito-frontal cortex is necessary for temporal context memory. *Journal of Cognitive Neuroscience* 22(8), 1819–1831 (2009).
73. Ranganath, C., Cohen, M. X., and Brozinsky, C. J. Working memory maintenance contributes to long-term memory formation: neural and behavioral evidence. *Journal of Cognitive Neuroscience* 17, 994–1010 (2005).
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Modeling Gene-Environment Interactions in Parkinson's Disease Using Patient-Derived Induced Pluripotent Stem Cells

Kevin Kumar

Parkinson's disease (PD) is a highly prevalent, progressive neurodegenerative disease characterized by loss of the dopaminergic neurons of the substantia nigra (SN) pars compacta. Although there are several proposed mechanisms for the pathophysiology of this debilitating illness, efforts to develop disease-modifying therapies have been hampered by the inability of existing model systems to completely reproduce the characteristic molecular and pathological features of PD. Given the potential of both environmental toxicants and genetic risk factors to modulate the onset and severity of PD, a model system that accounts for both would serve as a valuable tool in the study of PD-related environmental neurotoxicants. The advent of induced pluripotent stem cell (iPSC) technology has created the opportunity to evaluate personalized, toxicological susceptibility to specific environmental agents. Using this system, it is now possible to analyze cellular physiological pathways in human neurons, both developing and mature, and glial cells that play key roles in handling neurotoxicants. Furthermore, the utilization of living human cells with identical genetic determinants as the resource subjects, with or without PD, is a powerful resource for the development of therapeutics that modulate patient susceptibility to environmental toxicants.

Keywords: *Parkinson's disease, manganese, induced pluripotent stem cells*

Epidemiology of PD

Parkinson's disease (PD) is the second most common neurodegenerative disease. The prevalence of PD in the industrialized world is estimated at 0.3% of the general population and approximately 1% in individuals over the age of 60¹. Thus, PD is considered an age-related disease, with prevalence rising mainly after the age of 50²⁻¹². As the population ages, there is an increasing socioeconomic burden on society². The incidence of PD is 8 to 18 per 100,000 person-years². It has been noted that there is higher prevalence of PD in men than women, a finding hypothesized to be mediated by a neuroprotective role of estrogens^{2,3,6-8}.

PD diagnosis is contingent on presentation with at least two of the four cardinal symptoms: resting tremor, bradykinesia, rigidity, and postural instability². The clinical suspicion is further supported by the patients' respon-

siveness to levodopa, asymmetry of symptoms, or SPECT imaging with DaTSCAN, although the latter is seldom used as a primary diagnostic procedure^{13,14}. Furthermore, secondary causes of parkinsonism, such as drug-induced parkinsonism, must be excluded. Interestingly, the course of PD is highly variable; studies analyzing PD progression suggest that functional deterioration is accelerated both early in the disease course and among patients presenting with postural instability gait difficulty¹⁵⁻¹⁸. The majority (90%) of PD cases are sporadic in etiology, with the remaining 10% of cases having known genetic causes. Furthermore, there is profound heterogeneity in age of onset, neuropathological findings, and characteristic symptoms even among the genetic forms of PD.

Pathophysiology of PD

Although a complete understanding of the

Bradykinesia:

A slowness in the execution of movement. It is one of the three key symptoms of parkinsonism, which are bradykinesia, tremor and rigidity.

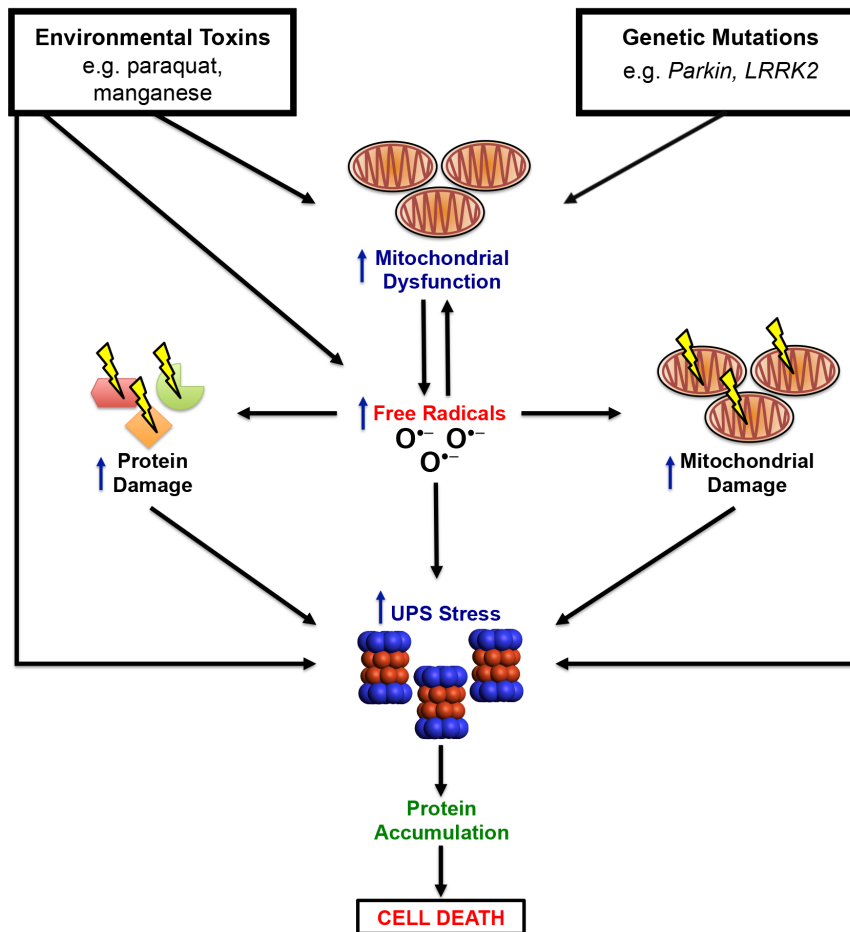


Figure 1: Gene-environment interactions in PD. Environmental and genetic factors result in the mitochondrial dysfunction, generation of free radicals, mitochondrial/protein damage, increased UPS stress, protein aggregation, and dopaminergic cell death in PD.

Ubiquitin proteasome system (UPS):

A multicomponent system that identifies and degrades unwanted proteins in the cytoplasm of all cells; involved in cell growth and differentiation, DNA replication and repair, apoptosis, and stress and immune responses.

pathogenesis of PD remains elusive, current evidence suggests that PD results from a multitude of factors, including: oxidative stress, protein aggregation, and mitochondrial dysfunction¹⁹. PD is characterized by loss of dopaminergic neurons of the substantia nigra and the presence of intraneuronal α -synuclein protein aggregates composed of α -synuclein known as Lewy bodies²⁰. Loss of dopamine levels in the striatum leads to downstream dysregulation of basal ganglia motor circuitry, resulting in the motor symptoms observed in PD. Studies exploring the genetic forms of PD have offered insight regarding central mechanisms in PD pathogenesis. Defective proteins in familial PD result from mutations in genes that function in critical cellular processes, such as the ubiquitin proteasome system (UPS), vesicle trafficking, mitochondrial function, and oxidative stress responses²¹⁻²³. These findings sug-

gest that while there is a common endpoint of decreased striatal dopamine levels, multiple pathways can influence an individual's pattern of neuronal cell death and the mechanism by which it occurs. For example, dysfunction in mitochondrial complex I results in upregulated free radical production causing protein damage. The damaged protein burden increases the stress on the UPS, leading to protein aggregation and subsequent neuronal death²⁴. However, this simple pathway could be influenced at any level by multiple inputs, such as environmental toxins, genetic risk, and enhanced oxidative stress (**Figure 1**). Thus, an individual patient's history of environmental exposure and genetic risk are critical to their clinical manifestation of PD.

Environmental Influences in PD

Epidemiological and laboratory research has revealed a number of environmental exposures and toxicants that contribute to PD. One of the most notable of PD-causing environmental toxicants is 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine (MPTP), a contaminant of a synthetic heroin analogue, which was identified as the culprit in the dramatic onset of parkinsonian symptoms in four individuals after self-administration of the drug²⁵. MPTP administration to both mice and non-human primates revealed a buildup of α -synuclein within nigral dopaminergic cell bodies^{26, 27}. These MPTP studies produced both a valuable model system of parkinsonism and launched a wave of investigation focusing on environmental exposures that contribute to PD.

Environmental Exposures: Pesticides, Heavy Metals, and Beyond

One of the most well-studied environmental associations with PD is that of pesticides and herbicides. Case control studies have revealed an association between exposure to herbicides, insecticides, and farming as an occupation and PD^{28, 29}. Individuals exposed to pesticides have a 70% higher incidence of PD compared to those not exposed³⁰. The timing of exposure has a profound impact on susceptibility; exposure at a younger age increases PD risk relative to those exposed over the age of 60³¹. However, these studies are limited to self-reporting by subjects, a variable that is highly dependent on the awareness of the individual and subject to recall bias. In addition, subjects recruited for these studies would most likely be unable to report prenatal or early-childhood exposures, further limiting the interpretation of epidemiological data. Despite these limitations, several major pesticides have been associated with PD, including: dieldrin, maneb, paraquat and rotenone^{32, 33}. Although it is outside the scope of this article to discuss the mechanism of these toxicants individually, paraquat is representative of how such exposures produce neurodegeneration resulting in parkinsonism, and will be given a brief discussion here.

The toxic properties of paraquat are caused by its redox cycle in which it is reduced by NADH-CYP450 reductase, NADPH-cytochrome *c* reductase, and mitochondrial complex I³⁴⁻³⁷. This process generates a paraquat monocation free radical that is rapidly re-oxidized, producing the superoxide radical (O_2^{2-}). This process sets off a cascade of reactions in which more reactive oxygen species are generated, leading to cellular stress and, when the cellular antioxidant capacity is overwhelmed, eventual degeneration. Furthermore, paraquat is known to generate selective neurodegeneration of SN dopaminergic neurons when administered to rodents in a manner similar to that of MPTP.³⁸ In addition to its acute effects, paraquat exposure during critical developmental periods generates progressive and permanent lesions of the SN dopaminergic system, rendering it hypersusceptible to adult neurotoxicant exposures³⁹.

In addition to pesticides, exposure to heavy metals is associated with PD. In particular, exposure to iron, manganese, lead, copper, zinc, amalgam, and aluminum have each been demonstrated to increase risk of PD⁴⁰. This is of direct clinical relevance, as studies have shown that high manganese exposure produces a secondary form of parkinsonism, known as manganism, that is clinically indistinguishable from idiopathic PD aside from age of onset.⁴¹ The mechanism of neurodegeneration in heavy metal toxicity is hypothesized to be due to deposition of the metals in the SN and increased oxidative stress.⁴⁰

Important to note, not all environmental exposures are neurotoxic, several agents have a demonstrated neuroprotective role such as coffee drinking, smoking, and statin use.^{42, 43} This suggests that environmental impact on PD is bidirectional, and recommendations regarding neuroprotective strategies may be of utility in those at high risk for developing the disease.

Dystonia:

A neurological movement disorder, in which sustained muscle contractions cause twisting and repetitive movements or abnormal postures.

Rasagiline:

An irreversible inhibitor of monoamine oxidase used as a monotherapy in early Parkinson's disease or as an adjunct therapy in more advanced cases.

Genetic Influences in PD

Several loci and genes have been identified as causative of the genetic forms of PD. These include the autosomal dominant *PARK1* and *PARK4* (*SNCA*/α-synuclein), *PARK5* (*UCHL1*), *PARK8* (*LRRK2*), *PARK11* (*GIGYF2*), *PARK13* (*Omi/Htra2*) and the autosomal recessive *PARK2* (*Parkin*), *PARK6* (*PINK1*), *PARK7* (*DJ-1*), and *PARK9* (*ATP13A2*)⁴⁴. These genetic forms vary significantly between each other and are different from the sporadic forms of PD in their age of onset, clinical course, and response to treatment. One example of how a genetic mutation can produce PD is illustrated by *PARK2* loss-of-function mutations, the most common cause of autosomal recessive juvenile parkinsonism (ARJP). In addition to the cardinal symptoms of PD, ARJP is distinguished by prominent lower limb dystonia, severe levodopa-induced dyskinesias, and early age of onset, usually before the age of 40⁴⁵. Notably, patients with ARJP caused by a mutation in *PARK2*, have loss of dopaminergic neurons without the presence of Lewy bodies²⁰. Parkin (the protein product of *PARK2*) functions as an E2-dependent E3 ubiquitin ligase that functions as a substrate-recognition molecule within the UPS⁴⁶. It is hypothesized that loss of Parkin function results in reduced ubiquitination of its substrates and subsequent protein accumulation and toxicity to dopaminergic neurons (**Figure 1**)^{47,48}. There is evidence that Parkin plays a critical role in engulfment of mitochondria with low membrane potential, leading to the hypothesis that failure to eliminate dysfunctional mitochondria contributes to PD pathogenesis.⁴⁹

In addition to these monogenic forms, sporadic forms of PD are likely influenced by an individual's global genetic variation. A number of genome-wide association studies (GWAS) have identified particular PD-associated loci. Susceptibility loci include regions within the monogenic causative genes such as *SNCA* (4q22) and *LRRK2* (12q12), as well as newly identified loci such as *PARK16* (1q32), *BST1* (4p15), and *HLA-DRA*⁵⁰⁻⁵³. Given that

the previously discussed environmental toxicants act on related processes, genetic findings have generated interest in the study of gene-environment interactions that may underlie the heterogeneity in presentation among PD patients.

Gene-environment interactions in PD – Modeling neurotoxicological risk

Examination of neurotoxicological risk in PD model systems has emerged as an area of active research given the strong evidence for the influence of both environmental and genetic factors on PD onset. The concept of mutations in individual genes or single nucleotide polymorphisms across multiple loci that alter the susceptibility of an individual to a given toxicant has been validated in many studies. For example, mutations in the gene *MDR1* predispose individuals to the injurious effects of pesticides and other P-glycoprotein transported xenobiotics, resulting in PD⁵⁴. Similarly, polymorphisms in metabolic enzymes such as *MAOB*, *CYP2D6*, and *GSTT1* have been associated with PD⁵⁵. On the other hand, experiments in yeast and animal model systems have revealed a protective role of certain genes, such as *PARK9* (*ATP13A2*) that protects against manganese toxicity and dopaminergic cell death due to α-synuclein overexpression⁵⁶. Despite these advances, there is a knowledge gap between clinical data from PD patients and laboratory data generated using model systems. It is assumed, however, that by modulating individual response to neurotoxicants, the clinical course of PD can be manipulated. This concept has been demonstrated in clinical trials demonstrating the beneficial effects of levodopa, tai chi, and rasagiline on PD symptoms and progression.⁵⁷⁻⁵⁹ Thus, the need to test different interactions between subject-specific genetic background and environmental exposures makes patient-specific iPSCs a powerful tool to predict clinical outcomes and guide clinical investigations and intervention.

The utility of iPSC technology for neurotoxicology

Background of iPSCs

In 2007, Takahashi *et al.* reported for the first time the possibility of reprogramming adult human-derived fibroblasts to pluripotent stem cells using four defined transcription factors, *OCT4*, *SOX2*, *c-MYC*, and *KLf4*⁶⁰. iPSCs exhibit the typical characteristic of inner cell mass-derived human embryonic stem cells, including self-renewal and the ability to differentiate into cell types of all three germ layers. This landmark study launched a new field of research focused on improving the efficiency of reprogramming and deriving cells from various patient types. Initial experiments utilized retroviruses for transduction, which introduced a set of drawbacks including mutagenesis at insertion sites and persistent expression of reprogramming factors. In efforts to overcome these obstacles, several alternative reprogramming strategies have been developed, including doxycycline-inducible expression, the use of *loxP* sites, *PiggyBac* transposons, adenovirus transduction, plasmid transfections, and episomal vectors⁶¹⁻⁶⁷. In addition, other groups are investigating the use of compounds that permit iPSC induction without the use of genetic material⁶⁸. These improvements in reprogramming enhance the utility of this system for the study of gene-environment interactions, as they minimize the contribution of the reprogramming process to genetic heterogeneity among iPSCs derived from different individuals. After reprogramming patient fibroblasts to iPSCs, the cells can be differentiated into a variety of neuronal and glial subtypes including functional midbrain dopaminergic neurons^{69,70}.

iPSCs as a model for gene-environment interactions in PD

One of the major advantages of utilizing patient-specific iPSCs to study neurotoxicological interaction is that an individual may be evaluated for environmental risk without *a priori* knowledge of their genetic risk factors⁷¹. The use of iPSCs for toxicological risk assessment is dependent on the assumption that cells derived from patients serve as a model system for understanding the influence of human genetic factors and their ability to modulate the vulnerability of differentiated cells to a given toxicant⁷¹. Although efforts to validate these assumptions are underway, iPSC technology remains an exciting opportunity to examine changes in the development and maintenance of neuronal function after genetic and toxicant perturbation.

There are a variety of exposure paradigms where iPSC-derived neurons and neural progenitors are of value. Through this experimental system, environmental insults or protectants can be screened across different temporal deliv-

ery patterns to understand response to acute and chronic exposures. The pattern of exposure is of interest, since toxicants such as methylmercury exhibit acute and latent effects with variable sensitivity based on developmental time point⁷²⁻⁷⁴. *In vitro* neuronal differentiation of hiPSCs permits assessment of interactions between early exposure and subsequent risk of neurodegenerative phenotypes in acute, multi-hit, and chronic exposure paradigms. Prior to iPSCs, such studies could only be performed utilizing primary cell culture or embryonic stem cells, which are high-cost alternatives that lack patient specificity.

Furthermore, developing neural progenitors can be exposed to chronic low concentrations of the agent to mimic the effect of cumulative toxicity across the lifetime of a neuron. The pluripotent nature of iPSCs allows the assessment of a diverse set of neuronal subtypes to a given exposure. For example, one could investigate if midbrain dopaminergic neurons have a heightened susceptibility to manganese during development compared to forebrain dopaminergic neurons derived from the same patient. Alternatively, polymorphisms at different loci between patients could heighten developmental sensitivity to a toxicant between PD patients and controls. Such findings from iPSCs can inform downstream *in vivo* vertebrate studies that account for endogenous processes such as detoxification, neuronal regeneration, and immune response.

Current challenges in modeling PD with iPSCs

Perhaps the greatest challenge utilizing iPSCs is assuring the coherence of genotype and phenotype. Many groups have identified methylation pattern and gene expression differences between iPSC lines from the same patient⁷⁵⁻⁷⁸. This could be due to a multitude of factors, including expression of reprogramming vectors, point mutations, and copy number variants generated in the reprogramming process⁷⁹. Any induced genetic or epigenetic abnormalities are of concern in the study of gene-environment interactions in PD since they may mask the effect of a patient's individual genetic variation. In order to account for these effects, karyotyping should be performed at minimum, and whole genome and bisulphite sequencing should also be considered. Furthermore, it should be noted that *in vitro* studies utilizing iPSC-derived cells are limited in interpretation because they lack complex extracellular environments, neuronal architecture, and glial interactions. However, there have been efforts to address these concerns through development of mixed neuronal-glial cultures, but results have been inconsistent thus far⁸⁰. Finally, since PD

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is an age-related disease, newly differentiated neurons from iPSCs fail to simulate long-term simulate age-related phenotypes.

The promise of PD patient-derived iPSCs for personalized medicine and risk assessment

PD has the potential to benefit tremendously from the wide utilization of iPSC technology. This system permits the study of gene-environment interactions utilizing cellular subtypes derived from patients afflicted with PD. iPSC technology provides a critical link between epidemiological studies and animal, cellular, and computational models. This feature has potential for direct clinical application, as iPSC-derived neurons can be used to design customized neuroprotective strategies and recommendations for patients at the preclinical or early clinical stages of PD. In a parallel fashion, drug development can be accelerated by the development of high-throughput assays utilizing iPSC-derived neurons. Although PD-derived iPSCs share the limitations of other *in vitro* model systems, the fact that they are derived from patients with a clinical diagnosis offers the ability to explore processes such as oxidative stress, protein aggregation, and mitochondrial dysfunction in response to toxicants without a full understanding of the genetic factors underlying PD pathogenesis.

References

1. Nussbaum, R.L. & Ellis, C.E. Alzheimer's disease and Parkinson's disease. *N Engl J Med* **348**, 1356-64 (2003).
2. de Lau, L.M. & Breteler, M.M. Epidemiology of Parkinson's disease. *Lancet Neurol* **5**, 525-35 (2006).
3. de Rijk, M.C., Breteler, M.M., Graveland, G.A., Ott, A., Grobbee, D.E., van der Meche, F.G. & Hofman, A. Prevalence of Parkinson's disease in the elderly: the Rotterdam Study. *Neurology* **45**, 2143-6 (1995).
4. Benito-Leon, J., Bermejo-Pareja, F., Morales, J.M., Vega, S. & Molina, J.A. Prevalence of essential tremor in three elderly populations of central Spain. *Mov Disord* **18**, 389-94 (2003).
5. Schoenberg, B.S., Osuntokun, B.O., Adeuja, A.O., Bademosi, O., Nottidge, V., Anderson, D.W. & Haerer, A.F. Comparison of the prevalence of Parkinson's disease in black populations in the rural United States and in rural Nigeria: door-to-door community studies. *Neurology* **38**, 645-6 (1988).
6. Tison, F., Dartigues, J.F., Dubes, L., Zuber, M., Alperovitch, A. & Henry, P. Prevalence of Parkinson's disease in the elderly: a population study in Gironde, France. *Acta Neurol Scand* **90**, 111-5 (1994).
7. Morgante, L., Rocca, W.A., Di Rosa, A.E., De Domenico, P., Grigoletto, F., Meneghini, F., Reggio, A., Savettieri, G., Castiglione, M.G., Patti, F. & et al. Prevalence of Parkinson's disease and other types of parkinsonism: a door-to-door survey in three Sicilian municipalities. The Sicilian Neuro-Epidemiologic Study (SNES) Group. *Neurology* **42**, 1901-7 (1992).
8. de Rijk, M.C., Tzourio, C., Breteler, M.M., Dartigues, J.F., Amaducci, L., Lopez-Pousa, S., Manubens-Bertran, J.M., Alperovitch, A. & Rocca, W.A. Prevalence of parkinsonism and Parkinson's disease in Europe: the EUROPARKINSON Collaborative Study. European Community Concerted Action on the Epidemiology of Parkinson's disease. *J Neurol Neurosurg Psychiatry* **62**, 10-5 (1997).
9. Claveria, L.E., Duarte, J., Sevillano, M.D., Perez-Sempere, A., Cabezas, C., Rodriguez, F. & de Pedro-Cuesta, J. Prevalence of Parkinson's disease in Cantalejo, Spain: a door-to-door survey. *Mov Disord* **17**, 242-9 (2002).
10. Mayeux, R., Marder, K., Cote, L.J., Denaro, J., Hemenegildo, N., Mejia, H., Tang, M.X., Lantigua, R., Wilder, D., Gurland, B. & et al. The frequency of idiopathic Parkinson's disease by age, ethnic group, and sex in northern Manhattan, 1988-1993. *Am J Epidemiol* **142**, 820-7 (1995).
11. Li, S.C., Schoenberg, B.S., Wang, C.C., Cheng, X.M., Rui, D.Y., Bolis, C.L. & Schoenberg, D.G. A prevalence survey of Parkinson's disease and other movement disorders in the People's Republic of China. *Arch Neurol* **42**, 655-7 (1985).
12. Fall, P.A., Axelson, O., Fredriksson, M., Hansson, G., Lindvall, B., Olsson, J.E. & Granerus, A.K. Age-standardized incidence and prevalence of Parkinson's disease in a Swedish community. *J Clin Epidemiol* **49**, 637-41 (1996).
13. Litvan, I., Bhatia, K.P., Burn, D.J., Goetz, C.G., Lang, A.E., McKeith, I., Quinn, N., Sethi, K.D., Shults, C. & Wenning, G.K. Movement Disorders Society Scientific Issues Committee report: SIC Task Force appraisal of clinical diagnostic criteria for Parkinsonian disorders. *Mov Disord* **18**, 467-86 (2003).
14. Tolosa, E., Borght, T.V. & Moreno, E. Accuracy of DaTSCAN (123I-Ioflupane) SPECT in diagnosis of patients with clinically uncertain parkinsonism: 2-year follow-up of an open-label study. *Mov Disord* **22**, 2346-51 (2007).
15. Jankovic, J. Parkinson's disease: clinical features and diagnosis. *J Neurol Neurosurg Psychiatry* **79**, 368-76 (2008).
16. Jankovic, J. & Kapadia, A.S. Functional decline in Parkinson disease. *Arch Neurol* **58**, 1611-5 (2001).
17. Lang, A.E. The progression of Parkinson disease: a hypothesis. *Neurology* **68**, 948-52 (2007).
18. Post, B., Merkus, M.P., de Haan, R.J. & Speelman, J.D. Prognostic factors for the progression of Parkinson's disease: a systematic review. *Mov Disord* **22**, 1839-51; quiz 1988 (2007).
19. Greenamyre, J.T. & Hastings, T.G. Biomedicine. Parkinson's--divergent causes, convergent mechanisms. *Science* **304**, 1120-2 (2004).
20. Wood-Kaczmar, A., Gandhi, S. & Wood, N.W. Understanding the molecular causes of Parkinson's disease. *Trends Mol Med* **12**, 521-8 (2006).
21. Jakes, R., Spillantini, M.G. & Goedert, M. Identification of

- two distinct synucleins from human brain. *FEBS Lett* **345**, 27-32 (1994).
22. Mori, F., Tanji, K., Yoshimoto, M., Takahashi, H. & Wakabayashi, K. Demonstration of alpha-synuclein immunoreactivity in neuronal and glial cytoplasm in normal human brain tissue using proteinase K and formic acid pretreatment. *Exp Neurol* **176**, 98-104 (2002).
 23. Nuytemans, K., Theuns, J., Cruts, M. & Van Broeckhoven, C. Genetic etiology of Parkinson disease associated with mutations in the SNCA, PARK2, PINK1, PARK7, and LRRK2 genes: a mutation update. *Hum Mutat* **31**, 763-80 (2010).
 24. Ali, S.F., Binienda, Z.K. & Imam, S.Z. Molecular aspects of dopaminergic neurodegeneration: gene-environment interaction in parkin dysfunction. *Int J Environ Res Public Health* **8**, 4702-13 (2011).
 25. Langston, J.W., Ballard, P., Tetrud, J.W. & Irwin, I. Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science* **219**, 979-80 (1983).
 26. Vila, M., Vukosavic, S., Jackson-Lewis, V., Neystat, M., Jakowec, M. & Przedborski, S. Alpha-synuclein up-regulation in substantia nigra dopaminergic neurons following administration of the parkinsonian toxin MPTP. *J Neurochem* **74**, 721-9 (2000).
 27. McCormack, A.L., Mak, S.K., Shenasa, M., Langston, W.J., Forno, L.S. & Di Monte, D.A. Pathologic modifications of alpha-synuclein in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated squirrel monkeys. *J Neuropathol Exp Neurol* **67**, 793-802 (2008).
 28. Gorell, J.M., Johnson, C.C., Rybicki, B.A., Peterson, E.L. & Richardson, R.J. The risk of Parkinson's disease with exposure to pesticides, farming, well water, and rural living. *Neurology* **50**, 1346-50 (1998).
 29. Ritz, B. & Yu, F. Parkinson's disease mortality and pesticide exposure in California 1984-1994. *Int J Epidemiol* **29**, 323-9 (2000).
 30. Ascherio, A., Chen, H., Weisskopf, M.G., O'Reilly, E., McCullough, M.L., Calle, E.E., Schwarzschild, M.A. & Thun, M.J. Pesticide exposure and risk for Parkinson's disease. *Ann Neurol* **60**, 197-203 (2006).
 31. Costello, S., Cockburn, M., Bronstein, J., Zhang, X. & Ritz, B. Parkinson's disease and residential exposure to maneb and paraquat from agricultural applications in the central valley of California. *Am J Epidemiol* **169**, 919-26 (2009).
 32. Betarbet, R., Sherer, T.B., MacKenzie, G., Garcia-Osuna, M., Panov, A.V. & Greenamyre, J.T. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nat Neurosci* **3**, 1301-6 (2000).
 33. Kamel, F., Tanner, C., Umbach, D., Hoppin, J., Alavanja, M., Blair, A., Comyns, K., Goldman, S., Korell, M., Langston, J., Ross, G. & Sandler, D. Pesticide exposure and self-reported Parkinson's disease in the agricultural health study. *Am J Epidemiol* **165**, 364-74 (2007).
 34. Dinis-Oliveira, R.J., Remiao, F., Carmo, H., Duarte, J.A., Navarro, A.S., Bastos, M.L. & Carvalho, F. Paraquat exposure as an etiological factor of Parkinson's disease. *Neurotoxicology* **27**, 1110-22 (2006).
 35. Clejan, L. & Cederbaum, A.I. Synergistic interactions between NADPH-cytochrome P-450 reductase, paraquat, and iron in the generation of active oxygen radicals. *Biochem Pharmacol* **38**, 1779-86 (1989).
 36. Fernandez, Y., Subirade, I., Anglade, F., Periquet, A. & Mitjavila, S. Microsomal membrane peroxidation by an Fe³⁺/paraquat system. Consequences of phenobarbital induction. *Biol Trace Elem Res* **47**, 9-15 (1995).
 37. Fukushima, T., Yamada, K., Isobe, A., Shiwaku, K. & Yamane, Y. Mechanism of cytotoxicity of paraquat. I. NADH oxidation and paraquat radical formation via complex I. *Exp Toxicol Pathol* **45**, 345-9 (1993).
 38. McCormack, A.L., Thiruchelvam, M., Manning-Bog, A.B., Thiffault, C., Langston, J.W., Cory-Slechta, D.A. & Di Monte, D.A. Environmental risk factors and Parkinson's disease: selective degeneration of nigral dopaminergic neurons caused by the herbicide paraquat. *Neurobiol Dis* **10**, 119-27 (2002).
 39. Thiruchelvam, M., Richfield, E.K., Goodman, B.M., Baggs, R.B. & Cory-Slechta, D.A. Developmental exposure to the pesticides paraquat and maneb and the Parkinson's disease phenotype. *Neurotoxicology* **23**, 621-33 (2002).
 40. Lai, B.C., Marion, S.A., Teschke, K. & Tsui, J.K. Occupational and environmental risk factors for Parkinson's disease. *Parkinsonism Relat Disord* **8**, 297-309 (2002).
 41. Racette, B.A., McGee-Minnich, L., Moerlein, S.M., Mink, J.W., Videen, T.O. & Perlmutter, J.S. Welding-related parkinsonism: clinical features, treatment, and pathophysiology. *Neurology* **56**, 8-13 (2001).
 42. Hernan, M.A., Takkouche, B., Caamano-Isorna, F. & Gestal-Otero, J.J. A meta-analysis of coffee drinking, cigarette smoking, and the risk of Parkinson's disease. *Ann Neurol* **52**, 276-84 (2002).
 43. Gao, X., Simon, K.C., Schwarzschild, M.A. & Ascherio, A. Prospective study of statin use and risk of Parkinson disease. *Arch Neurol* **69**, 380-4 (2012).
 44. Lesage, S. & Brice, A. Parkinson's disease: from monogenic forms to genetic susceptibility factors. *Hum Mol Genet* **18**, R48-59 (2009).
 45. Lucking, C.B., Abbas, N., Durr, A., Bonifati, V., Bonnet, A.M., de Broucker, T., De Michele, G., Wood, N.W., Agid, Y. & Brice, A. Homozygous deletions in parkin gene in European and North African families with autosomal recessive juvenile parkinsonism. The European Consortium on Genetic Susceptibility in Parkinson's Disease and the French Parkinson's Disease Genetics Study Group. *Lancet* **352**, 1355-6 (1998).
 46. Shimura, H., Hattori, N., Kubo, S., Mizuno, Y., Asakawa, S., Minoshima, S., Shimizu, N., Iwai, K., Chiba, T., Tanaka, K. & Suzuki, T. Familial Parkinson disease gene product, parkin, is a ubiquitin-protein ligase. *Nat Genet* **25**, 302-5 (2000).
 47. Sriram, S.R., Li, X., Ko, H.S., Chung, K.K., Wong, E., Lim, K.L., Dawson, V.L. & Dawson, T.M. Familial-associated mutations differentially disrupt the solubility, localization, binding and ubiquitination properties of parkin. *Hum Mol Genet* **14**, 2571-86 (2005).
 48. von Coelln, R., Dawson, V.L. & Dawson, T.M. Parkin-associated Parkinson's disease. *Cell Tissue Res* **318**, 175-84 (2004).
 49. **Narendra, D., Tanaka, A., Suen, D.F. & Youle, R.J. Parkin is recruited selectively to impaired mitochondria and promotes**

their autophagy. *J Cell Biol* **183**, 795-803 (2008).

This study helps establish the function of PARK2 (Parkin) in the elimination of impaired mitochondria. The authors conclude that failure of Parkin to promote autophagy of damaged mitochondria as a factor in the pathogenesis of PD. This article utilizes a number of molecular biology approaches to explore mitochondrial phenotypes in response to environmental and genetic insults. This is an example of study examining specific gene-environment interactions in PD.

50. Satake, W., Nakabayashi, Y., Mizuta, I., Hirota, Y., Ito, C., Kubo, M., Kawaguchi, T., Tsunoda, T., Watanabe, M., Takeda, A., Tomiyama, H., Nakashima, K., Hasegawa, K., Obata, F., Yoshikawa, T., Kawakami, H., Sakoda, S., Yamamoto, M., Hattori, N., Murata, M., Nakamura, Y. & Toda, T. Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. *Nat Genet* **41**, 1303-7 (2009).
51. Simon-Sanchez, J., Schulte, C., Bras, J.M., Sharma, M., Gibbs, J.R., Berg, D., Paisan-Ruiz, C., Lichtner, P., Scholz, S.W., Hernandez, D.G., Kruger, R., Federoff, M., Klein, C., Goate, A., Perlmutter, J., Bonin, M., Nalls, M.A., Illig, T., Gieger, C., Houlden, H., Steffens, M., Okun, M.S., Racette, B.A., Cookson, M.R., Foote, K.D., Fernandez, H.H., Traynor, B.J., Schreiber, S., Arepalli, S., Zonozi, R., Gwinn, K., van der Brug, M., Lopez, G., Chanock, S.J., Schatzkin, A., Park, Y., Hollenbeck, A., Gao, J., Huang, X., Wood, N.W., Lorenz, D., Deuschl, G., Chen, H., Riess, O., Hardy, J.A., Singleton, A.B. & Gasser, T. Genome-wide association study reveals genetic risk underlying Parkinson's disease. *Nat Genet* **41**, 1308-12 (2009).
- This article is recent work, within a population of European ancestry, suggesting the strong role of genetic variations in the etiology of Parkinson's Disease. The identification of the association between PD and MAPT was not found in a parallel study in the Japanese population suggesting that risk loci can be masked in analyses across populations. Most importantly, the study affirms the role of common genetic variants in contributing to PD risk in sporadic PD.*
52. Hamza, T.H., Zabetian, C.P., Tenesa, A., Laederach, A., Montimurro, J., Yearout, D., Kay, D.M., Doheny, K.F., Paschall, J., Pugh, E., Kusel, V.I., Collura, R., Roberts, J., Griffith, A., Samii, A., Scott, W.K., Nutt, J., Factor, S.A. & Payami, H. Common genetic variation in the HLA region is associated with late-onset sporadic Parkinson's disease. *Nat Genet* **42**, 781-5 (2010).
53. Nichols, W.C., Pankratz, N., Hernandez, D., Paisan-Ruiz, C., Jain, S., Halter, C.A., Michaels, V.E., Reed, T., Rudolph, A., Shults, C.W., Singleton, A. & Foroud, T. Genetic screening for a single common LRRK2 mutation in familial Parkinson's disease. *Lancet* **365**, 410-2 (2005).
54. Drozdzik, M., Bialecka, M., Mysliwiec, K., Honczarenko, K., Stankiewicz, J. & Sych, Z. Polymorphism in the P-glycoprotein drug transporter MDR1 gene: a possible link between environmental and genetic factors in Parkinson's disease. *Pharmacogenetics* **13**, 259-63 (2003).
55. Tan, E.K., Khajavi, M., Thornby, J.I., Nagamitsu, S., Jankovic, J. & Ashizawa, T. Variability and validity of polymorphism association studies in Parkinson's disease. *Neurology* **55**, 533-8 (2000).
56. Gitler, A.D., Chesi, A., Geddie, M.L., Strathearn, K.E., Hamamichi, S., Hill, K.J., Caldwell, K.A., Caldwell, G.A., Cooper, A.A., Rochet, J.C. & Lindquist, S. Alpha-synuclein is part of a diverse and highly conserved interaction network that includes PARK9 and manganese toxicity. *Nat Genet* **41**, 308-15 (2009).
57. Fahn, S., Oakes, D., Shoulson, I., Kieburtz, K., Rudolph, A., Lang, A., Olanow, C.W., Tanner, C. & Marek, K. Levodopa and the progression of Parkinson's disease. *N Engl J Med* **351**, 2498-508 (2004).
58. Li, F., Harmer, P., Fitzgerald, K., Eckstrom, E., Stock, R., Galver, J., Maddalozzo, G. & Batya, S.S. Tai chi and postural stability in patients with Parkinson's disease. *N Engl J Med* **366**, 511-9 (2012).
59. Olanow, C.W., Rascol, O., Hauser, R., Feigin, P.D., Jankovic, J., Lang, A., Langston, W., Melamed, E., Poewe, W., Stocchi, F. & Tolosa, E. A double-blind, delayed-start trial of rasagiline in Parkinson's disease. *N Engl J Med* **361**, 1268-78 (2009).
- This article is a major double-blind, delayed-start trial assessing rasagiline as a disease modifying therapy in PD. The authors establish the treatment of rasagiline at 1mg/day has a possible disease modifying effect. This study is an important example of how identification of compounds that act on protein products of genes with polymorphisms associated with PD have potential for clinical utility.*
60. Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K. & Yamanaka, S. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* **131**, 861-72 (2007).
61. Brambrink, T., Foreman, R., Welstead, G.G., Lengner, C.J., Wernig, M., Suh, H. & Jaenisch, R. Sequential expression of pluripotency markers during direct reprogramming of mouse somatic cells. *Cell Stem Cell* **2**, 151-9 (2008).
62. Yusa, K., Rad, R., Takeda, J. & Bradley, A. Generation of transgene-free induced pluripotent mouse stem cells by the piggyBac transposon. *Nat Methods* **6**, 363-9 (2009).
63. Okita, K., Nakagawa, M., Hyenjong, H., Ichisaka, T. & Yamanaka, S. Generation of mouse induced pluripotent stem cells without viral vectors. *Science* **322**, 949-53 (2008).
- This article establishes the episomal method of reprogramming of fibroblasts into induced pluripotent stem cells (iPSCs). This is the current method we utilize in the Bowman laboratory. Using this approach, we limit the risk of integration of the reprogramming vectors into the genome. By preventing integration, phenotypes observed with iPSC-derived neurons are more likely related to the genetic background of the patient than the reprogramming process itself.*
64. Stadtfeld, M., Nagaya, M., Utikal, J., Weir, G. & Hochedlinger, K. Induced pluripotent stem cells generated without viral integration. *Science* **322**, 945-9 (2008).
65. Zhou, W. & Freed, C.R. Adenoviral gene delivery can reprogram human fibroblasts to induced pluripotent stem cells. *Stem Cells* **27**, 2667-74 (2009).
66. Yu, J., Hu, K., Smuga-Otto, K., Tian, S., Stewart, R., Slukvin, II & Thomson, J.A. Human induced pluripotent stem cells free of vector and transgene sequences. *Science* **324**, 797-801 (2009).
67. Okita, K., Matsumura, Y., Sato, Y., Okada, A., Morizane, A., Okamoto, S., Hong, H., Nakagawa, M., Tanabe, K., Tezuka, K., Shibata, T., Kunisada, T., Takahashi, M., Takahashi, J., Saji, H. & Yamanaka, S. A more efficient method to generate integration-free human iPS cells. *Nat Methods* **8**, 409-12 (2011).
68. Lin, T., Ambasadhan, R., Yuan, X., Li, W., Hilcove, S., Abujarour, R., Lin, X., Hahm, H.S., Hao, E., Hayek, A. & Ding, S. A chemical platform for improved induction of human iPSCs. *Nat Methods* **6**, 805-8 (2009).

69. Swistowski, A., Peng, J., Liu, Q., Mali, P., Rao, M.S., Cheng, L. & Zeng, X. Efficient generation of functional dopaminergic neurons from human induced pluripotent stem cells under defined conditions. *Stem Cells* **28**, 1893-904 (2010).
70. Cho, M.S., Lee, Y.E., Kim, J.Y., Chung, S., Cho, Y.H., Kim, D.S., Kang, S.M., Lee, H., Kim, M.H., Kim, J.H., Leem, J.W., Oh, S.K., Choi, Y.M., Hwang, D.Y., Chang, J.W. & Kim, D.W. Highly efficient and large-scale generation of functional dopamine neurons from human embryonic stem cells. *Proc Natl Acad Sci U S A* **105**, 3392-7 (2008).
71. Kumar, K.K., Aboud, A.A. & Bowman, A.B. The potential of induced pluripotent stem cells as a translational model for neurotoxicological risk. *Neurotoxicology* **33**, 518-29 (2012).
72. Farina, M., Rocha, J.B. & Aschner, M. Mechanisms of methylmercury-induced neurotoxicity: evidence from experimental studies. *Life Sci* **89**, 555-63 (2011).
73. Ni, M., Li, X., Yin, Z., Sidoryk-Wegrzynowicz, M., Jiang, H., Farina, M., Rocha, J.B., Syversen, T. & Aschner, M. Comparative study on the response of rat primary astrocytes and microglia to methylmercury toxicity. *Glia* **59**, 810-20 (2011).
74. Weiss, B., Clarkson, T.W. & Simon, W. Silent latency periods in methylmercury poisoning and in neurodegenerative disease. *Environ Health Perspect* **110 Suppl 5**, 851-4 (2002).
75. Boulting, G.L., Kiskinis, E., Croft, G.F., Amoroso, M.W., Oakley, D.H., Wainger, B.J., Williams, D.J., Kahler, D.J., Yamaki, M., Davidow, L., Rodolfa, C.T., Dimos, J.T., Mikkilineni, S., MacDermott, A.B., Woolf, C.J., Henderson, C.E., Wichterle, H. & Eggan, K. A functionally characterized test set of human induced pluripotent stem cells. *Nat Biotechnol* **29**, 279-86 (2011).
76. Chin, M.H., Mason, M.J., Xie, W., Volinia, S., Singer, M., Peterson, C., Ambartsumyan, G., Aimiwu, O., Richter, L., Zhang, J., Khvorostov, I., Ott, V., Grunstein, M., Lavon, N., Benvenisty, N., Croce, C.M., Clark, A.T., Baxter, T., Pyle, A.D., Teitell, M.A., Pelegrini, M., Plath, K. & Lowry, W.E. Induced pluripotent stem cells and embryonic stem cells are distinguished by gene expression signatures. *Cell Stem Cell* **5**, 111-23 (2009).
77. Doi, A., Park, I.H., Wen, B., Murakami, P., Aryee, M.J., Irizarry, R., Herb, B., Ladd-Acosta, C., Rho, J., Loewer, S., Miller, J., Schlaeger, T., Daley, G.Q. & Feinberg, A.P. Differential methylation of tissue- and cancer-specific CpG island shores distinguishes human induced pluripotent stem cells, embryonic stem cells and fibroblasts. *Nat Genet* **41**, 1350-3 (2009).
78. Stadtfeld, M., Apostolou, E., Akutsu, H., Fukuda, A., Follett, P., Natesan, S., Kono, T., Shioda, T. & Hochedlinger, K. Aberrant silencing of imprinted genes on chromosome 12qF1 in mouse induced pluripotent stem cells. *Nature* **465**, 175-81 (2010).
79. Gore, A., Li, Z., Fung, H.L., Young, J.E., Agarwal, S., Antosiewicz-Bourget, J., Canto, I., Giorgetti, A., Israel, M.A., Kiskinis, E., Lee, J.H., Loh, Y.H., Manos, P.D., Montserrat, N., Panopoulos, A.D., Ruiz, S., Wilbert, M.L., Yu, J., Kirkness, E.F., Izpisua Belmonte, J.C., Rossi, D.J., Thomson, J.A., Eggan, K., Daley, G.Q., Goldstein, L.S. & Zhang, K. Somatic coding mutations in human induced pluripotent stem cells. *Nature* **471**, 63-7 (2011).
80. Haidet-Phillips, A.M., Hester, M.E., Miranda, C.J., Meyer, K., Braun, L., Frakes, A., Song, S., Likhite, S., Murtha, M.J., Foust, K.D., Rao, M., Eagle, A., Kammesheidt, A., Christensen, A., Mendell, J.R., Burghes, A.H. & Kaspar, B.K. Astrocytes from familial and sporadic ALS patients are toxic to motor neurons. *Nat Biotechnol* **29**, 824-8 (2011).

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Circuit Refinement in the Developing Nervous System: Uncovering the Molecular Mechanisms that Destabilize the Synapse

Tyne Miller

Developing neural circuits undergo extensive refinement, characterized by the dynamic addition and removal of synapses. Localization of synaptic connections is critical for circuit architecture and information flow. Incorrect or excess synapses are eliminated to produce the precise cellular connections that are characteristic of mature nervous systems. This process has been observed across phylogeny, suggesting that the underlying mechanisms may be evolutionarily conserved. Studies of the vertebrate neuromuscular junction and visual circuitry have defined some overall themes in synapse refinement; the process is modulated by circuit activity and is commonly characterized by competition between inputs. The molecular networks that connect circuit activity to refinement are largely unknown, suggesting the need for a simpler model system. This review will examine the current understanding of synaptic refinement and introduce *C. elegans* as a model system to examine the molecular underpinnings of this complex, conserved process.

Keywords: *Synapse refinement, synaptic pruning, input elimination, synaptic remodeling, activity dependence, C. elegans, synapse disassembly, ubiquitination*

Critical period:

Time interval during the development of an organism characterized by increased plasticity in the nervous system and an increased sensitivity to stimuli.

Heterochronic gene:

Gene that controls the timing of development.

Introduction

During development, nervous systems create many more synapses than will be maintained at maturity¹. It is unclear why this happens, but reduction involves large-scale removal of redundant synapses. The elimination of functional synapses is a hallmark of circuit refinement²⁻⁴. Studies of the neuromuscular junction (NMJ) have demonstrated that functional synapses are eliminated during circuit refinement²⁻⁵. Although it seems likely that functional synapses are also dismantled in the central nervous system, clear evidence of this phenomenon has been difficult to acquire²⁻⁵. Studies at the cerebellum show that incomplete elimination results in coordination defects in mice, suggesting that synapse elimination is important in creating functional neural circuits⁶⁻⁸. Synaptic refinement ranges from the disassembly of individual synapses to the complete removal of all connections between a presynaptic cell and its postsynaptic target^{4,6}. The refinement of neural circuits has been observed in diverse organisms ranging from metamorphosis in insects to the development

of the mammalian brain, indicating its evolutionary importance^{1,9}.

Synapse refinement is tightly regulated temporally¹⁰⁻¹¹. Synapse elimination occurs in adults during injury or disease; however, this process is much more prevalent in the developing nervous system¹²⁻¹³. Extensive refinement occurs during critical periods of vertebrate development in the visual system, auditory system, cerebellum, and skeletal muscle¹⁰⁻¹¹. Additionally, refinement is controlled temporally during the larval development of the invertebrate *C. elegans*, where the heterochronic gene *lin-14* controls the timing of GABAergic circuit remodeling¹⁴.

Much of what we know about the process of synapse refinement has come from studies of the vertebrate NMJ, due to its simplicity and accessibility¹⁵. Models of central nervous system (CNS) refinement include the visual system circuitry and climbing fibers in the cerebellum¹⁶. Additionally, the simple nervous systems in *Drosophila* and *C. elegans* provide excellent models for molecular and genetic

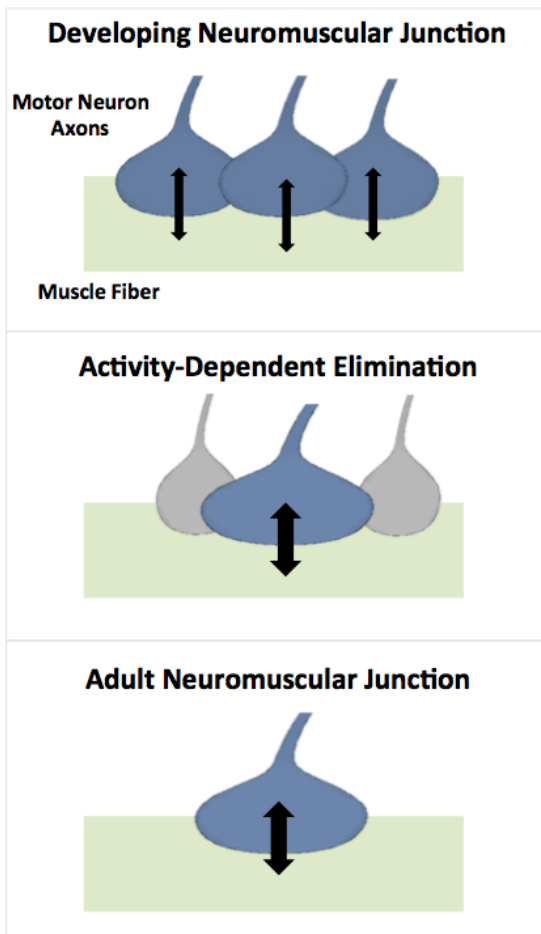


Figure 1: *Activity-dependent elimination at the neuromuscular junction (NMJ).* Initially, axons from multiple motor neurons innervate a single muscle fiber. Activity-dependent elimination results in the strengthening of one synapse (blue), while the other inputs become weaker and are eliminated (gray).

manipulation¹⁷⁻¹⁸. Our current understanding from studying these diverse model systems has led to overarching themes in synapse refinement, highlighted below.

Synaptic Activity and Circuit Refinement

Early studies of the visual system demonstrated that electrical activity plays a critical role in the refinement of neural circuits¹⁹⁻²². Axons from the left and right eye project to the lateral geniculate nucleus (LGN) of the thalamus²³. The thalamus sends projections to the visual, or striate, cortex, relaying information from the eyes²³. In the mature nervous system, inputs from each eye are segregated

into distinct segments in the LGN and cortex²³. The compartments in the visual cortex, termed ocular dominance columns, were identified by the striped patterns seen on cortical sections after injecting dye into one eye¹⁹⁻²³. In a classical experimental paradigm, vision was occluded in one eye of a developing cat^{19,22}. Even when monocular occlusion was restricted to a short window of time, a loss of ocular dominance columns was observed in the visual cortex¹⁹⁻²⁴. Additionally, the presynaptic arbors of the axons from the occluded eye were much smaller, whereas arbors from the non-deprived eye expanded²⁵⁻²⁶. Later studies found that the ocular dominance columns form before birth but can be lost during postnatal critical periods if circuit activity is interrupted²⁷. Extensive refinement also occurs at the LGN². The cells of the LGN are initially innervated by multiple retinal axons, but following eye opening, these synapses are eliminated so that only 1-3 inputs remain^{2,23}.

Similarly, a critical role for circuit activity at the vertebrate NMJ has been well characterized²⁸⁻²⁹. The NMJ consists of the presynaptic motor neuron and the postsynaptic muscle fiber, ensheathed by a Schwann cell³⁰. The presynaptic neuron releases acetylcholine (ACh), which binds to postsynaptic ACh receptors located on the membrane of the muscle fiber³⁰. Initially, multiple axons from different motor neurons synapse with the same muscle fiber²³. Excess inputs are eliminated over time, resulting in each muscle stabilizing innervation from only one motor neuron (**Figure 1**)³¹⁻³². During the elimination process, one input becomes stronger, whereas the other axons become weaker and retract¹⁵. Studies show that blocking activity results in defective elimination and increased numbers of inputs^{15,28-29,31}. Additionally, higher levels of activity can induce input elimination in less time^{15,31-33}. It appears that both presynaptic and postsynaptic activity play an important role in circuit refinement^{15,31,35}. Studies of the rodent NMJ have demonstrated that modulating activity in the axon or muscle cell can influence synaptic elimination^{15,34-35}.

Data now suggest that activity may not be sufficient to promote input elimination; rather, it is proposed that the pattern of neuronal firing dictates synapse refinement. Mature motor neurons that innervate the same muscle fire asynchronously³⁶⁻³⁷. Interestingly, *in vitro* studies of the mammalian NMJ show that activity is synchronous early in development, prior to synapse refinement³⁶⁻³⁷. During development, synchronous activity is replaced by

asynchronous firing, and this occurs around the onset of input elimination³⁶⁻³⁷. The imposition of synchronous activity on the NMJ inhibits the input elimination³⁶⁻³⁷. This study suggests that differential activity is needed at the NMJ to stabilize the active synapses and to prune the less active inputs. Additionally, visual system refinement is sensitive to the pattern of activity. Studies show that flashing strobe lights into the eyes of goldfish and frogs during the critical period of visual circuitry refinement delays synapse elimination and thus fails to maintain ocular dominance columns³⁸⁻³⁹. These results correlate with the Hebbian paradigm in which neurons that fire together are strengthened and maintained, whereas noncoincident activity weakens connections⁴⁰⁻⁴².

Taken together, these studies demonstrate that patterned circuit activity is a conserved player in remodeling the nervous system. Interestingly, not all synaptic refinement events are modulated by activity. Hormonal signaling controls metamorphosis in insects, whereas axon guidance molecules can mediate retraction in the vertebrate CNS^{17,43}. Additionally, refinement can occur at electrically silenced NMJ synapses *in vitro*⁴⁴. These results are important because they show that both activity-dependent and independent pathways can modulate synaptic remodeling.

Competition at the Vertebrate Neuromuscular Junction

Studies at the vertebrate NMJ have been critical in demonstrating that multiple synaptic inputs compete with one another to innervate a target muscle fiber^{25,31,45}. *In vitro* studies showed that in muscle fibers that are innervated by two motor neurons, stimulation of one neuron led to suppression of inputs from the other neuron³⁵. The mechanism of this effect was explored *in vivo* by genetic ablation of choline acetyl-transferase (ChAT) in a subset of motor neurons, thus selectively depleting biosynthesis of the neurotransmitter acetylcholine³¹. When competing with wild-type neural inputs, the ChAT-depleted inputs lost the competition to innervate³¹. This study demonstrates that more active inputs out-compete weaker inputs to stabilize innervations with target muscle. One motor neuron may lose innervation at one site but out-compete and stabilize connections at other sites, suggesting a mechanism to bias connections for maintenance or removal⁴⁶. Interestingly, the competition of motor neurons is reversible^{45,47-48}. Increasing the activity in weaker inputs can cause initially “losing” motor neurons to “win” the competition⁴⁵. Recent studies demonstrate that axons undergoing elimination will reverse their fate if the innervating axon is excised⁴⁵.

This finding suggests that the pruning process is not all-or-none but rather is a continually driven process³⁰. Another interesting characteristic of elimination is that at no time in the refinement process are muscle fibers without innervation, indicating that cellular mechanisms may exist to detect synaptic density and ensure all muscle fibers maintain input from at least one motor neuron⁴⁹.

Studies of competition at the vertebrate NMJ have led to the idea that axons may be competing for a limited trophic factor released from the post-synaptic muscle^{4,15,30}. The inputs with access to more trophic factor are stabilized, whereas inputs receiving less trophic factor are eliminated¹⁵. Studies show that trophic support is required for the maintenance of synapses⁵⁰. The loss of the neurotrophin-4 ligand or its receptor TrkB promotes synaptic elimination at the muscle and cerebellum⁵¹⁻⁵². Overexpression of neurotrophin-4/5 and brain derived neurotrophic factor (BDNF), ligands of the TrkB receptor, prevent elimination and promote synapse stabilization in the visual circuitry of cats⁵³⁻⁵⁴. Additionally, studies at the NMJ suggest that the postsynaptic muscle must have a mechanism to communicate with the presynaptic motor neurons; however, this signal has not yet been identified¹⁵. It remains speculative that neurotrophins are the retrograde signal at the NMJ that stabilize a single motor neuron while destabilizing others. Taken together, these studies suggest that the local control of trophic factors may contribute to the stabilization or removal of competing axons.

It is important to note that not all synapse elimination events involve competition between multiple inputs. At the *Drosophila* NMJ, motor neurons innervate target muscle without competing against other inputs; however, the size of the synapses increases greatly over time⁵⁵. In coordination with this synaptic growth, disassembly occurs in restricted areas and the postsynaptic muscle expands^{30,55-56}. Additionally, competition between inputs does not appear to dictate synapse disassembly in the refinement of *C. elegans* GABAergic circuitry although this process is activity-dependent^{9,57}. These results imply that competition between inputs may be characteristic of the more complex vertebrate nervous systems, while activity-dependent refinement is conserved in simpler invertebrate neural circuits.

Uncovering the Molecular Mechanisms of Synaptic Disassembly

It is unclear what molecular mechanisms connect activity in the nervous system to the cellular constituents that

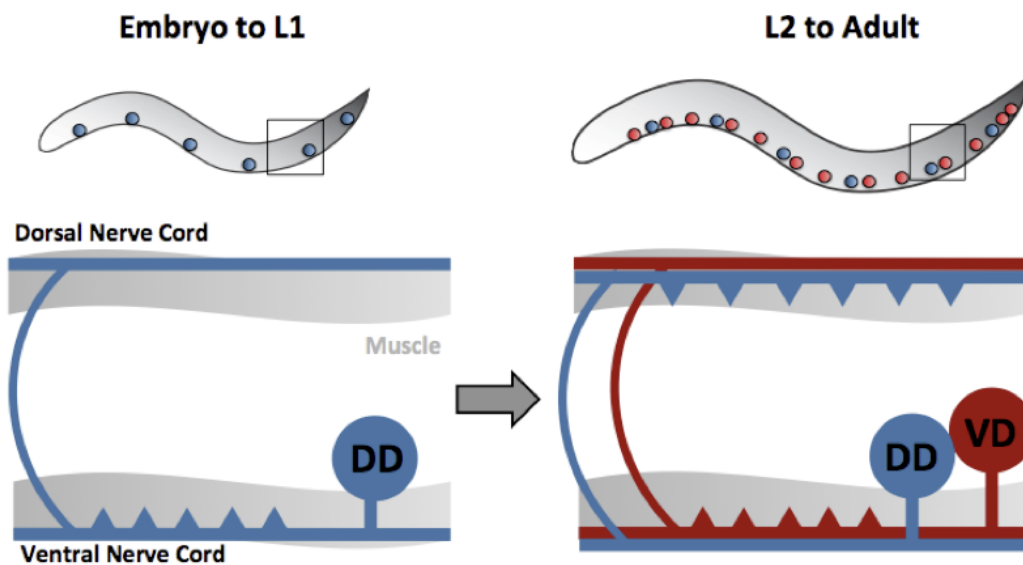


Figure 2: Remodeling the GABAergic motor circuit in *C. elegans*. Dorsal D (DD) motor neurons initially innervate ventral muscle (blue triangles), then relocate these synapses to dorsal muscles following the first larval stage (L1). The Ventral D (VD) motor neurons are generated in the second larval stage (L2) and synapse with ventral muscles (red triangles).

physically dismantle synapses. One hypothesis involves the activity-dependent destabilization of synapses by the ubiquitin proteasome system (UPS)⁵⁸⁻⁶³. The proteasome regulates protein concentrations in the cell and removes defective proteins by degradation⁶². Proteins are targeted for proteasomal destruction by ubiquitin molecules⁶². This process involves three enzymes: the ubiquitin-activating enzyme (E1), the ubiquitin-conjugating enzyme (E2), and the ubiquitin ligase (E3)⁶². The E3 ubiquitin ligase is responsible for substrate specificity⁶². Studies show that ubiquitination of both pre- and post-synaptic proteins occur at the synapse⁵⁸⁻⁶³. Interestingly, the ubiquitination of proteins at the synapse also appears to be modulated by activity⁵⁹⁻⁶⁰. A study in isolated vertebrate hippocampal neurons shows that in response to activity, the UPS degrades PSD-95, a scaffolding protein in the postsynaptic cell⁵⁸. Activity-induced ubiquitination of PSD-95 induces the internalization of AMPA receptors. Thus, this study connects circuit activity to protein turnover and molecular changes at the synapse⁵⁸. Another study shows that Shank, GKAP and AKAP79/150 postsynaptic scaffolds selectively undergo activity-dependent ubiquitination⁵⁹. The selective ubiquitination of scaffolding proteins and their associated proteins could be a mechanism to regulate synapse stability in response to activity⁵⁹. Additionally, it was found that during synapse removal, distinct scaffolds of proteins are eliminated at different times, showing that differential regulation of groups of proteins play an important role in the process of disassembly⁶⁴.

Examining the activity-dependent ubiquitination of pre-synaptic proteins has led to similar findings⁶⁰⁻⁶¹. Studies at the *Drosophila* NMJ show that proteasome inhibitors strengthen synaptic transmission through up-regulation of a vesicular priming component DUNC-13⁶⁰. An additional study in *C. elegans* proposes an interesting model for protecting synapses from ubiquitination and destruction^{61,65}. The immunoglobulin protein SYG-1 interacts with the E3 ubiquitin ligase SKR-1⁶¹. This interaction blocks the binding of SKR-1 to adaptor protein SEL-10, needed for ubiquitin-mediated target selection, thus blocking presynaptic ubiquitination in this area of the neuron⁶¹. This study also suggests that in different areas of the same axon that lack SYG-1, SKR-1 is free to join the active ubiquitin complex and dismantle synapses⁶¹. This model introduces a mechanism by which the presynaptic neuron can spatially dictate synaptic stabilization along a single axon⁶¹. Taken together, these studies demonstrate that ubiquitination of pre- and post-synaptic components may act as a mechanism connecting activity to the cellular processes that destabilize the synapse. More studies will be necessary to understand what specific proteins are targeted for degradation in these systems and how the loss of targeted proteins affects synaptic stability.

Circuit Remodeling in *C. elegans*: A Model of Synaptic Removal

While the examination of vertebrate systems has been helpful to our understanding of synaptic remodeling, it is becoming more apparent that refinement is complex and

COUP-TFII:

(chicken ovalbumin upstream promoter transcription factor-2) highly conserved transcription factor involved in patterning the nervous system in addition to controlling development of multiple organs in the body.

may involve the coordination of multiple cellular processes. Therefore, there is a demand for a simplified model to examine how different conserved pathways are acting with one another to regulate such a complex event. The nematode *C. elegans* is widely used for its ease of genetic manipulations and its highly conserved genome. Interestingly, a subset of neurons in the worm undergoes expansive remodeling during development⁹. The GABAergic Dorsal D (DD) motor neurons are born embryonically and make synapses onto ventral muscles⁹. At a critical window of development, specifically between the first (L1) and second (L2) larval stages, DD synapses with ventral muscles are removed as new DD connections are established with dorsal muscles⁹. Coincidentally, Ventral D's (VDs) are born and innervate the ventral muscle (**Figure 2**)⁹. The UNC-55/COUP-TFII transcription factor functions in VDs to inhibit remodeling⁶⁶⁻⁶⁷. When UNC-55 is genetically ablated, the VDs ectopically remodel, and conversely, over-expression of UNC-55 in DDs blocks remodeling⁶⁶⁻⁶⁸. Therefore, the UNC-55 transcription factor is necessary and sufficient to inhibit the GABAergic motor neuron remodeling program. Activity in the form of neurotransmitter release also modulates this process. Mutants that decrease synaptic vesicle fusion show delayed DD remodeling, whereas mutants that increase neurotransmitter release demonstrate precocious or early remodeling⁶⁹. Recent work demonstrates that this process is very complex. A microarray study was performed to uncover the targets of UNC-55, with the assumption that these would be candidate synaptic remodeling genes¹⁸. This approach identified 49 candidate genes with gene ontology categories ranging from ubiquitin regulation, calcium binding proteins, ion channels, enzymes, extracellular matrix components, transcription factors, cytoskeletal components, and proteins involved in neurotransmission¹⁸. This study demonstrates the complex nature of the remodeling process and yields candidate genes that may be conserved in refinement of the mammalian central nervous system.

Conclusions

The regulation of synaptic refinement is complex; however, with the use of model organisms we are developing deeper insight into the molecular mechanisms that govern this form of neural plasticity. Despite the complexity of the mammalian central nervous system, we have seen that synaptic refinement relies on the activity of neural circuitry and is characterized by competition between multiple inputs. Additionally, it appears that mechanisms to destabilize the synapse may play a role in activity-dependent synapse disassembly. It is unclear how the nervous system is able to coordinate the assembly, disassembly, and reassembly of a vast number of synapses during development. This suggests that the nervous system requires mechanisms to control synapse formation and retraction, in addition to mechanisms that balance the two. Much work will be necessary to elucidate the molecular constituents that refine functional neural circuitry. The utilization of model organisms with simplified nervous systems and malleable genetics will be of great value in exploring these intriguing questions.

References

1. Huttenlocher PR and de Courten C (1987). The development of synapses in striate cortex of man. *Human Neurobiology*. 6(1): 1-9.
1. Chen C and Regehr WG (2000). Developmental remodeling of the retinogeniculate synapse. *Neuron*. 28: 955-966.
2. Colman H, Nabekura J and Lichtman JW (1997). Alterations in synaptic strength preceding axon withdrawal. *Science*. 275(5298): 356-361.
3. Sanes JR and Lichtman JW (1999). Development of the vertebrate neuromuscular junction. *Annual Reviews Neuroscience*. 22: 389-442.
4. Trachtenberg JT, Chen BE, Knott GW, Feng G, Sanes JR, Welker E and Svoboda K (2002). Long-term in vivo imaging of experience-dependent synaptic plasticity in adult cortex. *Nature*. 420(6917): 788-794.
5. Sotelo C (1975). Anatomical, physiological and biochemical studies of the cerebellum from mutant mice. II morphological study of cerebellar cortical neurons and circuits in the weaver mouse. *Brain Research*. 94: 19-44.

6. Crepel F and Mariani J (1976). Multiple innervation of Purkinje cells by climbing fibers in the cerebellum of the Weaver Mutant Mouse. *Journal of Neurobiology*. 7(6): 579-582.
7. Crepel F, Delhaye-Bouchaud N, Guastavino JM and Sampaio I (1980). Multiple innervation of cerebellar Purkinje cells by climbing fibres in staggerer mutant mouse. *Nature*. 283: 483-484.
8. White JG, Albertson DG and Anness MAR (1978). Connectivity changes in a class of motoneurone during the development of a nematode. *Nature*. 271: 764-766.
9. Purves D and Lichtman JW (1980). Elimination of synapses in the developing nervous system. *Science*. 210: 153-157.
10. Hensch TK (2004). Critical period regulation. *Annual Review Neuroscience*. 27: 549-79.
11. Gan WB, Kwon E, Feng G, Sanes JR and Lichtman JW (2003). Synaptic dynamism measured over minutes to months: age-dependent decline in an autonomic ganglion. *Nature Neuroscience*. 6(9): 956-960.
12. Luo L and O'Leary DM (2005). Axon retraction and degeneration in development and disease. *Annu. Rev. Neurosci*. 28:127-156.
13. Hallam SJ and Jin Y (1998). *lin-14* regulates the timing of synaptic remodeling in *Caenorhabditis elegans*. *Nature*. 395: 78-82.
14. Nguyen QT and Lichtman JW (1996). Mechanism of synapse disassembly at the developing neuromuscular junction. *Current Opinion in Neurobiology*. 6: 104-112.
15. Kano M and Hashimoto K (2009). Synapse elimination in the central nervous system. *Current Opinion in Neurobiology*. 19:154-161.
16. Truman JW (1996). Steroid receptors and nervous system metamorphosis in insects. *Dev. Neurosci*. 18(1-2): 87-101
17. **Petersen SC, Watson JD, Richmond JE, Sarov M, Walthall WW and Miller DM (2011). A transcriptional program promotes remodeling of GABAergic synapses in *Caenorhabditis elegans*. *Journal of Neuroscience*. 31(43): 15362-15375.**
This article reveals candidate synaptic remodeling genes and introduces techniques to examine the role of these genes in remodeling.
18. Wiesel TN and Hubel DH (1963). Single-cell responses in striate cortex of kittens deprived of vision in one eye. *Journal of Neurophysiology*. 26(6): 1003-1017.
19. Katz LC and Shatz CJ (1996). Synaptic activity and the construction of cortical circuits. *Science*. 274: 1133-1138.
20. Wiesel TN (1982). Postnatal development of the visual cortex and the influence from the environment. *Nature*. 299: 583-591.
21. Hubel DH and Wiesel TN (1970). The period of susceptibility to the physiological effects of unilateral eye closure in kittens. *J. Physiology*. 206: 419-436.
22. Huberman AD (2007). Mechanisms of eye-specific visual circuit development. *Current Opinion in Neurobiology*. 17: 73-80.
23. Cline HT and Constantine-Paton M (1990). NMDA receptor agonist and antagonists alter retinal ganglion cell arbor structure in the developing frog retinotectal projection. *Journal of Neuroscience*. 10(4): 1197-1216.
24. Antonini A and Stryker MP (1993). Rapid remodeling of axonal arbors in the visual cortex. *Science*. 260: 1819-1821.
25. Antonini A and Stryker MP (1996). Plasticity of geniculocortical afferents following brief or prolonged monocular occlusion in the cat. *Journal of Comparative Neurology*. 369: 64-82.
26. Crowley JC and Katz LC (2002). Ocular dominance development revisited. *Current Opinion Neurobiology*. 12: 104-109.
27. Duxson MJ (1982). The effect of postsynaptic block on development of the neuromuscular junction in postnatal rats. *Journal of Neurocytology*. 11(3): 395-408.
28. Thompson W, Kuffler DP and Jansen JK (1979). The effect of prolonged, reversible block of nerve impulses on the elimination of polyn neuronal innervation of new-born rat skeletal muscle fibers. *Neuroscience*. 4(2): 271-281.
29. Goda Y and Davis GW (2003). Mechanisms of synapse assembly and disassembly. *Neuron*. 40(2): 243-264.
30. **Buffelli M, Burgess RW, Feng G, Lobe CG, Lichtman JW and Sanes JR (2003). Genetic evidence that relative synaptic efficacy biases the outcome of synaptic competition. *Nature*. 424(6947): 430-434.**
This article shows that competition between inputs at the neuromuscular junction is activity-dependent.
31. Balice-Gordon RJ, Chua CK, Nelson CC and Lichtman JW (1993). Gradual loss of synaptic cartels precedes axon withdrawal at developing neuromuscular junctions. *Neuron*. 11(5): 801-815.
32. Thompson W (1983). Synapse elimination in neonatal rat muscle is sensitive to pattern of muscle use. *Nature*. 302(5909): 614-616.
33. Lo YJ, Lin YC, Sanes DH and Poo M (1994). Depression of developing neuromuscular synapses induced by repetitive postsynaptic depolarizations. *Journal of Neuroscience*. 14: 4694-4704.
34. Lo YJ and Poo MM (1991). Activity-dependent synaptic competition in vitro: heterosynaptic suppression of developing synapses. *Science*. 254: 1019-1022.
35. Busetto G, Buffelli M, Cangiano L and Cangiano A (2003). Effects of evoked and spontaneous motoneuronal firing on synapse competition and elimination in skeletal muscle. *Journal of Neurocytology*. 32: 795-802.
36. Buffelli M, Busetto G, Cangiano L and Cangiano A (2002). Perinatal switch from synchronous to asynchronous activity of motoneurons: link with synapse elimination. *Proc. Natl. Acad. Sci*. 99(20): 13200-13205.
37. Brickley SG, Dawes EA, Keating MJ and Grant Simon (1998). Synchronizing retinal activity in both eyes disrupts binocular map development in the optic tectum. *Journal of Neuroscience*. 18(4): 1491-1504.
38. Schmidt JT and Eisele LE (1985). Stroboscopic illumination and dark rearing block the sharpening of the regenerated retinotectal map in goldfish. *Neuroscience*. 14(2): 535-546.
39. Hebb D (1949). *The Organization for Behavior*. New York: Wiley.
40. Stent GS (1973). A physiological mechanism for Hebb's postulate of learning. *Proc. Natl. Acad. Sci*. 70: 997-1001.

CANDIDATE REVIEWS

41. Zhang LI and Poo M (2001). Electrical activity and development of neural circuits. *Nature Neuroscience*. 4:1207-1214.
42. Bagri A, Cheng HJ, Yaron A, Pleasure SJ and Tessier-Lavigne M (2003). Stereotyped pruning of long hippocampal axon branches triggered by retraction inducers of the semaphorin family. *Cell*. 113: 285-299.
43. Constanzo EM, Barry JA and Ribchester RR (2000). Competition at silent synapses in reinnervated skeletal muscle. *Nature Neuroscience*. 3(7): 694-700.
44. Turney SG and Lichtman JW (2012). Reversing the outcome of synapse elimination at developing neuromuscular junctions in vivo: evidence for synaptic competition and its mechanism. *PLoS Biology*. 10(6): 1-15.
45. Kasthuri N and Lichtman JW (2003). The role of neuronal identity in synaptic competition. *Nature*. 424: 426-430.
46. Walsh MK and Lichtman JW (2003). In vivo time-lapse imaging of synaptic takeover associated with naturally occurring synapse elimination. *Neuron*. 37: 67-73.
47. Antonini A, Gillespie DC, Crair MC and Stryker MP (1998). Morphology of single geniculocortical afferents and functional recovery of the visual cortex after reverse monocular deprivation in the kitten. *Journal of Neuroscience*. 18(23): 9896-9909.
48. Buffelli M, Busetto G, Bidoia C, Favero M and Cangiano A (2004). Activity-dependent synaptic competition at mammalian neuromuscular junctions. *News Physiol. Sci*. 19: 85-91.
49. McAllister KA, Katz LC and Lo DC (1999). Neurotrophins and synaptic plasticity. *Ann. Rev. Neurosci*. 22: 295-318.
50. Gonzalez M, Ruggiero P, Chang Q, Shi Y, Rich MM, Kraner S and Balice-Gordon RJ (1999). Disruption of TrkB-mediated signaling induces disassembly of postsynaptic receptor clusters at neuromuscular junctions. *Neuron*. 24: 567-583.
51. Rico B, Baoji X and Reichardt LF (2002). TrkB receptor signaling is required for establishment of GABAergic synapses in the cerebellum. *Nature Neuroscience*. 5(3): 225-233.
52. Cabelli RJ, Hohn A and Shatz CJ (1995). Inhibition of ocular dominance formation by infusion of NT-4/5 or BDNF. *Science*. 267(5204): 1662-1666.
53. Riddle DR, Lo DC and Katz LC (1995). NT-4-mediated rescue of lateral geniculate neurons from effects of monocular deprivation. *Nature*. 378: 189-191.
54. Keshishian H, Broadie K, Chiba A and Bate M (1996). The drosophila neuromuscular junction: a model system for studying synaptic development and function. *Annu. Rev. Neuroscience*. 19: 545-575.
55. Schuster CM, Davis GW, Fetter RD and Goodman CS (1996). Genetic dissection of structural and functional components of synaptic plasticity. I. Fasciclin II controls synaptic stabilization and growth. *Neuron*. 17: 641-654.
56. White JG, Southgate E, Thomson JN and Brenner S (1976). The structure of the ventral nerve cord of *Caenorhabditis elegans*. *Phil. Trans. R. Soc. Lond. B*. 275: 327-348.
57. Colledge M, Snyder EM, Crozier RA, Soderling JA, Jin Y, Langeberg LK, Lu H, Bear M, Scott JD (2003). Ubiquitination regulates PSD-95 degradation and AMPA receptor surface expression. *Neuron*. 40: 595-607.
58. Ehlers MD (2003). **Activity level controls postsynaptic composition and signaling via the ubiquitin-proteasome system.** *Nature*. 6(3): 231-242.
- This article provides a link between activity-dependent ubiquitination and postsynaptic cellular changes that can alter the stability of synapses.*
59. Speese SD, Trotta N, Rodesch CK, Aravamudan B and Broadie K (2003). The ubiquitin proteasome system acutely regulates presynaptic protein turnover and synaptic efficacy. *Current Biology*. 13: 899-910.
60. Ding M, Chao D, Wang G and Shen K (2007). Spatial regulation of an E3 ubiquitin ligase directs selective synapse elimination. *Science*. 317: 947-951.
61. Mabb AM and Ehlers MD (2010). Ubiquitination in postsynaptic function and plasticity. *Annu. Rev. Cell Dev. Biol*. 26: 179-210.
62. Waites CL, Craig AM and Garner CG (2005). Mechanisms of vertebrate synaptogenesis. *Annu. Rev. Neurosci*. 28: 251-274.
63. Culican SM, Nelson CC and Lichtman JW (1998). Axon withdrawal during synapse elimination at the neuromuscular junction is accompanied by disassembly of the postsynaptic specialization and withdrawal of Schwann cell processes. *Journal of Neuroscience*. 18(13): 4953-4965.
64. Miller DM (2007). Synapses here and not everywhere. *Science*. 317: 907-908.
65. Walthall WW and Plunkett JA (1995). Genetic transformation of the synaptic pattern of a motoneuron class in *Caenorhabditis elegans*. *Journal of Neuroscience*. 15(2): 1035-1043.
66. Zhou HM and Walthall WW (1998). UNC-55, an orphan nuclear hormone receptor, orchestrates synaptic specificity among two classes of motor neurons in *Caenorhabditis elegans*. *Journal of Neuroscience*. 18(24): 10438-10444.
67. Shan G, Kim K, Li C and Walthall WW (2005). Convergent genetic programs regulate similarities and differences between related motor neuron classes in *Caenorhabditis elegans*. *Developmental Biology*. 280: 494-503.
68. **Thompson-Peer KL, Bai J, Hu Z and Kaplan JM (2012). HBL-1 patterns synaptic remodeling in *C. elegans*.** *Neuron*. 73: 453-465.
- This article shows that synaptic remodeling of the GABAergic circuit in *C. elegans* is activity-dependent.*

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Roles of Serotonin Signaling in Embryonic and Postnatal Neurogenesis

Elaine Ritter

Serotonin (5-hydroxytryptamine, 5-HT) is perhaps the most well-studied of the classical neurotransmitters and has been implicated in many behavioral adaptations as well as essential physiological functions. In addition to its role in these processes, 5-HT is also a vital regulator of development and begins acting as early as the end of gastrulation and continues throughout postnatal development. A rich body of research has established that 5-HT is intimately involved in the survival, proliferation, migration, differentiation, and maturation of neurons, and the formation of functional synapses. This review surveys experiments that have demonstrated the role of 5-HT signaling in each of these processes, most of which are mediated by G-protein coupled receptor signaling cascades. However, 5-HT₃, the only ligand-gated cation channel member of the 5-HT receptor gene family, has been understudied in the context of development. Evidence presented here suggests a significant role for the 5-HT₃ receptor in neurogenesis, but much is still left to be learned about the mechanisms by which it acts. The information gained from the study of 5-HT signaling in neurogenesis has important implications for a more thorough understanding of the development of the central and peripheral nervous systems.

Keywords *Serotonin, serotonin receptors, neurogenesis, development, signaling cascade, molecular biology*

Introduction

Serotonin (5-hydroxytryptamine, 5-HT) is an evolutionarily ancient molecule¹ that is critical for the modulation of many physiological and behavioral adaptations. Among these adaptations are appetite, gastrointestinal function, nociception, sexual behavior, mood, learning and memory. The role of 5-HT in these processes has been extensively studied for decades. Outside the realm of classical neurotransmission, 5-HT is considered to be a key regulator of neurogenesis through neurotrophic effects^{2,3}, and this notion is well supported in the literature. 5-HT expression is highly abundant in the developing central and peripheral nervous systems⁴. Additionally, serotonergic neurons housed in the brainstem are among the earliest born^{5,6} and they innervate most regions of the brain⁷. One of the foundational experiments identifying 5-HT in neurodevelopment demonstrated that pharmacological depletion of 5-HT

during rat embryogenesis results in impaired neural differentiation in the brain⁸. Since then, advances in molecular biology and pharmacology have permitted study of the modulation of neurogenesis by 5-HT during embryonic and postnatal development, specifically by the activity of 5-HT receptors and the 5-HT transporter, SERT. This review highlights research that attests to the importance of 5-HT signaling throughout neurogenesis in mammalian development and identifies questions in the field that remain unanswered.

Serotonin in the Modulation of Survival, Proliferation and Differentiation

Numerous studies support the hypothesis that 5-HT promotes the proliferation and survival of neuronal progenitors early in embryogenesis. One way 5-HT acts in these processes is by mediating the activity of glycogen synthase kinase-3 β (GSK3 β). GSK3 β

Neurogenesis:

The process by which neurons are generated from neural stem and/or precursor cells.

Differentiation:

The developmental process of neural progenitors acquiring the genetic and phenotypic characteristics of their terminal neuronal fates.

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Wnt signaling pathway:

Highly conserved cellular signal transduction pathway that plays a crucial role in embryonic and fetal development; known downstream effects include modulation of gene expression, cytoskeletal dynamics, and intracellular calcium levels.

Specification:

Also referred to as lineage segregation or divergence, the means by which neural precursors are assigned neuronal identities but have yet to become determined or undergo differentiation.

acts as a negative regulator of neurogenesis by inhibiting β -catenin in the canonical Wnt signaling pathway. Inhibition of GSK3 β allows β -catenin to enter the nucleus and activate target genes that promote the survival and proliferation of neuronal progenitors⁹. The effect of 5-HT on GSK3 β activity was first observed in the mouse brain—upon administration of d-fenfluramine, a SERT inhibitor, GSK3 β becomes phosphorylated and subsequently inhibited in the cerebral cortex and hippocampus¹⁰. A later experiment showed a similar effect in animals lacking functional tryptophan hydroxylase-2 (Tph2), a key enzyme in neuronal 5-HT synthesis: Tph2 null mice also display phosphorylated, inhibited GSK3 β ¹¹. In both experiments, inhibited GSK3 β results in perpetuation of the neural stem cell profile, such that survival and proliferation programs are maintained. Interestingly, the selective serotonin reuptake inhibitor (SSRI) fluoxetine has been proposed to mediate its antidepressant effects by fostering neurogenesis in the hippocampus¹²⁻¹⁴. This hypothesis is supported by the observation that fluoxetine increases phosphorylation and inhibition of GSK3 β , thereby promoting survival and proliferation^{15,16}. Further experimentation revealed that inhibition of GSK3 β occurs by signaling through the serotonin type 1A receptor (5-HT1A). Specifically, 5-HT1A signaling stimulates the kinase activity of Akt^{17,18}, which is known to phosphorylate and inhibit GSK3 β ¹⁹⁻²¹. In favor of this mechanism, it was shown that 5-HT1A-specific antagonists block phosphorylation of GSK3 β by Akt¹⁰. Collectively these findings demonstrate that 5-HT1A signaling causes inhibition of GSK3 β , stimulating survival and proliferation in neurogenesis.

While there is no evidence substantiating a direct role for 5-HT in specification of neurons, there is support for 5-HT regulating mechanisms of differentiation. Contradictory to its influence on GSK3 β inhibition, 5-HT1A signaling mediates effects that oppose the proliferation and survival of neuronal precursors by promoting neuronal

differentiation. Autoinhibition of the 5-HT1A receptor moderates serotonergic neuronal differentiation in the developing raphe nucleus, as evidenced by increased serotonergic density in this region in animals lacking 5-HT1A²². It has also been noted that the 5-HT1A receptor is expressed early in the mouse fetal brain, and over developmental time its autoinhibitory activity results in decreased 5-HT1A expression as serotonergic neuronal differentiation increases²³. Corroborating these results, other experiments showed that the absence of SERT leads to excessive stimulation of 5-HT1A receptors and subsequent inhibition of serotonergic neuronal development²⁴. Aside from serotonergic neurons, 5-HT signaling also stimulates differentiation of glutamatergic neurons in the embryonic cerebral cortex²⁵, demonstrating that 5-HT affects the differentiation of multiple neuronal subtypes. In an effort to tease apart the mechanisms underlying these effects, work in cultured cells from the developing brainstem and cortex of fetal mice showed that 5-HT1A activity stimulates the release of S100 β from astroglial cells expressing this receptor²⁶⁻²⁹. S100 β , when secreted from astroglial cells and taken in by neighboring neuronal progenitors, is anti-apoptotic^{30,31} and promotes neuronal differentiation³². This factor appears to be involved in regulating the stability of tubulin in the construction of microtubules, thus serving to help stabilize the neuronal cytoskeletal architecture required for differentiation³³. These collective reports indicate that the 5-HT1A receptor plays opposing roles in neurogenic processes and further study is needed to dissect its mechanisms of action in various settings.

The influence of 5-HT signaling on survival, proliferation and differentiation of neuronal progenitors is not unique to the central nervous system. While 5-HT2B promotes survival in cortical neuronal progenitors^{34,35}, it also modulates survival and differentiation during peripheral neurogenesis. Depletion of 5-HT2B at the onset of neurulation in mice results in precocious neuronal differentiation

and failure of cranial neural crest cells to migrate to the periphery³⁶. In the enteric nervous system (ENS), it was shown by Michael Gershon's group that this receptor is important for initiating differentiation of enteric neurons³⁷. Moreover, they have also published studies demonstrating the requirement of 5-HT₄ for the generation and survival of neurons in the developing postnatal enteric nervous system³⁸. By ablation of Tph2 gene expression, they showed that 5-HT is necessary for the proper differentiation and maintenance of dopaminergic neurons in the gut³⁹. These experiments collectively emphasize the significance of 5-HT signaling in the development of several neuronal populations, although the underlying molecular mechanisms have yet to be dissected. How 5-HT functions in the neurodevelopment of other visceral organs outside the ENS is still largely unknown. Clearly, the means by which serotonin signaling impacts the beginning stages of neurogenesis are highly diverse and the downstream effects of several 5-HT receptors vary throughout the course of development.

Serotonin in Neuronal Maturation and Synaptogenesis

An extensive body of literature reports the effects of 5-HT signaling on the maturation of the neuronal phenotype following differentiation and the formation of functional synapses. The involvement of 5-HT in neurite outgrowth and dendritic arborization is especially well-supported by numerous studies. In embryonic development of the mouse raphe nucleus, loss of the 5-HT_{1A} receptor results in an increase in neurite number and length²². A comparable autoinhibitory effect of this receptor was noted in cultured fetal rat cortical neurons⁴⁰. However, deprivation of 5-HT during postnatal rat development ultimately leads to deficient dendritic branching on granule cells of the dentate gyrus, which can be rescued with a 5-HT_{1A} receptor agonist⁴¹. As described earlier in the context of proliferation and differentiation, it appears that 5-HT_{1A} activity has varying effects either by promoting or inhibiting neuritic outgrowth and maturation. Other 5-HT receptors are also influential in neuronal maturation. For example, in fetal mice, enhanced neurite outgrowth is seen when 5-HT_{1B} receptors expressed in thalamic neurons are stimulated⁴². Similarly, detailed studies have revealed that the 5-HT₇ receptor activity regulates neuronal architecture in the construction of cortical columns as serotonergic inputs connect to Cajal-Retzius cells⁴³. Coupling between stimulated 5-HT₇ and the $\text{G}\alpha\text{-12}$ protein activates the RhoA and Cdc42 signaling cascades. When activated by $\text{G}\alpha\text{-12}$,

these factors promote and impede, respectively, neurite outgrowth and growth cone motility^{20,44-46}. Conversely, 5-HT₄ activity, by coupling to the heterotrimeric G₁₃ protein and activating the RhoA signaling cascade, inhibits neurite outgrowth and cell-rounding during neurogenesis in the hippocampus^{47,48}. Analogous to the complexity of actions seen in survival and proliferation, the opposing effects of 5-HT in neurite outgrowth and maturation attest to the diversity of the roles this neurotransmitter can play in multiple neurogenic processes.

In addition to its influence on dendritic morphology, 5-HT has also been implicated in moderating axon guidance mechanisms in development. Work from Levitt and colleagues in recent years have clearly demonstrated a unique role for 5-HT signaling in guidance of thalamocortical axons in the developing forebrain of rodents and humans. Using 5-HT_{1A/1D} receptor-specific drugs, they found that activation of the G_{i/o}-protein signaling pathway through these receptors, which inhibits adenylyl cyclase and decreases intracellular levels of cAMP⁴⁹, causes the axonal attractant netrin-1 to become a repulsive cue for migrating axons⁵⁰. Additionally, segregation of axons in developing thalamic sensory projections relies on appropriate levels of 5-HT signaling in the early postnatal brain—overstimulation of 5-HT_{1B} receptors results in cytoarchitectural aberrations of somatosensory projections to the thalamus and thalamic projections to the barrel field cortex⁵¹. All the studies reported here emphasize the importance of 5-HT signaling in neuronal maturation processes.

The role of 5-HT signaling in neurogenesis extends from neuronal maturation and axon guidance to the formation of synapses during embryonic and postnatal development. Recalling the early neurogenic effects of 5-HT signaling on s100 β expression, 5-HT continues to act through s100 β to support synaptogenesis and synaptic plasticity. Transient depletion of 5-HT during rat postnatal brain development leads to a subsequent loss of s100 β , which ultimately results in a thinning of synaptic density in the hippocampus of adult rats⁵². The consequence of 5-HT and s100 β loss during this time is permanent, such that even after restoration of endogenous 5-HT these synapses cannot be reformed in adulthood. The actions of 5-HT signaling on s100 β not only affect synapse formation but also plasticity. When s100 β is lost early in mouse development, the remaining hippocampal synapses are less likely to adapt to learning and memory formation compared to control animals⁵³. It

is important to recognize that the effects of 5-HT signaling during developmental neurogenesis continue to impact adult brain structure and function.

A Potential Role for the 5-HT₃ Receptor in Neurogenesis

Throughout this review of serotonin's modulation of neurogenesis, the focus has been on the members of the 5-HT receptor family that are G-protein coupled receptors. The 5-HT₃ receptor is the only serotonin receptor that is a ligand-gated ion channel^{54,55}, and there are several lines of evidence that implicate an important role for 5-HT₃ in central nervous system development. Loss of 5-HT₃ activity in cortical development results in decreased levels of reelin. The expression of reelin, an extracellular matrix glycoprotein involved in neurogenesis, has been demonstrated in Cajal-Retzius cells in layer I of the cortex in mice⁵⁶. Reelin expression in this cell population is dependent upon the reception of excitatory input through 5-HT_{3A} receptors⁵⁷. It has also been demonstrated that reelin helps coordinate the migration of sympathetic preganglionic neuronal progenitors to the spinal cord in fetal development⁵⁸, which suggests a role for 5-HT signaling in this system. 5-HT₃ receptors have also been shown to be involved in the migratory patterns and maturation of GABAergic interneurons in mouse cortical development. Application of a 5-HT₃ agonist causes this cell population to have abnormally long neuritic processes with few branches, while a 5-HT₃ antagonist results in the formation of numerous, short processes and failure to migrate to the cortical plate⁵⁹. Moreover, fluoxetine has been demonstrated to functionally suppress the activity of 5-HT₃ receptors⁶⁰, and very recently it was discovered that prenatal exposure to fluoxetine reduces dendritic complexity by nearly 50% in pyramidal neurons of the cortex in mice⁶¹. This receptor's actions in neurogenesis extend into the construction of neuronal architecture. Elegant experiments have demonstrated an interaction between the intracellular portion of the 5-HT₃ receptor and F-actin, implying that 5-HT₃ modulates cytoskeletal structure during neuronal migration and maturation⁶². 5-HT₃ activity appears to mediate multiple neurogenic processes throughout the course of neurodevelopment—yet, the causal mechanisms underlying the function of 5-HT₃ are poorly understood.

It is widely known that regulation of calcium flux in neural progenitors is required for proper nervous system development to occur. Interestingly, presynaptically

localized 5-HT₃ is permeable to calcium in neurons of the corpus striatum, hippocampus and amygdala^{63,64}. 5-HT₃ receptors have enhanced calcium permeability in several neuroblastoma cell lines as well^{65,66}. Given this information, it is plausible that 5-HT₃ exerts its effects on neurogenesis via calcium signaling. Research supporting this hypothesis has only been conducted fairly recently. PC12 cells express 5-HT₃, and an increase in intracellular calcium levels was observed in response to treatment with a 5-HT₃ agonist⁶⁷. This effect led to Nerve Growth Factor (NGF) upregulation, resulting in neurite outgrowth and differentiation, and is blocked by a 5-HT₃ antagonist. Surprisingly, the L-type calcium channel antagonist nifedipine also inhibits the 5-HT-induced increase in intracellular calcium and its stimulation of NGF. The response to nifedipine suggests that 5-HT₃ stimulation likely affects the activity of voltage-gated calcium channels, providing another mechanism by which 5-HT₃ receptors are responsible for regulating calcium flux in developing neurons. This hypothesis is especially intriguing, as the calcium-dependent promotion of Brain Derived Neurotrophic Factor (BDNF) transcription by the CREB pathway is mediated by L-type calcium channels⁶⁸⁻⁷⁰. It was also shown that 5-HT₃ receptors are expressed transiently in the glutamatergic granule cells of the developing cerebellum, where they were shown to be critical for promoting plasticity during synaptogenesis of Purkinje cells and parallel fibers⁷¹. The authors of this work postulate that 5-HT₃ may be mediating its effects via control of calcium flux, either through 5-HT₃ itself or its activation of L-type calcium channels, but this possibility has not yet been pursued.

In contrast to the intriguing studies conducted in the CNS, the role of 5-HT₃ in PNS development is almost entirely unknown. In 1996, Johnson and Heinemann reported expression of 5-HT₃ in the neural crest cells of rat embryos aged 15 days post coitus (dpc), including sympathetic and parasympathetic ganglia of the enteric nervous system and the dorsal root ganglia (DRG)⁷². This observation is corroborated by demonstration of 5-HT₃ gene expression in the DRG of 14.5 dpc fetal mice⁷³. There has been no follow up on these findings, so the function of 5-HT₃ throughout neural crest development remains unclear. As mentioned previously, the development of some aspects of the PNS is poorly studied; among these is the innervation of the lower urinary tract (LUT). Proper 5-HT₃ function is critical for maintaining the autonomic innervation of the LUT in adult mice⁷⁴, but its role in the development of this system has not yet been explored. Based on the importance

of 5-HT signaling in neurogenesis and the preliminary observations outlined here, there is strong support for the hypothesis that signaling through the 5-HT₃ receptor influences neural crest development and the innervation of the LUT.

To investigate the function of 5-HT₃ in neural crest cells, a wide variety of tools is available. Transgenic reporter mouse lines⁷⁵ allow visualization of 5-HT₃ expression throughout embryonic and postnatal development. A 5-HT₃ knockout line exists⁷⁶ and would be valuable to study the effects of loss of this receptor on neural crest survival, differentiation and migration to the LUT. *In vitro* study of mechanisms by which 5-HT₃ mediates neuronal specification of neural crest progenitors is possible with numerous drugs affecting 5-HT₃ activity⁴. Pharmacological agents may also be used to tease apart the downstream signaling cascades regulating neurogenesis that are stimulated by 5-HT₃. Additionally, emerging technology facilitating imaging of calcium flux in live cells permits examination of the functionality of 5-HT₃ in neurogenesis⁷⁷. These and other molecular biology techniques will allow researchers to begin to elucidate the role of 5-HT₃ signaling in PNS neurogenesis.

Conclusions

A broad body of evidence, including studies described here and others not mentioned, underscores the importance of 5-HT signaling in multiple processes that comprise neurogenesis. 5-HT has been shown to be intimately involved in the survival, proliferation and differentiation of neuronal progenitors. Additionally, 5-HT functions to modulate the migration of differentiating neurons, the augmentation of neurites, and the construction of synapses and cellular architecture. The diversity of the processes regulated by 5-HT is reflected in the myriad signaling mechanisms by which 5-HT acts via its receptors. In fact, the same receptor can even serve oppositional functions throughout the course of development, as is the case with 5-HT_{1A} and 5-HT₇. While some of the signaling cascades mediating these processes have been dissected, much work remains to be done in order to discover the ways in which 5-HT signaling is able to take on so many roles throughout the course of neurogenesis. Especially compelling for future study is the 5-HT₃ receptor, the only ligand-gated ion channel in the serotonin receptor family. Several publications implicate a significant role of 5-HT₃ in neurogenesis—however, no one has yet assembled the pieces of the puzzle to understand precisely the ways by

which this receptor affects neurogenic processes. The subject of 5-HT₃ in peripheral neurogenesis is still largely untouched. Fortunately, pharmacological and molecular tools currently available make it possible to investigate how 5-HT₃ guides neurogenesis in embryonic and postnatal development.

References

1. Turlejski K (1996). Evolutionary ancient roles of serotonin: long-lasting regulation of activity and development. *Acta Neurobiol Exp*. 56: 619-636.
2. Baker P and Quay W (1969). 5-hydroxytryptamine metabolism in early embryogenesis, and the development of brain and retinal tissues. *Brain Res*. 12: 273-295.
3. McMahon D (1974). Chemical messengers in development: a hypothesis. *Science*. 185: 1012-1021.
4. Nichols DE and Nichols CD (2008). Serotonin receptors. *Chem Rev*. 108 (5): 1614-1641.
5. Olson L and Seiger A (1972). Early prenatal ontogeny of central monoamine neurons in the rat: fluorescence histochemical observations. *Z Anat EntwGesch*. 137: 301-316.
6. Lauder J and Bloom F (1974). Ontogeny of monoamine neurons in the locus coeruleus, raphe nuclei, and substantia nigra of the rat: I. *Cell Differentiation*. *J Comp Neurol*. 155: 469-481.
7. Jacobs B and Azmitia EC (1992). Structure and function of the brain serotonin system. *Physiol Rev*. 72: 165-229.
8. **Lauder J and Krebs H (1978). Serotonin as a differentiation signal in early neurogenesis. *Dev Neurosci*. 1: 15-30.**
This paper was one of the earliest studies that examined the role of serotonin in embryonic neurogenesis. The researchers found that serotonin signaling is necessary for differentiation and maintenance of neurons in the brain.
9. Kim W, Wang X, Wu Y, Doble B, Patel S, Woodgett J and Snider W (2009). GSK-3 is a master regulator of neural progenitor homeostasis. *Nat Neurosci*. 12 (11): 1390-1399.
10. Li X, Zhu W, Roh M, Friedman A, Rosborough K and Jope R (2004). In vivo regulation of glycogen synthase kinase-3beta (GSK3beta) by serotonergic activity in mouse brain. *Neuropsychopharmacology*. 29: 1426-1431.
11. Beaulieu J, Zhang X, Rodriguez R, Sotnikova T, Cools M, Wetsel W, Gainetdinov R and Caron M (2008). Role of GSK3 beta in behavioral abnormalities induced by serotonin deficiency. *Proc Natl Acad Sci U S A*. 105 (4): 1333-1338.
12. Bachis A, Mallei A, Cruz MI, Wellstein A and Mocchetti I (2008). Chronic antidepressant treatments increase basic fibroblast growth factor and fibroblast growth factor-binding protein in neurons. *Neuropharmacology*. 55 (7): 1114-1120.
13. Czeh B, Muller-Keuker J, Rygula R, Abumaaria N, Hiemke C, Domenici E and Fuchs E (2007). Chronic social stress inhibits cell proliferation in the adult medial prefrontal cortex: hemispheric asymmetry and reversal by fluoxetine treatment. *Neuropsychopharmacology*. 32: 1490-1503.

CANDIDATE REVIEWS

14. Encinas JM, Vaahtokari A and Enikolopov G (2006). Fluoxetine targets early progenitor cells in the adult brain. *Proc Natl Acad Sci U S A.* 103 (21): 8233-8238.
15. McManus E, Sakamoto K, Armit L, Ronaldson L, Shpiro N, Marquez R and Alessi D (2005). Role that phosphorylation of GSK3 plays in insulin and Wnt signalling defined by knockin analysis. *EMBO J.* 24: 1571-1583.
16. Polter A, Beurel E, Yang S, Garner R, Song L, Miller C, Sweatt J, McMahon L, Bartolucci A, Li X and Jope R (2010). Deficiency in the inhibitory serine-phosphorylation of glycogen synthase kinase-3 increases sensitivity to mood disturbances. *Neuropsychopharmacology.* 35: 1761-1774.
17. Cowen DS, Sowers R and Manning D (1996). Activation of a mitogen-activated protein kinase (ERK2) by the 5-hydroxytryptamine 1A receptor is sensitive not only to inhibitors of phosphatidylinositol 3-kinase, but to an inhibitor of phosphatidylcholine hydrolysis. *J Biol Chem.* 271 (22): 297-322.
18. Cowen DS, Johnson-Farley N and Travkina T (2005). 5-HT1A receptors couple to activation of Akt, but not extracellular-regulated kinase (ERK) in cultured hippocampal neurons. *J Neurochem.* 93: 910-917.
19. Fang X, Yu S, Lu Y, Bast RJ, Woodgett J and Mills G (2000). Phosphorylation and inactivation of glycogen synthase kinase 3 by protein kinase A. *Proc Natl Acad Sci U S A.* 97 (22): 11960-11965.
20. Li M, Wang X, Meintzer M, Laessig T, Birnbaum M and Heidenreich K (2000). Cyclic AMP promotes neuronal survival by phosphorylation of glycogen synthase kinase 3beta. *Mol Cell Biol.* 20: 9356-9363.
21. Cross D, Alessi D, Cohen P, Andjelkovich M and Hemmings B (1995). Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature.* 378: 785-789.
22. Rumajogee P, Verge D, Hanoun N, Brisorgueil MJ, Hen R, Lesch KP, Hamon M and Miquel MC (2004). Adaption of the serotonergic neuronal phenotype in the absence of 5-HT autoreceptors or the 5-HT transporter: involvement of BDNF and cAMP. *Eur J Neurosci.* 19 (4): 937-944.
23. Hillion J, Milne-Edwards JB, Catelon J, de Vitry F, Gros F and Hamon M (1993). Prenatal developmental expression of rat brain 5-HT1A receptor gene followed by PCR. *Biochem Biophys Res Commun.* 191 (3): 991-997.
24. Galter D and Unsicker K (2000). Brain-derived neurotrophic factor and trkB are essential for cAMP-mediated induction of the serotonergic neuronal phenotype. *J Neurosci Res.* 61 (3): 295-301.
25. Lavdas AA, Blue ME, Lincoln J and Parnavelas JG (1997). Serotonin promotes the differentiation of glutamate neurons in organotypic slice cultures of the developing cerebral cortex. *J Neurosci.* 17 (20): 7872-7880.
26. Whitaker-Azmitia P and Azmitia EC (1989). Stimulation of astroglial serotonin receptors produces media which regulates development of serotonergic neurons. *Brain Res.* 497: 80-85.
27. Azmitia EC, K D and Whitaker-Azmitia P (1990). S-100B, but not NGF, EGF or insulin functions as a serotonergic growth factor. *Brain Res.* 516: 354-360.
28. Whitaker-Azmitia P, Clarke C and Azmitia EC (1993). Localization of 5-HT-1A receptors to astroglial cells in adult rats. *Synapse.* 14: 201-205.
29. Ramos A, Tagliaferro P, Lopez E, Pecci Saavedra J and Brusco A (2000). Neuroglial interactions in a model of para-chlorophenylalanine-induced serotonin depletion. *Brain Res.* 883 (1): 1-14.
30. Brewton L, Haddad L and Azmitia EC (2001). Colchicine-induced cytoskeletal collapse and apoptosis in N-18 neuroblastoma cultures is rapidly reversed by applied S-100b. *Brain Res.* 912 (1): 9-16.
31. Bhattacharyya A, Oppenheim RW, Prevet D, Moore BW, Brackenbury R and Ratner N (1992). S100 is present in developing chicken neurons and schwann cells and promotes motor neuron survival in vivo. *J Neurobiol.* 23 (4): 451-466.
32. Selinfreund RH, Barger SW, Welsh MJ and van Eldik LJ (1990). Antisense inhibition of glial S100B production results in alterations in cell morphology, cytoskeletal organization, and cell proliferation. *J Cell Biol.* 111: 2021-2028.
33. Hesketh J and Baudier J (1986). Evidence that S100 proteins regulate microtubule assembly and stability in rat brain extracts. *Int J Biochem.* 18 (8): 691-695.
34. Stankovski L, Alvarez C, Ouimet T, Vitalis T, El-Hachimi KH, Price D, Deneris ES, Gaspar P and Cases O (2007). Developmental cell death is enhanced in the cerebral cortex of mice lacking the brain vesicular monoamine transporter. *J Neurosci.* 27 (6): 1315-1324.
35. Persico AM, Baldi A, Dell'Acqua ML, Moessner R, Murphy DL, Lesch KP and Keller F (2003). Reduced programmed cell death in brains of serotonin transporter knockout mice. *Neuroreport.* 14 (3): 341-344.
36. Choi D, Kellerman O, Richard S, Colas J, Bolanos-Jimenez F, Tournois C, Launay J and Maroteaux L (1998). Mouse 5-HT2B receptor-mediated serotonin trophic functions. *Annals of the New York Academy of Sciences.* 861: 67-73.
37. Fiorica-Howells E, Maroteaux L and Gershon MD (2000). Serotonin and the 5-HT(2B) receptor in the development of enteric neurons. *J Neurosci.* 20 (1): 294-305.
38. Liu MT, Kuan YH, Wang J, Hen R and Gershon MD (2009). 5-HT4 receptor-mediated neuroprotection and neurogenesis in the enteric nervous system of adult mice. *J Neurosci.* 29 (31): 9683-9699.
39. **Li Z, Chalazonitis A, Huang YY, Mann JJ, Margolis KG, Yang QM, Kim DO, Cote F, Mallet J and Gershon MD (2011). Essential roles of enteric neuronal serotonin in gastrointestinal motility and the development/survival of enteric dopaminergic neurons. *J Neurosci.* 31 (24): 8998-9009.**
This paper provides strong evidence for a significant role of serotonin signaling in neurogenesis during the fetal development of the peripheral nervous system. The researchers were able to use a wide variety of molecular biology techniques to demonstrate the effect of 5-HT signaling on the development of dopaminergic neurons in the gut in vivo.
40. Sikich L, Hickok JM and Todd RD (1990). 5-HT1A receptors control neurite branching during development. *Dev Brain Res.* 56 (2): 269-274.
41. Yan W, Wilson CC and Haring JH (1997). 5-HT1a receptors

- mediate the neurotrophic effect of serotonin on developing dentate granule cells. *Dev Brain Res.* 98 (2): 185-190.
42. Lotto B, Upton L, Price DJ and Gaspar P (1999). Serotonin receptor activation enhances neurite outgrowth of thalamic neurones in rodents. *Neurosci Lett.* 269 (2): 87-90.
43. Janusonis S, Gluncic V and Rakic P (2004). Early serotonergic projections to Cajal-Retzius cells: relevance to cortical development. *J Neurosci.* 24 (7): 1652-1659.
44. Kobe F, Guseva D, Jensen TP, Wirth A, Renner U, Hess D, Muller M, Medrihan L, Zhang W, Zhang M, Braun K, Westerholz S, Herzog A, Radyushkin K, El-Kordi A, Ehrenreich H, Richter DW, Rusakov DA and Ponimaskin E (2012). 5-HT7R/G12 signaling regulates neuronal morphology and function in an age-dependent manner. *J Neurosci.* 32 (9): 2915-2930.
45. Lee T, Winter C, Marticke SS, Lee A and Luo L (2000). Essential roles of *Drosophila* RhoA in the regulation of neuroblast proliferation and dendritic but not axonal morphogenesis. *Neuron.* 25 (2): 307-316.
46. Newey SE, Velamoor V, Govek EE and van Aelst L (2005). Rho GTPases, dendritic structure, and mental retardation. *J Neurobiol.* 64 (1): 58-74.
47. Ponimaskin E, Profirovic J, Vaiskunaite R, Richter DW and Voyno-Yasenetskaya TA (2002). 5-Hydroxytryptamine 4(a) receptor is coupled to the G α subunit of the heterotrimeric G13 protein. *J Biol Chem.* 277 (23): 20812-20819.
48. Kvachnina E, Liu G, Dityatev A, Renner U, Dumuis A, Richter DW, Dityateva G, Schachner M, Voyno-Yasenetskaya TA and Ponimaskin EG (2005). 5-HT7 receptor is coupled to G α subunits of heterotrimeric G12-protein to regulate gene transcription and neuronal morphology. *J Neurosci.* 25 (34): 7821-7830.
49. Hamblin MW and Metcalf MA (1991). Primary structure and functional characterization of a human 5-HT_{1D}-type serotonin receptor. *Mol Pharmacol.* 40 (2): 143-148.
50. Bonnin A, Torii M, Wang L, Rakic P and Levitt P (2007). Serotonin modulates the response of embryonic thalamocortical axons to netrin-1. *Nat Neurosci.* 10 (5): 588-597.
51. Salichon N, Gaspar P, Upton AL, Picaud S, Hanoun N, Hamon M, De Maeyer E, Murphy DL, Moessner R, Lesch KP, Hen R and Seif I (2001). Excessive activation of serotonin (5-HT) 1B receptors disrupts the formation of sensory maps in monoamine oxidase A and 5-HT transporter knock-out mice. *J Neurosci.* 21 (3): 884-896.
52. Mazer C, Muneyyirci J, Taheny K, Raio N, Borella A and Whitaker-Azmitia P (1997). Serotonin depletion during synaptogenesis leads to decreased synaptic density and learning deficits in the adult rat: a possible model of neurodevelopmental disorders with cognitive deficits. *Brain Res.* 760: 68-73.
53. Nishiyama H, Knopfel T, Endo S and Itohara S (2002). Glial protein S100B modulates long-term neuronal synaptic plasticity. *Proc Natl Acad Sci U S A.* 99 (6): 4037-4042.
54. Derkach V, Surprenant A and North R (1989). 5-HT₃ receptors are membrane ion channels. *Nature.* 339: 706-709.
55. Boess FG, Beroukhim R and Martin IL (1995). Ultrastructure of the 5-Hydroxytryptamine 3 receptor. *J Neurochem.* 64: 1401-1405.
56. Alcantara S, Ruiz M, D'Archangelo G, Ezan F, de Lecea L, Curran T, Sotelo C and Soriano E (1998). Regional and cellular patterns of reelin mRNA expression in the forebrain of the developing and adult mouse. *J Neurosci.* 18 (19): 7779-7799.
57. **Chameau P, Inta D, Vitalis T, Monyer H, Wadman WJ and van Hooft JA (2009). The N-terminal region of reelin regulates postnatal dendritic maturation of cortical pyramidal neurons. *Proc Natl Acad Sci U S A.* 106 (17): 7227-7232.**
- The authors of this work report that 5-HT₃ activity affects embryonic cortical development by modulation of reelin. It is one of the first demonstrations of serotonin signaling through this receptor having a direct impact on neurogenesis in the mouse brain.*
58. Yip YP, Mehta N, Magdaleno S, Curran T and Yip JW (2009). Ectopic expression of reelin alters migration of sympathetic preganglionic neurons in the spinal cord. *J Comp Neurol.* 515: 260-268.
59. Vitalis T and Parnavelas JG (2003). The role of serotonin in early cortical development. *Dev Neurosci.* 25 (2): 245-256.
60. Eisensamer B, Rammes G, Gimpl G, Shapa M, Ferrari U, Hapfelmeyer G, Bondy B, Parsons C, Gilling K, Zieglgansberger W, Holsboer F and Rupprecht R (2003). Antidepressants are functional antagonists at the serotonin type 3 (5-HT₃) receptor. *Mol Psychiatry.* 8: 994-1007.
61. Smit-Rigter LA, Noorlander CW, von Oerthel L, Chameau P, Smidt MP and van Hooft JA (2012). Prenatal fluoxetine exposure induces life-long serotonin 5-HT₃ receptor-dependent cortical abnormalities and anxiety-like behaviour. *Neuropharmacology.* 62 (2): 865-870.
62. Emerit MB, Doucet E, Darmon M and Hamon M (2002). Native and cloned 5-HT_{3A}(S) receptors are anchored to F-actin in clonal cells and neurons. *Mol Cell Neurosci.* 20 (1): 110-124.
63. **Nichols RA and Mollard P (1996). Direct observation of serotonin 5-HT₃ receptor-induced increases in calcium levels in individual brain nerve terminals. *J Neurochem.* 67 (2): 581-592.**
- This paper demonstrates that 5-HT₃ receptors modulate calcium flux not only by its own channel activity, but also by stimulating other voltage-gated calcium channels found on the same synapses. This observation underlies the foundation of the hypothesis that 5-HT₃ activity regulates neuronal calcium flux in several ways that are conducive for instigating neurogenic programs during development.*
64. Nayak S, Ronde P, Spier A, Lummis SC and Nichols RA (1999). Calcium changes induced by presynaptic 5-hydroxytryptamine-3 serotonin receptors on isolated terminals from various regions of the rat brain. *Neurosci.* 91 (1): 107-117.
65. Reiser G, Donie F and Binmoller F-J (1989). Serotonin regulates cytosolic Ca²⁺ activity and membrane potential in a neuronal and in a glial cell line via 5-HT₃ and 5-HT₂ receptors by different mechanisms. *J Cell Sci.* 93: 545-555.
66. Yang J (1990). Ion permeation through 5-hydroxytryptamine-gated channels in neuroblastoma N18 cells. *J Gen Physiol.* 96 (6): 1177-1198.
67. Homma K, Kitamura Y, Ogawa H and Oka K (2006). Serotonin induces the increase in intracellular Ca²⁺ that enhances neurite outgrowth in PC12 cells via activation of 5-HT₃ receptors and voltage-gated calcium channels. *J Neurosci Res.* 84: 316-325.

CANDIDATE REVIEWS

68. Shieh PB, Hu S-C, Boob K, Timmusk T and Ghosh A (1998). Identification of a signaling pathway involved in calcium regulation of BDNF expression. *Neuron*. 20 (4): 727-740.
69. Tao X, Finkbeiner S, Arnold DB, Shaywitz AJ and Greenberg ME (1998). Ca²⁺ influx regulates BDNF transcription by a CREB family transcription factor-dependent mechanism. *Neuron*. 20 (4): 709-726.
70. Chen WG, Chang Q, Lin Y, Meissner A, West AE, Griffith EC, Jaenisch R and Greenberg ME (2003). Derepression of BDNF transcription involves calcium-dependent phosphorylation of MeCP2. *Science*. 302: 885-889.
71. Oostland M, Sellmeijer J and van Hooft JA (2011). Transient expression of function serotonin 5-HT₃ receptors by glutamatergic granule cells in the early postnatal mouse cerebellum. *J Physiol*. 589 (Pt 20): 4837-4846.
72. Johnson D and Heinemann S (1995). Embryonic expression of the 5-HT₃ receptor subunit, 5-HT_{3R-A}, in the rat: an in situ hybridization study. *Mol Cell Neurosci*. 6: 122-138.
73. Diez-Roux G, Banfi S, Sultan M, Geffers L, Anand S, Rozado D, Magen A, Canidio E, Pagani M, Peluso I, Lin-Marq N, Koch M, Bilio M, Cantiello I, Verde R, De Masi C, Bianchi SA, Cicchini J, Perroud E, Mehmeti S, Dagand E, Schrinner S, Nurnberger A, Schmidt K, Metz K, Zwingmann C, Brieske N, Springer C, Hernandez AM, Herzog S, Grabbe F, Sieverding C, Fischer B, Schrader K, Brockmeyer M, Dettmer S, Helbig C, Alunni V, Battaini MA, Mura C, Henrichsen CN, Garcia-Lopez R, Echevarria D, Puellas E, Garcia-Calero E, Kruse S, Uhr M, Kauck C, Feng G, Milyaev N, Ong CK, Kumar L, Lam M, Semple CA, Gyenesi A, Mundlos S, Radelof U, Lehrach H, Sarmientos P, Raymond A, Davidson DR, Dolle P, Antonarakis SE, Yaspo ML, Martinez S, Baldock RA, Eichele G and Ballabio A (2011). A high-resolution anatomical atlas of the transcriptome in the mouse embryo. *PLoS Biol*. 9 (1): e1000582.
74. Bhattacharya A, Dang H, Zhu QM, Schnegelsberg B, Rozengurt N, Cain G, Prantil R, Vorp DA, Guy N, Julius D, Ford AP, Lester HA and Cockayne DA (2004). Urothelial observations in mice expressing a constitutively active point mutation in the 5-HT_{3A} receptor subunit. *J Neurosci*. 24 (24): 5537-5548.
75. Gong S, Zheng C, Doughty ML, Losos K, Didkovsky N, Schambra UB, Nowak NJ, Joyner A, Leblanc G, Hatten ME and Heintz N (2003). A gene expression atlas of the central nervous system based on bacterial artificial chromosomes. *Nature*. 425 (6961): 917-925.
76. Zeitz KP, Guy N, Malmberg AB, Dirajlal S, Martin WJ, Sun L, Bonhaus DW, Stucky CL, Julius D and Basbaum AI (2002). The 5-HT₃ subtype of serotonin receptor contributes to nociceptive processing via a novel subset of myelinated and unmyelinated nociceptors. *J Neurosci*. 22 (3): 1010-1019.
77. Tian L, Akerboom J, Schreiter E and Looger L (2012). Neural activity imaging with genetically encoded calcium indicators. *Prog Brain Res*. 196: 79-94.

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The Role of the Principal Sensory Nucleus in Discriminative Touch

Eva Sawyer

Specialized facial somatosensory organs have evolved in diverse groups of animals, and the sense of touch that these organs transduce is important for normal behavior. The principal sensory nucleus of the spinal trigeminal complex is the first relay for facial discriminative touch in the central nervous system. Much of the work done on this nucleus is done in rodents, where the ability to trace the central representations of whisker follicle innervation has been a useful tool for experimenters. Questions remain about the role of the nucleus, from uncertainties about the basic anatomy to its role in forming the disproportionate representation of the body seen in the cortical somatosensory maps. Comparative neurobiology points out that some non-rodent animals with specialized trigeminal somatosensory organs, such as the star-nosed mole, have a much larger principal sensory nucleus than one would expect for a mammal of their size. Complementing rodent work with studies on these species has the potential to help resolve puzzles about the entire spinal trigeminal complex, and the principal sensory nucleus in particular.

Keywords: *Somatosensory, trigeminal, segmentation, principal sensory nucleus, Barrels, star-nosed mole*

Introduction

Facial somatosensory specializations help animals navigate their world. Examples include whiskers on the face of rodents and seals^{1,2}, corpuscles of Herbst and Grandry on the beak of ground-probing birds^{3,4}, push-rod receptors on the bills of montremes⁵, integumentary sense organs on the jaws of crocodylians⁶, and Eimer's organs on the noses of talpid moles⁷. These adaptations are associated with exploration, foraging and feeding^{8,9}. The co-evolution of sense organs and central processing centers is a theme in neurobiology^{10,11}. Accordingly, when researchers have looked at the first relay of the trigeminal somatosensory stream, these specialized trigeminal touch organs tend to be paired with central specializations¹²⁻¹⁵.

The spinal trigeminal complex (STC) is the main target for the primary somatosensory receptors innervating the scalp, face and oral structures. The complex consists of the principal sensory nucleus (PrV) at the most rostral position and the spinal trigeminal nucleus (STN) more caudally. The latter consists of three subnuclei with subnucleus pars oralis (SpVo), subnucleus pars interpolaris (SpVi), and subnucleus pars caudalis (SpVc) found at progressively more caudal positions, respectively^{16,17}. SpVc merges with the dorsal horn of the spinal cord at its most caudal extent.

The complex receives most of its sensory input from the somatosensory components of the trigeminal nerve (but also from the somatosensory components of the facial, glossopharyngeal and vagus nerves)¹⁶. Upon entering the brainstem, the trigeminal branch splits into an ascending branch that projects to the PrV and a descending branch that projects to the subdivisions of the STN.

Traditionally, there is a perception that in the somatosensory brainstem there is a division of labor so that the PrV mediates light touch sensation and the STN mediates pain¹⁸. In this view, the PrV is analogous to the dorsal column nuclei and the STN to the dorsal horn^{19,20}. Broadly, there is truth in the vital importance of the PrV to discriminative touch and the STN, especially the more caudal regions, to pain, but a strict view of non-converging labeled lines has weak support. This article will focus on the PrV, but it would be misleading to present the nucleus as if it were completely independent from the STN. Therefore, the STN will be mentioned where appropriate.

Form and Function

Ramón y Cajal illustrated trigeminal afferent axons branching to form an ascending path to the PrV and a descending path to the STN²¹. He writes that he cannot be sure that all of these axons bifurcate because he cannot rely on the

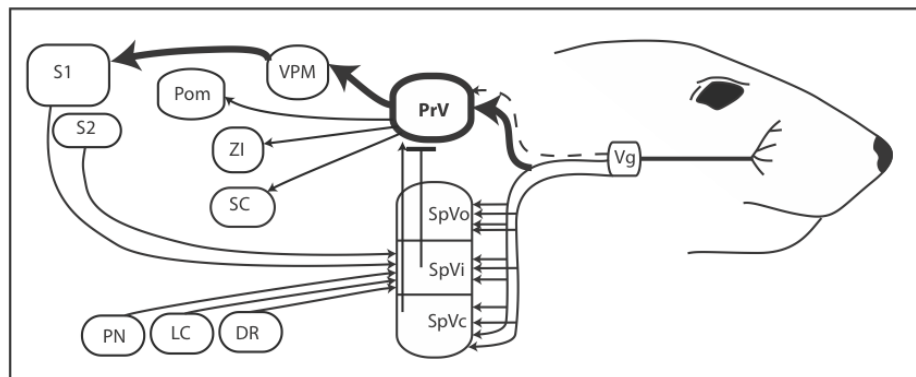


Figure 1: Schematic of main connections of the principal sensory nucleus (PrV) and the regulatory input directed through spinal trigeminal nucleus pars interpolaris (SpVi). The dotted line represents an unconfirmed class of sensory neurons projecting solely to the PrV. Thick lines represent the main pathway for low-threshold mechanoreception to the cortex. DR, dorsal raphe nucleus; LC, locus coeruleus; PN, Pontine nucleus; Pom, medial posterior nucleus; SC, superior colliculus; SpVc, spinal trigeminal nucleus pars caudalis; SpVo, spinal trigeminal nucleus pars oralis; ZI, zona incerta.

silver stain as an unbiased technique, but many, if not all, of the fibers he saw bifurcated. In a reinvestigation with silver stains Anstrom states he found the bifurcating fibers Ramón y Cajal reported, as well as descending non-bifurcating fibers projecting to only the STN²². He wrote that he did not observe, but could not rule out the possibility of, ascending non-bifurcating fibers projecting only to the PrV. Presumably, if these fibers exist, they would be mechanosensory axons that project exclusively to the PrV.

Another anatomist found such fibers. Windle, like Ramón y Cajal and Anstrom, used silver stains, but his studies were on fetal pigs instead of fetal and young mice²³. He found three sub-populations of trigeminal afferents: 52% of axons bifurcated, 42% (mostly thin fibers) descended without bifurcating, and 6% (mostly thick fibers) ascended without bifurcating. He struggled to explain why other investigators did not observe large diameter non-bifurcating ascending fibers. One possibility he does not mention is that mice, with vibrissa used for whisking, and pigs, with a large glabrous nose used for rooting in soil, differ in their facial somatosensory specializations. Since the mechanoreceptors in these specializations differ, the animals may have different proportions of bifurcating and non-bifurcating trigeminal afferents.

Subsequent literature on ascending non-bifurcating axons is sparse, though the perception that this pathway exists is maintained in modern reviews and texts²⁴⁻²⁷. One method for studying projection patterns of sensory neurons in the brainstem is interaxonal injections of tracer combined with reconstruction of the labeled axons. This has been used in the STC with interaxonal injections of the neuronal tracers horseradish peroxidase or neurobiotin into the spinal trigeminal tract. Unfortunately, the injections are almost always at the level of SpVo or SpVi²⁸⁻³³, a method which could not isolate an ascending non-bifurcating population. A less biased technique would use interaxonal injections

upstream of the bifurcation, as done by Shigenaga et al.³⁴, or to inject a far-reaching tracer into single ganglion cells, as Jaquin et al.³⁵ piloted in a methods paper. Neither study found ascending non-bifurcating axons. The absence of this class could be because such neurons are not present in rodents. However, given that neurons with the ascending non-bifurcating branching pattern made up only 6% of trigeminal afferents in pigs, a combined sample size of 12 axons in rodents is unlikely to represent sufficient sampling to warrant a strong conclusion that these cells are absent in rodents.

Within the PrV, SpVo and SpVi the somatotopic map of the face is inverted so that afferents from the mandibular branch project dorsally, the maxillary branch intermediately and the ophthalmic branch ventrally^{28,31}. The anterior receptors are represented medially and the more posterior receptors, laterally. The arrangement in the SpVc is less well understood. Some studies find that the dorsal-ventral representation is unchanged but the medial-lateral representation is flipped, so that the anterior receptors are represented laterally and the more posterior receptors, medially^{17,36-38}. They also find that in SpVc there is rostral-caudal skew that results in the more rostral afferents being represented more rostrally in the nucleus. This arrangement is reminiscent of the rostral-caudal mapping of dermatomes found in the dorsal horn for the rest of the body. Despite this work, reviews sometimes depict the SpVc as organized like the other subdivisions²⁶, and recently the somatotopy of SpVc has reemerged as an issue³⁷. It is noteworthy that there is still confusion about basic anatomy of the STC.

In addition to the main sensory input from primary sensory neurons, the PrV receives modulatory input from the STN and the cortex. Inhibitory GABAergic interneurons from the SpVi and excitatory glutamatergic interneurons from the SpVc project to the PrV⁴⁰. These connections let the STN influence the sensitivity of the PrV⁴¹. Projections

from the primary and secondary somatosensory areas (S1 and S2) to the STC could also facilitate top-down reduction of PrV sensitivity⁴²⁻⁴³. In rodents, the cortex-STN-PrV pathway is thought to be particularly important during active whisking, when the somatosensory signals induced by body movement, and not by the characteristics of a substrate, are irrelevant. Such a circuit could explain how the sensitivity of the PrV is reduced during active whisking⁴⁴. Other inputs are from the pontine tegmental nucleus⁴⁵⁻⁴⁶, the raphe nucleus⁴⁷ and the locus coeruleus⁴⁸. These likely reflect modulation of sensitivity based on the animal's level of alertness.

From the PrV, neurons project mainly to the contralateral ventral posterior medial nucleus (VPM) of the thalamus, which sends strong projections to S1^{12,49-51}. This trigeminal lemniscal pathway is particularly notable in rodents because every station on this pathway (the PrV, the VPM and S1) has a pattern of cell-dense patches that correspond in a one-to-one manner with the whiskers on the animal's snout, termed barrelettes¹⁷, barreloids⁵¹, and barrels¹² in each location, respectively. We will return to barrels later in the essay. Other important direct projections are to zona incerta⁵², the posterior medial nucleus of the thalamus⁴⁹ and the superior colliculus⁵³. These projections likely contribute to the regulation of movement. The main connections of the PrV are summarized in **Figure 1**.

Electrophysiological work in a variety of species complements the above anatomical findings. As would be expected from the termination patterns of large diameter bifurcating axons branching to every STC subdivision, the PrV and the STN contain mechanosensory neurons^{14,54-58}. Likewise, the anatomy shows that many small diameter fibers are non-bifurcating descending axons. If these are nociceptive c-fibers, then electrophysiological studies should find the STN enriched with nociceptive neurons. Indeed, electrophysiological studies that test for it fail to find nociceptive neurons in the PrV, but isolate them in STN^{54,56,59-61}.

Another promising area for animal studies is to use the power of genetic manipulations in model species to dissect the pathways of mechanoreceptors. For example, Li et al.⁶² drove expression of reporter proteins in different classes of low-threshold mechanoreceptors in order to follow sensory neurons from the receptors in the skin to their projections in the dorsal horn. The same techniques are yet to be applied to neurons projecting to the dorsal column nuclei or to the STC. Both studies would be valuable. In the whisker pathway, it would be interesting to see if the

different classes of low-threshold mechanoreceptors have unique projection patterns to the subdivisions of the STC, which has so far been undetected with electrophysiology and tract tracing.

Human case studies provide strong results that support the view of parallel pain and touch pathways. Lesions in the PrV cause deficits in touch sensation with sparing of temperature sense and nociception in the face⁶³, while lesions in the SpVc spare touch sensation but usually lead to the loss of nociception and temperature perception¹⁸. Thus, surgical damage to this area is a treatment for intractable orofacial pathogenic pain. With case studies such as these providing most of the background for the understanding of the human trigeminal system, a recent fMRI study was surprising. The study looked for changes in the blood oxygen level in humans who were experiencing noxious and non-noxious cutaneous and muscle stimulation to their face. As expected, noxious cutaneous and noxious muscle stimulation elicited changes in regions of the STN, but unexpectedly, the noxious muscle stimulation also elicited activation in PrV⁶⁴.

These unexpected results showing integration of pain and sensory information in the brainstem fit with a history of confusion about trying to connect the anatomical data, the electrophysiological data and now the fMRI data — which all show some integration of low-threshold mechanoreception and nociception — with human case studies, which show a strong division of touch and pain sensation between the nuclei^{18,22,64}. One problem is that lesion studies rely on eliminating an entire node of a network, which provides strong but crude results. Another problem is that there is a push to make labeled lines for pain and touch explain the anatomical divisions of the STC, despite evidence that the divisions will not fit well into those categories. That debate, however, is beyond the scope of this article (see ⁶⁵ for review).

Comparative neurobiology

Rodents have been seen as well suited for studies of the trigeminal touch pathway because their barrel system is more amenable to experimentation than non-patterned areas. The cortical barrels are impressive. For example, in rats, barrels cover 20% of S1, a total area of 9 mm²⁶⁶. The development of this pattern is dependent on an intact PrV⁶⁷. As others have pointed out, it is odd that all the input for such a large cortical representation is funneled through a small PrV⁵⁴, 0.56mm³ in the case of a rat¹³. Part of the resolution is that the cortex also receives connections from the

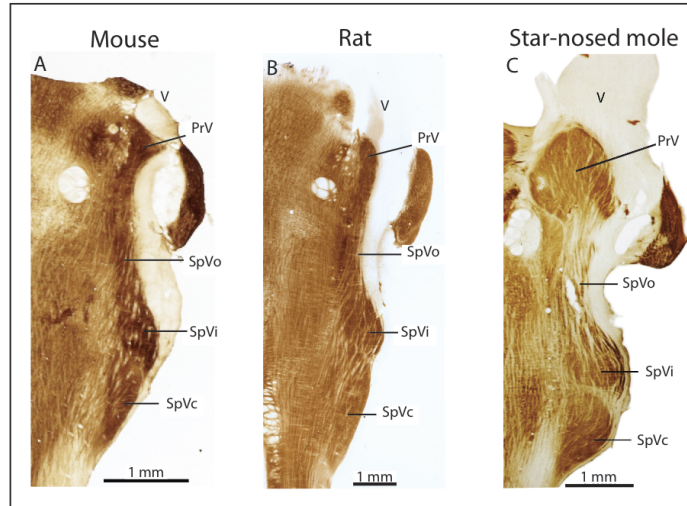


Figure 2: *Cytochrome oxidase stained sections of a mouse, rat and star-nosed mole brainstem cut in the horizontal plane. Chosen sections maximized the volume of the principal sensory nucleus (PrV). Compared to the mouse and the rat, the star-nosed mole has a large PrV. SpVc, spinal trigeminal pars caudalis; SpVi, spinal trigeminal pars interpolaris; SpVo, spinal trigeminal pars oralis.*

STN. Specifically, the caudal portions of SpVi project to the ventrolateral portion of the VPM in the thalamus. The VPM projects to the inter-barrel space, termed septa, in the cortex⁶⁸.

In rats, the volume of the SpVi is 1.66mm³, almost three times larger than the PrV, and has more distinct barrelettes than the PrV^{17,69}. Earlier we saw that the SpVi has a role in modulating sensory sensitivity based on directed movements of the whiskers. A large part of rodent exploratory behavior is active whisking — coordinated movements of the six muscles innervating each whisker pad to move the whiskers against the surface being examined^{2,70-72}. All this suggests that rats and other rodents are good models for studying the SpVi, particularly because the subnucleus relates to active sensory behavior. But other animals may be better suited for investigating the PrV.

Comparative studies point out that in some species with elaborate somatosensory trigeminal sensory organs, the PrV is hypertrophied^{9,14}. The case of the star-nosed mole is particularly informative because of the amount already known about its nervous system. The star consists of 22 fleshy appendages covered with Eimer's organs. Eimer's organs are composed of regular geometric arrangements of Merkel cell-neurite complexes, laminated corpuscles and free nerve-endings⁷. The star can be moved forward as a whole, and groups of appendages can be extended to bring the organ surface into contact with a substrate⁷³. When moles forage, they rapidly move the star, touching it to the surfaces of the damp soil of their habitat to locate small food items in the mud⁷⁴. There is a behavioral preference to use the two medial ventral rays when inspecting potential food items⁷⁵. The neuroanatomical correlate of that prefer-

ence is a larger representation of that ray in S1, and smaller receptive field size in the representation of the medial ventral rays than in the other rays^{73,76}. The behavioral preference and the increased resolution suggest those rays as a somatosensory analog of the retina's fovea⁷⁵.

To put the size of the star-nosed mole PrV into perspective, the absolute volume of the PrV of a 55g star-nosed mole is larger than the PrV of a 274g rat — it is about 630% the size expected based on the proportions of a rat¹³⁻¹⁴. In comparison, the SpVi subdivision of the star-nosed mole is only about 50% larger than expected⁹ (unpublished result). Acknowledging that the comparisons are crude and cover a wide taxonomic range, the results still show that the star-nosed mole has a large PrV even when compared to other somatosensory specialists (**Figure 2**).

The size of the PrV in the star-nosed mole is likely related to the exceptional spatial resolution of the star. Multi-unit receptive fields in the cortex average 0.82 mm² in the non-foveal part of the star and 0.52 mm² in the foveal regions, which are both smaller than receptive fields reported for primate fingertips⁷⁶⁻⁷⁸. If the PrV is the nucleus for fine touch, the extraordinary resolution of the star would be expected to distinguish this nucleus. There is already evidence that this is the case: within the PrV, as in the cortex, the medial ventral rays have a larger representation than the other rays. Interestingly, the greater size of the representation of the foveal rays in the cortex and PrV is not explained by greater innervation of these rays¹⁴. Combined with the smaller receptive fields in foveal than in non-foveal areas in S1, this suggests that within the lemniscal pathway foveal afferents converge less than the afferents for other rays.

Finding that a behaviorally important area of skin is over-represented in the central nervous system is not new⁷⁹. But finding that the size of the somatosensory representation cannot be predicted by counting the number of fibers innervating that structure and multiplying by a constant “afferent scaling factor” is special⁸⁰. This result is important because it suggests the mole PrV, and perhaps the rest of the lemniscal pathway, could be used to address questions about how the central over-representation of a foveal area of a sensory epithelium comes about.

Conclusion

The role of the PrV, compared to other regions of the STC is relatively understudied. The oversight is surprising given its vital role in organizing the somatosensory cortex. The lack of focus on the PrV might be due to the relatively unimpressive PrV in rodents compared to other trigeminal somatosensory specialists. There are many unresolved questions that could be addressed with comparative work. Just some include: Anatomically, what contributes to a hypertrophied PrV? Are there unique afferents? Is there less convergence? Within a nucleus, what contributes to the “foveal” area of higher resolution? Understanding these points will inform us on the forces that link the evolution of sensory surfaces and their central representations.

References

1. Dehnhardt G, Mauck B, Hanke W and Bleckmann H (2001). Hydrodynamic trail-following in harbor seals (*Phoca vitulina*). *Science*. 293 (5527): 102-104.
2. Ahl AS (1986). The role of vibrissae in behavior: a status review. *Vet Res Commun*. 10 (4): 245-268.
3. Dubbeldam JL (1980). Studies on the somatotopy of the trigeminal system in the mallard, *Anas platyrhynchos* L. II. Morphology of the principal sensory nucleus. *J Comp Neurol*. 191 (4): 557-571.
4. Pettigrew JD and Frost BJ (1985). A tactile fovea in the Scolopacidae? *Brain Behav Evol*. 26 (3-4): 105-195.
5. Proske U, Gregory JE and Iggo A (1998). Sensory receptors in monotremes. *Philos Trans R Soc Lond B Biol Sci*. 353 (1372): 1187-1198.
6. Leitch DB and Catania KC (2012). Structure, innervation and response properties of integumentary sensory organs in crocodylians. *J Exp Biol*. 215 (Pt 23): 4217-4230.
7. Catania KC (1996). Ultrastructure of the Eimer's organ of the star-nosed mole. *J Comp Neurol*. 365 (3): 343-354.
8. Catania KC (2005). Evolution of sensory specializations in insectivores. *Anat Rec A Discov Mol Cell Evol Biol*. 287 (1): 1038-1050.
9. Gutierrez-Ibanez C, Iwaniuk AN and Wylie DR (2009). The independent evolution of the enlargement of the principal sensory nucleus of the trigeminal nerve in three different groups of birds. *Brain Behav Evol*. 74 (4): 280-294.
10. Van der Loos H and Dorfl J (1978). Does the skin tell the somatosensory cortex how to construct a map of the periphery? *Neurosci Lett*. 7 (1): 23-30.
11. Krubitzer LA and Seelke AMH (2012). Cortical evolution in mammals: The bane and beauty of phenotypic variability. *Proc Natl Acad Sci U S A*. 109: 10647-10654.
12. Woolsey TA and Van der Loos H (1970). The structural organization of layer IV in the somatosensory region (SI) of mouse cerebral cortex. The description of a cortical field composed of discrete cytoarchitectonic units. *Brain Res*. 17 (2): 205-242.
13. Ashwell KW, Hardman CD and Paxinos G (2006). Cyto- and chemoarchitecture of the sensory trigeminal nuclei of the echidna, platypus and rat. *J Chem Neuroanat*. 31 (2): 81-107.
14. **Catania KC, Leitch DB and Gauthier D (2011). A Star in the Brainstem Reveals the First Step of Cortical Magnification. *Plos One*. 6 (7).**
This article reports on a star representation in the hypertrophied principal sensory nucleus of the star-nosed mole. Importantly, the authors find that the size of the behaviorally important “fovea” rays is magnified at the level of the brainstem.
15. Sarko DK, Johnson JI, Switzer RC, 3rd, Welker WI and Reep RL (2007). Somatosensory nuclei of the manatee brainstem and thalamus. *Anat Rec (Hoboken)*. 290 (9): 1138-1165.
16. Olszewski J (1950). On the anatomical and functional organization of the spinal trigeminal nucleus. *J Comp Neurol*. 92 (3): 401-413.
17. Ma PM (1991). The barrelettes--architectonic vibrissal representations in the brainstem trigeminal complex of the mouse. I. Normal structural organization. *J Comp Neurol*. 309 (2): 161-199.
18. Gerard MW (1923). Afferent impulses of the trigeminal nerve. *AMA Arch Neurol Psychiat*. 9: 306-338.
19. Darian-Smith I, Phillips G and Ryan RD (1963). Functional Organization in the Trigeminal Main Sensory and Rostral Spinal Nuclei of the Cat. *J Physiol*. 168: 129-146.
20. Dubner R and Bennett GJ (1983). Spinal and Trigeminal Mechanisms of Nociception. *Annual Review of Neuroscience*. 6: 381-418.
21. Ramon y Cajal S (1896). Beitrag zum Studium der Medulla oblongata, des Kleinhirns und des Ursprungs der Gehirnnerven. Leipzig: J. A. Barth.
22. Anstrom KE (1953). On the central course of afferent fibres in the trigeminal, facial, glossopharyngeal, and vagal nerves and their nuclei in the mouse. *Acta physiol scand Suppl*. 106: 209-320.
23. **Windle WF (1926). Non-bifurcating nerve fibers of the trigeminal nerve. *The Journal of Comparative Neurology* 40 (1): 229-240.**

CANDIDATE REVIEWS

- This article reports on the projection patterns of trigeminal sensory afferents. Windle finds non-bifurcating ascending axons, an observation which is the basis for the modern perception that such axons exist. The finding has not been replicated, though it is not clear that the hypothesis that these axons exist has been well tested.*
24. Butler AB and Hodos W (2005). Comparative vertebrate neuroanatomy : evolution and adaptation, 2nd Edition. Hoboken, N.J.: Wiley-Interscience.
 25. Martin JH (2003). Neuroanatomy : text and atlas, 3rd Edition. New York, N.Y.: McGraw-Hill.
 26. Bosman LW, Houweling AR, Owens CB, Tanke N, Shevchouk OT, Rahmati N, Teunissen WH, Ju C, Gong W, Koekkoek SK and De Zeeuw CI (2011). Anatomical pathways involved in generating and sensing rhythmic whisker movements. *Front Integr Neurosci.* 5: 53.
 27. Usunoff KG, Marani E and Schoen JH (1997). The trigeminal system in man. *Adv Anat Embryol Cell Biol.* 136: I-X, 1-126.
 28. Hayashi H (1980). Distributions of vibrissae afferent fiber collaterals in the trigeminal nuclei as revealed by intra-axonal injection of horseradish peroxidase. *Brain Res.* 183 (2): 442-446.
 29. Hayashi H (1985). Morphology of terminations of small and large myelinated trigeminal primary afferent fibers in the cat. *J Comp Neurol.* 240 (1): 71-89.
 30. Jacquin MF, Renehan WE, Rhoades RW and Panneton WM (1993). Morphology and topography of identified primary afferents in trigeminal subnuclei principalis and oralis. *J Neurophysiol.* 70 (5): 1911-1936.
 31. Shortland PJ, Demaro JA, Shang F, Waite PM and Jacquin MF (1996). Peripheral and central predictors of whisker afferent morphology in the rat brainstem. *J Comp Neurol.* 375 (3): 481-501.
 32. Chiaia NL, Hess PR, Hosoi M and Rhoades RW (1987). Morphological characteristics of low-threshold primary afferents in the trigeminal subnuclei interpolaris and caudalis (the medullary dorsal horn) of the golden hamster. *J Comp Neurol.* 264 (4): 527-546.
 33. Tsuru K, Otani K, Kajiyama K, Suemune S and Shigenaga Y (1989). Central terminations of periodontal mechanoreceptive and tooth pulp afferents in the trigeminal principal and oral nuclei of the cat. *Brain Res.* 485 (1): 29-61.
 34. Shigenaga Y, Otani K and Suemune S (1990). Morphology of central terminations of low-threshold trigeminal primary afferents from facial skin in the cat--intra-axonal staining with HRP. *Brain Res.* 523 (1): 23-50.
 35. Jacquin MF, Hu JW, Sessle BJ, Renehan WE and Waite PM (1992). Intra-axonal Neurobiotin injection rapidly stains the long-range projections of identified trigeminal primary afferents in vivo: comparisons with HRP and PHA-L. *J Neurosci Methods.* 45 (1-2): 71-86.
 36. Arvidsson J (1982). Somatotopic organization of vibrissae afferents in the trigeminal sensory nuclei of the rat studied by transganglionic transport of HRP. *J Comp Neurol.* 211 (1): 84-92.
 37. da Silva S, Hasegawa H, Scott A, Zhou X, Wagner AK, Han BX and Wang F (2011). Proper formation of whisker barrelettes requires periphery-derived Smad4-dependent TGF-beta signaling. *Proc Natl Acad Sci U S A.* 108 (8): 3395-3400.
 38. Jacquin MF, Renehan WE, Mooney RD and Rhoades RW (1986). Structure-function relationships in rat medullary and cervical dorsal horns. I. Trigeminal primary afferents. *J Neurophysiol.* 55 (6): 1153-1186.
 39. Sjöqvist O (1938). Studies on pain conduction in the trigeminal nerve; a contribution to the surgical treatment of facial pain. Helsingfors: Mercators tryckeri.
 40. Furuta T, Timofeeva E, Nakamura K, Okamoto-Furuta K, Togo M, Kaneko T and Deschenes M (2008). Inhibitory gating of vibrissal inputs in the brainstem. *J Neurosci.* 28 (8): 1789-1797.
 41. Timofeeva E, Lavallee P, Arsenault D and Deschenes M (2004). Synthesis of multiwhisker-receptive fields in subcortical stations of the vibrissa system. *J Neurophysiol.* 91 (4): 1510-1515.
 42. Aronoff R, Matyas F, Mateo C, Ciron C, Schneider B and Petersen CC (2010). Long-range connectivity of mouse primary somatosensory barrel cortex. *Eur J Neurosci.* 31 (12): 2221-2233.
 43. Haque T, Akhter F, Kato T, Sato F, Takeda R, Higashiyama K, Moritani M, Bae YC, Sessle BJ and Yoshida A (2012). Somatotopic direct projections from orofacial areas of secondary somatosensory cortex to trigeminal sensory nuclear complex in rats. *Neuroscience.* 219: 214-233.
 44. Lee S, Carvell GE and Simons DJ (2008). Motor modulation of afferent somatosensory circuits. *Nat Neurosci.* 11 (12): 1430-1438.
 45. Timofeeva E, Dufresne C, Sik A, Zhang ZW and Deschenes M (2005). Cholinergic modulation of vibrissal receptive fields in trigeminal nuclei. *J Neurosci.* 25 (40): 9135-9143.
 46. Beak SK, Hong EY and Lee HS (2010). Collateral projection from the forebrain and mesopontine cholinergic neurons to whisker-related, sensory and motor regions of the rat. *Brain Res.* 1336: 30-45.
 47. Lee SB, Lee HS and Waterhouse BD (2008). The collateral projection from the dorsal raphe nucleus to whisker-related, trigeminal sensory and facial motor systems in the rat. *Brain Res.* 1214: 11-22.
 48. Moore RY and Bloom FE (1979). Central catecholamine neuron systems: anatomy and physiology of the norepinephrine and epinephrine systems. *Annual Review of Neuroscience.* 2: 113-168.
 49. Veinante P and Deschenes M (1999). Single- and multi-whisker channels in the ascending projections from the principal trigeminal nucleus in the rat. *J Neurosci.* 19 (12): 5085-5095.
 50. Furuta T, Kaneko T and Deschenes M (2009). Septal neurons in barrel cortex derive their receptive field input from the lemniscal pathway. *J Neurosci.* 29 (13): 4089-4095.
 51. Van der Loos H (1976). Barreloids in mouse somatosensory thalamus. *Neurosci Lett.* 2 (1): 1-6.
 52. Kolmac CI, Power BD and Mitrofanis J (1998). Patterns of connections between zona incerta and brainstem in rats. *J Comp Neurol.* 396 (4): 544-555.
 53. Steindler DA (1985). Trigemino-cerebellar, trigemino-tectal, and trigeminothalamic projections: a double retrograde

- axonal tracing study in the mouse. *J Comp Neurol.* 237 (2): 155-175.
54. Kirkpatrick DB and Kruger L (1975). Physiological properties of neurons in the principal sensory trigeminal nucleus of the cat. *Exp Neurol.* 48 (3 Pt 1): 664-690.
55. Zeigler HP and Witkovsky P (1968). The main sensory trigeminal nucleus in the pigeon: a single-unit analysis. *J Comp Neurol.* 134 (3): 255-264.
56. Gordon G, Landgren S and Seed WA (1961). The functional characteristics of single cells in the caudal part of the spinal nucleus of the trigeminal nerve of the cat. *J Physiol.* 158: 544-559.
57. Silver R and Witkovsky P (1973). Functional characteristics of single units in the spinal trigeminal nucleus of the pigeon. *Brain Behav Evol.* 8 (4): 287-303.
58. Molenaar GJ, Fizaan-Oostveen JL and van der Zalm JM (1979). Infrared and tactile units in the sensory trigeminal system of python reticulatus. *Brain Res.* 170 (2): 372-376.
59. Azerad J, Woda A and Albe-Fessard D (1982). Physiological Properties of neurons in different parts of the cat trigeminal sensory complex. *Brain Res.* 246 (1): 7-21.
60. Mosso JA and Kruger L (1972). Spinal trigeminal neurons excited by noxious and thermal stimuli. *Brain Res.* 38 (1): 206-210.
61. Dallel R, Clavelou P and Woda A (1989). Effects of tractotomy on nociceptive reactions induced by tooth pulp stimulation in the rat. *Exp Neurol.* 106 (1): 78-84.
62. **Li LS, Rutlin M, Abraira VE, Cassidy C, Kus L, Gong SC, Jankowski MP, Luo WQ, Heintz N, Koerber HR, Woodbury CJ and Ginty DD (2011). The Functional Organization of Cutaneous Low-Threshold Mechanosensory Neurons. *Cell.* 147 (7): 1615-1627.**
- The authors use transgenic mice and immunohistochemistry to selectively label 4 classes of low-threshold mechanoreceptors. This allows them to trace the peripheral and central projections of somatosensory neurons, a goal that has been largely elusive in somatosensory research.*
63. Kamitani T, Kuroiwa Y and Hidaka M (2004). Isolated hypesthesia in the right V2 and V3 dermatomes after a midpontine infarction localised at an ipsilateral principal sensory trigeminal nucleus. *J Neurol Neurosurg Psychiatry.* 75 (10): 1508-1509.
64. Nash PG, Macefield VG, Klineberg IJ, Murray GM and Henderson LA (2009). Differential activation of the human trigeminal nuclear complex by noxious and non-noxious orofacial stimulation. *Hum Brain Mapp.* 30 (11): 3772-3782.
65. Craig AD (2003). Pain mechanisms: labeled lines versus convergence in central processing. *Annual Review of Neuroscience.* 26: 1-30.
66. Welker C (1971). Microelectrode delineation of fine grain somatotopic organization of (SmI) cerebral neocortex in albino rat. *Brain Res.* 26 (2): 259-275.
67. Killackey HP and Fleming K (1985). The role of the principal sensory nucleus in central trigeminal pattern formation. *Brain Res.* 354 (1): 141-145.
68. Pierret T, Lavallee P and Deschenes M (2000). Parallel streams for the relay of vibrissal information through thalamic barreloids. *J Neurosci.* 20 (19): 7455-7462.
69. Lo FS, Guido W and Erzurumlu RS (1999). Electrophysiological properties and synaptic responses of cells in the trigeminal principal sensory nucleus of postnatal rats. *J Neurophysiol.* 82 (5): 2765-2775.
70. Vincent SB (1912). The function of vibrissae in the behavior of the white rat. *Behav Monogr.* 1: 1-82.
71. Dorfl J (1982). The musculature of the mystacial vibrissae of the white mouse. *J Anat.* 135 (Pt 1): 147-154.
72. Hartmann MJ (2011). A night in the life of a rat: vibrissal mechanics and tactile exploration. *Ann N Y Acad Sci.* 1225: 110-118.
73. Catania KC and Kaas JH (1995). Organization of the somatosensory cortex of the star-nosed mole. *J Comp Neurol.* 351 (4): 549-567.
74. Catania KC and Remple FE (2005). Asymptotic prey profitability drives star-nosed moles to the foraging speed limit. *Nature.* 433 (7025): 519-522.
75. Catania KC and Kaas JH (1997). Somatosensory fovea in the star-nosed mole: behavioral use of the star in relation to innervation patterns and cortical representation. *J Comp Neurol.* 387 (2): 215-233.
76. Sachdev RN and Catania KC (2002). Receptive fields and response properties of neurons in the star-nosed mole's somatosensory fovea. *J Neurophysiol.* 87 (5): 2602-2611.
77. Pons TP, Wall JT, Garraghty PE, Cusick CG and Kaas JH (1987). Consistent features of the representation of the hand in area 3b of macaque monkeys. *Somatosens Res.* 4 (4): 309-331.
78. Xerri C, Merzenich MM, Peterson BE and Jenkins W (1998). Plasticity of primary somatosensory cortex paralleling sensorimotor skill recovery from stroke in adult monkeys. *J Neurophysiol.* 79 (4): 2119-2148.
79. Penfield W and Rasmussen T (1950). The cerebral cortex of man; a clinical study of localization of function. New York,: Macmillan.
80. **Welker E and Van der Loos H (1986). Quantitative correlation between barrel-field size and the sensory innervation of the whiskerpad: a comparative study in six strains of mice bred for different patterns of mystacial vibrissae. *J Neurosci.* 6 (11): 3355-3373.**
- This article finds tests the relationship between whisker innervation and cortical barrel area in six strains of mice bred for their different whisker patterns. The authors find that within a strain there is a linear relationship between the variables, but the slope of the relationship differs between strains. The existence of a linear relationship can be contrasted with case in the star-nosed mole.*

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Investigating Atypical Multisensory Processing in Individuals with Autism Spectrum Disorders

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Blending information from multiple senses together into a perceptual Gestalt is necessary to understand the world. Autism spectrum disorders (ASD) are classically defined by a triad of symptoms, although sensory impairments have been consistently reported as well. Recently, there has been an increased interest in how individuals with ASD combine and integrate information from multiple senses. This article discusses current topics in ASD including neurobiology and sensory impairments with a focus on how atypical processing of multisensory information may be related to the symptoms found in ASD.

Keywords: *Autism, sensory, multisensory integration, temporal processing, audiovisual*

Functional connectivity:

A process used to describe how well brain regions are connected based on the temporal synchronization of the activity between these areas.

Autism Spectrum Disorders: Overview

Autism spectrum disorders are complex neurodevelopmental disorders characterized by repetitive or restricted behaviors, impairments in language and communication, and deficits in social or reciprocal behavior¹. Currently, it is estimated that 1 in 88 individuals is diagnosed with an ASD and that males are 4 times more likely to be diagnosed than females². Diagnoses of ASD normally occur by 3 years of age, but clinical symptoms may be present earlier in development. Based on the DSM-IV, Autism, Asperger's syndrome, and Pervasive Developmental Disorder Not Otherwise Specified (PDD-NOS) are three disorders that fall within this spectrum. Individuals with autism can be further characterized as either classic or high functioning based on IQ measures. Asperger's syndrome and PDD-NOS differ from autism in that individuals with Asperger's do not have delays in communication and individuals with PDD-NOS either lack impairments in communication or repetitive behaviors or present mild forms of the triad of symptoms³. While DSM-IV classification is currently in use it should be recognized that the proposed draft of the DSM-V modifies the definition of ASD.

Neurobiological and Neurophysiological

Findings in ASD

In order to investigate possible causes for the symptoms found in autism spectrum disorders, there have been a variety of studies that have focused on determining the neurobiology of ASD. Autism has been characterized by multiple descriptions of atypical structural and functional connections within the brain. A consistent finding is that children with autism tend to have larger brains and greater head circumferences that increase in the first few years of development, then reaches a plateau and growth slows later in life^{4, 5}. It is thought that this overgrowth occurs during a critical time period in the development of the frontal and temporal lobes, but does not seem to impact the occipital lobes⁶. This thought stems from atypical morphology demonstrated by an increase in cerebral grey and white matter in the frontal and temporal regions⁷. Another developmental finding in ASD is altered minicolumnar structure. A cortical minicolumn is the fundamental unit in the cortex that is comprised of excitatory pyramidal cells surrounded by inhibitory interneurons⁸. Post-mortem studies revealed that in the frontal and temporal lobes, there were increased numbers of minicolumns, pyramidal neurons in the minicolumnar structure, and the overall structure was narrower in individuals with

autism^{8,9}. More excitatory pyramidal neurons along with a greater number of narrower minicolumns could result in over activation of neurons, leading to local cortical connections more likely and long range connections less likely to develop¹⁰. These neurobiological and structural findings prompted the investigation of possible altered functional connections between specific brain regions in ASD. There have been numerous studies demonstrating a reduced functional connectivity between multiple brain regions in individuals with ASD¹¹⁻¹⁷. It was shown that the functional connectivity between Broca and Wernicke's areas, two regions used for speech, was reduced in individuals with ASD compared to typically developed controls¹⁸. Based on impaired long range connections between regions used for speech processing, this finding provides neurological evidence for possible deficits in communication in ASD.

Autism Spectrum Disorders: Theories

In order to explain the neurobiology along with the symptoms found in this heterogeneous disorder a multitude of theories have been proposed. Weak central coherence, the temporal binding deficit hypothesis, the cortical underconnectivity theory, and an imbalance of excitation/inhibition signaling are relevant theories in explaining disturbances found in ASD¹⁹.

Central coherence is based on the concept that individuals are capable of processing and combining information for a higher level of understanding²⁰. The theory of weak central coherence (WCC) states that individuals with autism have impairments in integrating information from a local or detailed perspective to a more global concept²¹. If there is a preference for local over global processing, this may explain aspects of restricted interests and could cause the global meaning of social situations and communication to be impaired. Brock's temporal binding deficit hypothesis tried to explain weak central coherence by looking at potential timing deficits. This theory proposed that long range connections between brain regions may not be as temporally correlated as brain regions that are in closer proximity, which could result in intact local, but impaired global processing²². The cortical underconnectivity theory explained this concept in more detail, when functional magnetic resonance imaging (fMRI) studies demonstrated that long distance connections between brain regions were less functionally connected in individuals with autism^{18,23}. The cortical underconnectivity theory has been consistently used to describe atypical processing found in ASD.

Just *et al* proposed this theory by using an fMRI sentence comprehension task to demonstrate atypical functional connectivity in individuals with ASD¹⁸. This theory complements the previous two theories because it provides a possible reason for global information processing deficits since this type of processing requires the proper timing and integration of information from multiple brain regions²⁴. A final explanation is a possible imbalance of the ratio of excitation/inhibition signaling in the brain. This is supported by post-mortem studies in ASD patients demonstrating an increased number of excitatory pyramidal cells^{4,10,25}. While no one theory can explain all of the symptoms, there are aspects from all of these theories that may lend to greater insights in explaining the impairments that are seen in individuals with ASD.

Sensory Impairments in ASD

In addition to the three classical symptoms that impact individuals with autism, sensory abnormalities have been consistently observed and reported. Sensory dysfunction is not one of the classic symptoms of autism spectrum disorders, yet it is found to impact up to 85% of individuals with ASD²⁶. Kanner first reported children with autism spectrum disorders to have sensory disruptions and to date there have been a number of studies demonstrating atypical sensitivity to sensory stimuli^{26,27}. Individuals can be hyper- or hyposensitive to a variety of different stimuli spanning multiple modalities^{28,29}. Using a visual stimulus as an example, individuals with ASD may immediately cover their eyes or stare at bright lights for long periods of time, depending on the individual's sensitivity to the stimulus. This dysfunction can be further classified as either sensory aversion or sensory seeking behavior. Depending on the type of behavior, individuals with autism spectrum disorders may be highly sensitive to or engaged in stimuli that may be considered mundane. Based on these observations, there have been several studies that have correlated sensory impairments with ASD severity³⁰⁻³³. Of all the sensory modalities impacted, dysfunctions in the visual and auditory domains have been studied most extensively. One reason for why this may be the case is that these modalities are needed for communication as well as understanding aspects of social behavior. A consistent finding is that individuals with ASD tend to perform well on visual tasks that require the processing of individual features^{34,35}. For example, individuals with ASD can excel on the Embedded Figures and Block Design tasks which are timed and require the identification of smaller objects within a larger figure or the recreation

of a detailed pattern using a set of blocks^{20, 21}, respectively. While there are reports of enhanced performance on tasks requiring the identification of specific features there have been observations demonstrating impaired global processing when these local features are needed to be combined. For instance, individuals with autism typically are less accurate in identifying biological motion, motion coherence, and visual form compared to controls³⁶⁻³⁸. An interesting finding from these studies however, was that while the high functioning autism group showed deficits in these areas, individuals with Asperger's syndrome performed as well as typically developed individuals, which speaks to the heterogeneity of ASD³⁷⁻³⁸. Similar to the findings in the visual domain, it is known that depending on the task, individuals with ASD perform atypically compared to controls in the auditory domain as well³⁹. For example, it has been shown that individuals with ASD tend to excel on tasks that require pitch discrimination specifically of musical tones^{40, 41}.

Enhanced visual and auditory discrimination to detailed information, yet overall impaired global processing in these modalities are consistent findings in ASD. Communication and behaving in social contexts require a high level of global processing and if individuals with ASD have deficits in this type of processing this could result in information being lost or improperly understood. Besides the possible impaired processing, both communication and social behaviors require the appropriate combination of sensory information from multiple modalities. Based on these observations and studies describing unisensory dysfunction in ASD, there has been an increased focus in determining how multisensory integration may be impacted when sensory stimuli are combined.

Multisensory Integration and Temporal Processing

Multisensory integration can be described as the merging of sensory information from different modalities⁴². Studies have shown that the combination of information from multiple senses can produce behavioral enhancements that can increase accuracy and reduce reaction times⁴³. Three principles: space, time, and effectiveness are used to describe multisensory integration⁴³. Stimuli from different modalities that are presented in close temporal and/or spatial proximity can result in a maximal multisensory gain^{44, 45}. The enhancement seen in behavior from the combination of multiple stimuli compared to one of the sensory stimuli presented alone describes multisensory gain⁴³. One effective way to measure multisensory integration is by us-

ing cross modal illusions. These illusions can produce interesting behavioral responses based on how one modality affects another. For example, the sound induced flash illusion demonstrates that the auditory domain can alter the visual information that is perceived by individuals⁴⁶. In this illusion when two auditory beeps are played in close temporal proximity to a single visual flash, most individuals perceive multiple visual flashes^{46, 47}. Also, it was shown that as the presentation of the second beep is further delayed in time, participants are less likely to perceive the illusion⁴⁸. Therefore, this illusion not only utilizes the temporal principle of multisensory integration, but also demonstrates the concept of a multisensory temporal binding window (TBW). The multisensory TBW can be described as the time interval in which two cross modal stimuli are bound together as a single unified perceived event⁴⁹.

It was demonstrated that individuals with ASD performed comparable to typically developed controls for the sound induced illusion, which illustrates that there is intact multisensory integration for simple cross modal stimuli⁴⁹. However, using this illusion, it appears that children with ASD have an extended temporal binding window^{50, 51}. Foss-Feig *et al* showed that individuals with ASD perceived this illusion more often when the presentations of the second beep were delayed for longer periods of time⁵¹. This study demonstrated a multisensory temporal binding window for individuals with ASD that was almost twice as wide as that of typically developed controls. An extended TBW and overall atypical temporal processing could impact communication and the proper understanding of social behaviors, both of which are known to be impaired in autism spectrum disorders. Importantly, it is known that the multisensory temporal binding windows tend to be wider for children in general and it is possible to narrow the TBW with the implementation of multisensory training paradigms^{52, 53}. The plasticity of the TBW may allow for future developments of remediation tools for individuals with ASD.

In general, there have been a variety of studies supporting the findings of temporal deficits when individuals with ASD are asked to process multisensory information^{54, 55}. Bebko *et al* simultaneously showed children identical videos on two monitors with the only difference being that one of the video tracks was temporally delayed⁵⁵. This study demonstrated that while typically developed children preferentially looked at the videos that were synchronous, children with autism did not have a preference looking at either video, specifically when the stimuli were

speech related. This suggests that the ASD group may not have noticed that one of the videos was temporally out of synchrony, which was why there was no preferential looking for this group. If long range connections between brain regions in ASD are not as temporally correlated as in typically developed individuals then this could explain some of these results especially when more complex stimuli such as speech were used. Overall, these findings suggest that there are multisensory and temporal deficits in autism spectrum disorders, which will likely have a major impact on communication and speech comprehension.

Communication in ASD

The integration of visual and auditory information makes speech naturally, multisensory. With the known impairments of communication in autism, speech and audiovisual integration have been greatly studied topics. It has been shown that individuals with ASD tend to perform worse on audiovisual tasks that use human faces and voices, but perform typically on tasks using non-human stimuli⁵⁶. One task that uses human speech stimuli to measure multisensory integration is the McGurk effect. In this illusion, participants see an individual's face and lips move to form the utterance of /ga/, while simultaneously hearing the utterance /ba/ which tends to cause participants to report the perception of hearing /da/⁵⁷. The McGurk effect can be thought of as a measure of multisensory integration because the McGurk percept of /da/ represents a fusion of both visual and auditory information. Similar to the sound induced flash illusion the McGurk effect is temporally restricted⁵⁸. Multiple studies have shown that children with ASD tend to perceive the McGurk illusion less often than typically developed individuals^{59, 60}. This would suggest a possible impairment in multisensory integration since the fused percept was reported less often. An interesting finding however was that adolescents and adults with ASD perceived this illusion as often as typically developed individuals, which suggests that these impairments may improve over time^{60, 61}.

In addition to a decreased McGurk percept, studies have found that children with ASD tend to perform worse on the visual only condition⁵⁹. This is demonstrated by an impaired ability to lip read, which would mean that individuals with ASD may not benefit from the added visual information especially in a noisy environment in order to understand speech appropriately^{62, 63}. Smith *et al* presented individuals with sentences within varying levels of noise

and participants were asked to identify specific words when the words were only heard or were seen and heard simultaneously⁶². This study demonstrated that individuals with ASD performed typically compared to controls on the auditory only condition, but were worse at identifying words when both audio and visual information were presented. Impaired multisensory and audiovisual integration in noisy environments, similar to those in the real world, could be one explanation for the communication impairments that are found in ASD. Recently, it has been shown that throughout typical development children can improve on speech in noise tasks⁶⁴. In addition to this finding it was demonstrated that after training on a lip reading paradigm, individuals with ASD reported the McGurk percept as often as controls⁵⁹. These findings provide evidence that this type of training may be another possible remediation tool for individuals with ASD, which may be the most beneficial if implemented earlier during development.

Overall, it has been shown that there are multiple levels of sensory dysfunction in autism spectrum disorders. Not only are there unisensory deficits, but multisensory impairments are also apparent. Atypical multisensory and temporal processing have been most evident in individuals with ASD especially as the stimuli become more complex and related to language. As studies have further characterized this sensory dysfunction, a few examples of possible remediation tools to improve multisensory integration, temporal processing, and speech comprehension have also been described. Multiple behavioral findings have described atypical unisensory and multisensory processing in ASD, which have been insightful in order to better understand this disorder.

Concluding Remarks

ASD is a heterogeneous neurodevelopmental disorder that impacts many aspects of an individual's life. Although multiple reports have described atypical structural and functional connections in ASD, there have not been many neurophysiological studies investigating deficits in multisensory integration and the brain regions that are potentially impacted. Russo *et al* published one of the first studies describing impairments in multisensory integration using EEG measures. This investigation demonstrated that multisensory integration within individuals with ASD was present specifically at later time intervals compared to typically developed individuals⁶⁵. This further suggests that individuals with ASD are capable of integrating stimuli from

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multiple modalities, yet temporal impairments are still evident. This study again supports the various findings of atypical multisensory and temporal processing. Based on functional imaging studies with typically developed individuals, one brain region that has been implicated in multisensory integration and temporal processing is the superior temporal sulcus⁶⁶⁻⁶⁹. This makes this region a potential area of interest in investigating multisensory impairments in ASD. Although there have been few neurophysiological studies describing multisensory and temporal processing in autism spectrum disorders, this allows for the opportunities of innovative research studies to be pursued.

The primary goal of this review was not only to give an overview of autism spectrum disorders and theories that have been used to explain these disorders, but also to specifically demonstrate how impairments in multisensory integration and temporal processing may relate to the common symptoms that characterize ASD. There have been numerous findings of atypical processing and sensitivity to unisensory stimuli spanning a variety of modalities. By studying how combinations of sensory stimuli are perceived, investigators have been able to further characterize the impairments in multisensory integration in individuals with ASD. Deficits in multisensory integration have led to findings of impaired temporal processing, which have generated an increased interest on how this could impact symptoms in ASD such as communication. This knowledge has generated behavioral studies focused on modulating audiovisual and speech stimuli. In addition to characterizing ASD behaviorally, there have been a variety of studies devoted to determining the neurobiology of ASD and how atypical development may impact connections and activity between brain regions in these disorders. A large number of neurophysiological studies have now characterized structural and functional impairments in individuals with ASD, but only a few have investigated multisensory processing. Additional studies that help to characterize atypical multisensory processing will not only improve our understanding of the impairments, but will also raise the possibility of developing better remediation tools for individuals with ASD in the future.

References

1. APA (2000). Diagnostic and Statistical Manual of Mental Disorders DSM-IV-TR. In: Washington, D.C: American Psychiatric Association.
2. CDC (2008). Prevalence of autism spectrum disorders – Autism and developmental disabilities monitoring network. In: Morbidity and Mortality Weekly Report Surveillance Summary, pp 1-19. United States: Centers for Disease Control and Prevention
3. DiCicco-Bloom E, Lord C, Zwaigenbaum L, Courchesne E, Dager SR, Schmitz C, Schultz RT, Crawley J and Young LJ (2006). The developmental neurobiology of autism spectrum disorder. *The Journal of Neurosci.* 26 (26): 6897-6906.
4. Casanova MF (2007). The neuropathology of autism. *Brain pathology.* 17 (4): 422-433.
5. White S, O'Reilly H and Frith U (2009). Big heads, small details and autism. *Neuropsychologia.* 47 (5): 1274-1281.
6. Courchesne E, Pierce K, Schumann CM, Redcay E, Buckwalter JA, Kennedy DP and Morgan J (2007). Mapping early brain development in autism. *Neuron.* 56 (2): 399-413.
7. Courchesne E (2004). Brain development in autism: early overgrowth followed by premature arrest of growth. *Ment Retard Dev Disabil Res Rev.* 10 (2): 106-111.
8. Casanova MF, Buxhoeveden DP, Switala AE and Roy E (2002). Minicolumnar pathology in autism. *Neurology.* 58 (3): 428-432.
9. Courchesne E, Redcay E, Morgan JT and Kennedy DP (2005). Autism at the beginning: Microstructural and growth abnormalities underlying the cognitive and behavioral phenotype of autism. *Development and Psychopathology.* 17 (3): 577-597.
10. Rubenstein JL and Merzenich MM (2003). Model of autism: increased ratio of excitation/inhibition in key neural systems. *Genes, brain, and behavior.* 2 (5): 255-267.
11. David O, Cosmelli D and Friston KJ (2004). Evaluation of different measures of functional connectivity using a neural mass model. *Neuroimage.* 21 (2): 659-673.
12. Wass S (2011). Distortions and disconnections: disrupted brain connectivity in autism. *Brain Cogn.* 75 (1): 18-28.
13. Schroeder JH, Desrocher M, Bebko JM and Cappadocia MC (2010). The neurobiology of autism: Theoretical applications. *Research in Autism Spectrum Disorders.* 4 (4): 555-564.
14. Minshew NJ and Keller TA (2010). The nature of brain dysfunction in autism: functional brain imaging studies. *Curr Opin Neurol.* 23 (2): 124-130.
15. Hughes JR (2007). Autism: the first firm finding = underconnectivity? *Epilepsy Behav.* 11 (1): 20-24.
16. Rippon G, Brock J, Brown C and Boucher J (2007). Disordered connectivity in the autistic brain: challenges for the "new psychophysiology". *International journal of psychophysiology : official journal of the International Organization of Psychophysiology.* 63 (2): 164-172.
17. Belmonte MK, Allen G, Beckel-Mitchener A, Boulanger LM, Carper RA and Webb SJ (2004). Autism and abnormal development of brain connectivity. *J Neurosci.* 24 (42): 9228-9231.
18. **Just MA, Cherkassky VL, Keller TA and Minshew NJ (2004). Cortical activation and synchronization during sentence**

comprehension in high-functioning autism: evidence of underconnectivity. Brain : a journal of neurology. 127 (Pt 8): 1811-1821.

This study proposes the theory of cortical underconnectivity in ASD, which discusses neurological impairments to describe atypical processing in ASD.

19. Levy F (2007). Theories of autism. The Australian and New Zealand journal of psychiatry. 41 (11): 859-868.
20. Happe F (1999). Autism: cognitive deficit or cognitive style? Trends Cogn Sci. 3 (6): 216-222.
21. Happe F and Frith U (2006). The weak coherence account: detail-focused cognitive style in autism spectrum disorders. J Autism Dev Disord. 36 (1): 5-25.
22. Brock J, Brown CC, Boucher J and Rippon G (2002). The temporal binding deficit hypothesis of autism. Development and psychopathology. 14 (2): 209-224.
23. Kana RK, Libero LE and Moore MS (2011). Disrupted cortical connectivity theory as an explanatory model for autism spectrum disorders. Phys Life Rev. 8 (4): 410-437.
24. Sgado P, Dunleavy M, Genovesi S, Provenzano G and Bozzi Y (2011). The role of GABAergic system in neurodevelopmental disorders: a focus on autism and epilepsy. Int J Physiol Pathophysiol Pharmacol. 3 (3): 223-235.
25. Di Cristo G (2007). Development of cortical GABAergic circuits and its implications for neurodevelopmental disorders. Clinical genetics. 72 (1): 1-8.
26. Iarocci G and McDonald J (2006). Sensory integration and the perceptual experience of persons with autism. Journal of Autism and Developmental Disorders. 36 (1): 77-90.
27. Kanner L (1943). Autistic disturbances of affective contact. Nervous Child. 2: 217-250.
28. Minshew NJ and Hobson JA (2008). Sensory sensitivities and performance on sensory perceptual tasks in high-functioning individuals with autism. Journal of Autism and Developmental Disorders. 38 (8): 1485-1498.
29. Schoen SA, Miller LJ, Brett-Green BA and Nielsen DM (2009). Physiological and behavioral differences in sensory processing: a comparison of children with autism spectrum disorder and sensory modulation disorder. Frontiers in integrative neuroscience. 3: 29.
30. Kern JK, Trivedi MH, Grannemann BD, Garver CR, Johnson DG, Andrews AA, Savla JS, Mehta JA and Schroeder JL (2007). Sensory correlations in autism. 11 (2): 123-134.
31. Hilton CL, Harper JD, Kueker RH, Lang AR, Abbacchi AM, Todorov A and LaVesser PD (2010). Sensory Responsiveness as a Predictor of Social Severity in Children with High Functioning Autism Spectrum Disorders. Journal of Autism and Developmental Disorders. 40 (8): 937-945.
32. Klintwall L, Holm A, Eriksson M, Carlsson LH, Olsson MB, Hedvall A, Gillberg C and Fernell E (2011). Sensory abnormalities in autism. A brief report. Res Dev Disabil. 32 (2): 795-800.
33. Rogers SJ and Ozonoff S (2005). Annotation: What do we know about sensory dysfunction in autism? A critical review of the empirical evidence. Journal of Child Psychology and Psychiatry. 46 (12): 1255-1268.
34. Dakin S and Frith U (2005). Vagaries of visual perception in autism. Neuron. 48 (3): 497-507.
35. Chen Y, Norton DJ, McBain R, Gold J, Frazier JA and Coyle JT (2012). Enhanced local processing of dynamic visual information in autism: evidence from speed discrimination. Neuropsychologia. 50 (5): 733-739.
36. Blake R, Turner LM, Smoski MJ, Pozdol SL and Stone WL (2003). Visual recognition of biological motion is impaired in children with autism. Psychological science. 14 (2): 151-157.
37. Spencer JV and O'Brien JM (2006). Visual form-processing deficits in autism. Perception. 35 (8): 1047-1055.
38. Tsermentseli S, O'Brien JM and Spencer JV (2008). Comparison of form and motion coherence processing in autistic spectrum disorders and dyslexia. Journal of Autism and Developmental Disorders. 38 (7): 1201-1210.
39. Marco EJ, Hinkley LB, Hill SS and Nagarajan SS (2011). Sensory processing in autism: a review of neurophysiologic findings. Pediatric research. 69 (5 Pt 2): 48R-54R.
40. O'Connor K (2012). Auditory processing in autism spectrum disorder: a review. Neurosci Biobehav Rev. 36 (2): 836-854.
41. O'Riordan M and Passetti F (2006). Discrimination in autism within different sensory modalities. Journal of Autism and Developmental Disorders. 36 (5): 665-675.
42. Meredith MA and Stein BE (1983). Interactions among Converging Sensory Inputs in the Superior Colliculus. Science. 221 (4608): 389-391.
43. Stein BE and Stanford TR (2008). Multisensory integration: current issues from the perspective of the single neuron. Nature reviews Neuroscience. 9 (4): 255-266.
44. Meredith MA and Stein BE (1986). Spatial Factors Determine the Activity of Multisensory Neurons in Cat Superior Colliculus. Brain Res. 365 (2): 350-354.
45. Meredith MA, Nemitz JW and Stein BE (1987). Determinants of Multisensory Integration in Superior Colliculus Neurons .1. Temporal Factors. Journal of Neuroscience. 7 (10): 3215-3229.
46. Shams L, Kamitani Y and Shimojo S (2000). Illusions. What you see is what you hear. Nature. 408 (6814): 788.
47. Shimojo S and Shams L (2001). Sensory modalities are not separate modalities: plasticity and interactions. Current opinion in neurobiology. 11 (4): 505-509.
48. Shams L, Kamitani Y and Shimojo S (2002). Visual illusion induced by sound. Brain Res Cogn Brain Res. 14 (1): 147-152.
49. van der Smagt MJ, van Engeland H and Kemner C (2007). Brief report: Can you see what is not there? Low-level auditory-visual integration in autism spectrum disorder. Journal of Autism and Developmental Disorders. 37 (10): 2014-2019.
50. Kwakye LD, Foss-Feig JH, Cascio CJ, Stone WL and Wallace MT (2011). Altered auditory and multisensory temporal processing in autism spectrum disorders. Frontiers in integrative neuroscience. 4: 129.
51. Foss-Feig JH, Kwakye LD, Cascio CJ, Burnette CP, Kadivar H,

CANDIDATE REVIEWS

- Stone WL and Wallace MT (2010). An extended multisensory temporal binding window in autism spectrum disorders. *Experimental Brain Research*. 203 (2): 381-389.
52. Hillock AR, Powers AR and Wallace MT (2011). Binding of sights and sounds: age-related changes in multisensory temporal processing. *Neuropsychologia*. 49 (3): 461-467.
53. Powers AR, Hillock AR and Wallace MT (2009). Perceptual Training Narrows the Temporal Window of Multisensory Binding. *Journal of Neuroscience*. 29 (39): 12265-12274.
54. Nakano T, Ota H, Kato N and Kitazawa S (2010). Deficit in visual temporal integration in autism spectrum disorders. *Proceedings Biological sciences / The Royal Society*. 277 (1684): 1027-1030.
55. Bebko JM, Weiss JA, Demark JL and Gomez P (2006). Discrimination of temporal synchrony in intermodal events by children with autism and children with developmental disabilities without autism. *Journal of child psychology and psychiatry, and allied disciplines*. 47 (1): 88-98.
56. Mongillo EA, Irwin JR, Whalen DH, Klaiman C, Carter AS and Schultz RT (2008). Audiovisual processing in children with and without autism spectrum disorders. *J Autism Dev Disord*. 38 (7): 1349-1358.
57. McGurk H and Macdonald J (1976). Hearing Lips and Seeing Voices. *Nature*. 264 (5588): 746-748.
58. van Wassenhove V, Grant KW and Poeppel D (2007). Temporal window of integration in auditory-visual speech perception. *Neuropsychologia*. 45 (3): 598-607.
59. Williams JH, Massaro DW, Peel NJ, Bosseler A and Suddendorf T (2004). Visual-auditory integration during speech imitation in autism. *Research in developmental disabilities*. 25 (6): 559-575.
60. **Taylor N, Isaac C and Milne E (2010). A comparison of the development of audiovisual integration in children with autism spectrum disorders and typically developing children. *Journal of Autism and Developmental Disorders*. 40 (11): 1403-1411**
- This study demonstrates that atypical multisensory integration may improve in individuals with ASD throughout development.*
61. Keane BP, Rosenthal O, Chun NH and Shams L (2010). Audiovisual integration in high functioning adults with autism. *Research in Autism Spectrum Disorders*. 4 (2): 276-289.
62. Smith EG and Bennetto L (2007). Audiovisual speech integration and lipreading in autism. *Journal of child psychology and psychiatry, and allied disciplines*. 48 (8): 813-821.
63. Iarocci G, Rombough A, Yager J, Weeks DJ and Chua R (2010). Visual influences on speech perception in children with autism. *Autism: the international journal of research and practice*. 14 (4): 305-320.
64. **Ross LA, Molholm S, Blanco D, Gomez-Ramirez M, Saint-Amour D and Foxe JJ (2011). The development of multisensory speech perception continues into the late childhood years. *Eur J Neurosci*. 33 (12): 2329-2337.**
- This study demonstrates that typically developed children improve throughout development on speech in noise tasks, which shows a potential for improvement in multisensory processing.*
65. **Russo N, Foxe JJ, Brandwein AB, Altschuler T, Gomes H and Molholm S (2010). Multisensory processing in children with autism: high-density electrical mapping of auditory-somatosensory integration. *Autism research : official journal of the International Society for Autism Research*. 3 (5): 253-267.**
- This is one of the first studies investigating multisensory integration in ASD using EEG methods to determine possible temporal processing impairments.*
66. Driver J and Noesselt T (2008). Multisensory interplay reveals crossmodal influences on 'sensory-specific' brain regions, neural responses, and judgments. *Neuron*. 57 (1): 11-23.
67. Nath AR, Fava EE and Beauchamp MS (2011). Neural correlates of interindividual differences in children's audiovisual speech perception. *J Neurosci*. 31 (39): 13963-13971.
68. Powers AR, 3rd, Hevey MA and Wallace MT (2012). Neural correlates of multisensory perceptual learning. *J Neurosci*. 32 (18): 6263-6274.
69. Stevenson RA, VanDerKlok RM, Pisoni DB and James TW (2011). Discrete neural substrates underlie complementary audiovisual speech integration processes. *Neuroimage*. 55 (3): 1339-1345.

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Tactile Motion on the Glabrous Hand of Human and Non-Human Primates

Jeremy Winberry

Tactile motion is a complex perceptual experience that requires a nervous system built from the ground up to allow rapid processing of spatial and temporal sensory information. Tactile motion relies primarily on Meissner corpuscles (RA-1), although Merkel cell-neurite complexes (SA-1) and Pacini corpuscles (RA-2) also play roles. Primary afferent projections from these receptors transmit sensory information to the brain in large fast fibers, and with high fidelity. The organization of primary somatosensory cortex is optimized for location and receptor type. Receptive fields of neurons in SI adapt with motion in order to increase feature selectivity. Neurons responding to motion, direction, and orientation can all be found in SI. Complex stimulus features, such as motion velocity, are resolved by population coding. Higher cortical areas for motion processing, such as the human motion complex (hMT+), are probably multisensory. Vision, in particular, seems to share motion processing architecture with the tactile modality. As in vision, tactile motion illusions may shed light on the cortical processing of motion, particularly when paired with functional imaging techniques. Apparent motion and the tactile motion aftereffect are two such illusions discussed in this review.

Keywords: *Somatosensation, motion perception, primates, humans*

In order to discuss tactile motion, it is first necessary to reveal the relevant biological pathways in cutaneous motion perception. The real action in tactile motion perception begins at the receptor surface of the somatosensory system, namely the skin. When the skin is deformed by physical stimulation, specialized mechanoreceptive neurons called primary afferent neurons are depolarized. This is possible because the terminal ends of these neurons contain mechanotransducer channels. These channels are normally closed but open when flexed. The exact mechanisms for this opening are varied¹⁻² and are not as well characterized as the mechanically gated channels of stereocilia in cochlear hair cells³. The induced currents from open channels can be recorded from the cell soma in the dorsal root ganglion⁴. Upon mechanical stimulation, these channels open, and cations, such as Na⁺ and Ca²⁺, rush into the terminal. If the inward rush of positive current is sufficient, then an action potential is produced. This basic transduction mechanism underlies the broad spectrum of mechanical somatosensation, including motion.

Mechanoreceptors of motion

The wide range of sensory percepts experienced is due to

different morphologies and anatomic locations of primary afferent terminals. In order to discuss motion on the glabrous skin of the hand, it is necessary to consider at least these afferent terminals: Meissner corpuscles, Merkel cell neurite complexes, and Pacinian corpuscles. These different mechanoreceptors are typically classified by their rate of adaptation to a stimulus. The rapidly adapting (RA) mechanoreceptors are Meissner corpuscles (RA-I) and Pacinian corpuscles (RA-II), whereas the slowly adapting (SA) mechanoreceptor is the Merkel cell neurite complex (SA-I). A second class of slowly adapting mechanoreceptors, Ruffini corpuscles (SA-II), are not present in the glabrous hand of the primate and exist only in tiny numbers in the glabrous hand of humans⁵. SA-I afferents terminate at Merkel cells at the base of the epidermis, the outermost layer of skin⁶. These afferents are densely populated in the glabrous skin of the hand, and provide precise localization and pressure information. RA-I afferents terminate in Meissner corpuscles just below the epidermis⁷, are layered with specialized Schwann cells, and respond well to slip, flutter and motion. RA-II afferents terminate at the base of the dermis in Pacinian corpuscles⁸ and respond preferentially to vibration. Each of these mechanoreceptor types contributes to motion in a unique way.

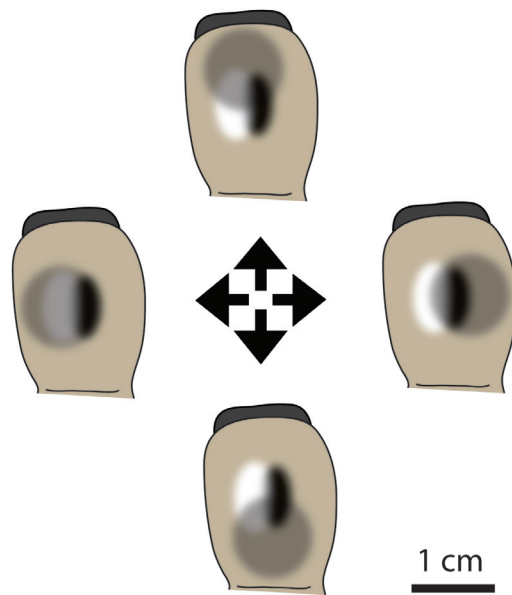


Figure 1: Receptive fields as seen in Area 3b neurons. On these four monkey fingertips, fixed excitatory (white), smaller fixed inhibitory (black) and larger lagged inhibitory (gray) receptive fields can be seen. The arrows in the center of the image signal the direction of a motion stimulus passing over the nearest fingerpad. The lagged inhibitory receptive field slides in the same direction as that of the motion stimulus, thereby modifying the overall receptive field. Note that these receptive fields are for illustration only and are not drawn to scale.

Dermatopic map:

A continuous representation of the somatic sensory surface, the skin. In this map, each fiber is beside the fiber that innervates the adjacent skin, and there are no breaks.

Somatotopic map:

A representation of body parts, some of which may be disconnected from the representation of adjacent skin areas. For example, the somatotopic map in primary somatosensory cortex has the thumb mapped next to the lower lip.

Modality:

A term that refers to the type of primary afferent, such as RA-1, SA-1, etc. Conservation of modality means that information from different primary afferent types stays segregated.

Mechanoreceptive afferent fibers can be individually recorded by microneurography⁹⁻¹². This has allowed characterization of the output of the different mechanoreceptor types. This has also allowed direct measurement of receptive fields. Receptive field sizes for RA-I and SA-I fibers are 6.2mm and 4.8mm, respectively¹³. These receptive fields are not homogeneous, as they contain hotspots where some terminal branches are more sensitive than others¹⁴. For example, SA-I and RA-I fibers increase their firing rates linearly with indentation¹⁵⁻¹⁶. It was also discovered that making a second indentation in the skin just outside the receptive field produces a suppressive effect by means of relieving skin pressure in the receptive field¹⁷.

Sensory transmission to the Central Nervous System

Mechanoreceptor afferent fibers are all large, myelinated A-beta fibers with conduction velocities between 36-73 m/s¹⁸. These properties are beneficial to rapid signal transmission to the central nervous system. Large-fibered mechanoreceptors project through the dorsal columns¹⁹ of the spinal cord to the dorsal column nuclei. Dermatopy is preserved in the dorsal column, where sacral fibers are

the most medial and cervical fibers are the most lateral²⁰⁻²¹. These fibers rearrange before synapsing in the dorsal column nuclei, shifting from a dermatopic map to a somatotopic map²². The primate cuneate nucleus contains a complete somatotopic representation of the sensory surfaces of the hand²³.

Medial lemniscal pathway. Under normal physiological conditions, dorsal column nuclei relay cells faithfully transmit the impulses of the primary afferent neurons²⁴⁻²⁷. These cells project through the medial lemniscus to type I relay cells in the lateral division of the ventral posterior nucleus (VPN) of thalamus (Vc in humans). The VPN is also somatotopically organized²⁸⁻²⁹. The region of the VPN containing cutaneous afferents from the lemniscal tract projects to layers 4 and 3 of Areas 3b and 1 of somatosensory cortex³⁰.

Thalamus and SI cortex. Place and modality information are conserved from thalamus to Area 3b and 1³¹, although signal transmission depends on vigilance. The amount of convergence from peripheral receptor to Area 3b is so restrained that Area 3b receptive field sizes are just 2-3 times the size of primary afferent receptive fields³². Dense microelectrode mapping demonstrates that there are somatotopic maps in areas 1 and 3b³³⁻³⁴, demonstrating the

conservation of place information. The fingers representations in Area 1 and 3b point away from each other³⁵. The hand representation can also be located histologically³⁶, or, in humans, anatomically³⁷⁻⁴¹ or by electrical cortical stimulation in waking humans⁴²⁻⁴⁴. These areas display a columnar structure⁴⁵, with different cortical columns representing different modalities, SA or RA⁴⁶.

All of the information so far illustrates that sensory information transduced by primary afferents in the skin is transmitted to cortex with high fidelity of place and modality information.

In some cases, primary afferent responses closely mimic psychophysical responses. For example, both primary afferent firing rates¹⁵⁻¹⁶ and psychophysically perceived pressure⁴⁷ increase linearly with skin indentation. Another example is that RA-1 tactile thresholds match psychophysical thresholds⁴⁸⁻⁴⁹. Furthermore, a single impulse in a single RA-1 fiber from the fingerpad produces a tactile percept¹². The cortex is essentially the bridge between stimulus transduction and tactile perception. Additional evidence linking cortical processing to this percept comes from finding BOLD response⁵⁰ and evoked potentials⁵¹ from the same stimulus.

The rapidity and fidelity of tactile information, as noted above, allow for motion processing. If a finger is held on a surface without moving, spatial features of that surface are only weakly observed. However, even surface features a few microns tall can be perceived on a moving surface⁵². RA-1 afferents are thought to be responsible for this gain of function with movement^{14,53}. This is especially true regarding slip of a smooth surface⁵⁴. Because RA-1 afferents are silent when held motionless on a surface, only the SA-1 afferents collect useful information. When the surface is moved, the RA-1 and SA-1 afferents can respond to raised dots as small as 2-4 μm and 8 μm in height, respectively⁵². The detection of edges, as opposed to dots, is possible at sub-micron heights⁵⁵. Interest-

ingly, sensitivity does not change over a wide range of velocities (10-40 mm/s)⁵². However, optimal velocity ranges do differ depending on skin type and, presumably, receptor density. Essick et al.⁵⁶ showed that optimal velocities for motion to be perceived on the fingertip of humans ranged from 15 to 94 mm/s whereas for the proximal forearm the optimal velocities were 115-312 mm/s. Strokes of movement had to be 5.9 times longer on the forearm than on the fingertip in order to obtain the same sensitivity⁵⁶. While increased force and velocity increase firing rate, the spatial pattern of firing does not change⁵⁷.

Cortical processing of motion

Primary somatosensory cortex processes basic sensory input to reveal complex features. One important cortical process is to use population coding. For example, localization discrimination thresholds on the fingertip can be as low as 0.38 mm for a 1.9 mm tactile probe⁵⁸. Although there is overlap in the area that the probe depresses in such intervals, the population responses are different enough to discriminate. Significantly, this discrimination threshold is smaller than the receptive fields of single primary afferents. Other complex percepts, such as curvature, orientation, movement, and direction, also require population coding.

Receptive fields of Area 3b neurons have been described in detail^{32,59-61}. 95% of these neurons have an excitatory field of about 24 mm² on the skin, with a range of 3-43 mm². However, about 5% of 3b cells in the same study had two or more excitatory receptive field regions. They also have an adjacent inhibitory field of about 18 mm², with a range of 1-47 mm². This configuration enhances feature contrast and preference. Remarkably, there is also a dynamic, delayed inhibitory field, whose position is not fixed, but rather biases in the direction of motion³². The 30 ms delay of this inhibitory field could serve to suppress minor features in the scanning direction on smooth surfaces and to emphasize novelty. At sufficient scanning speeds, it may even serve to confer directional

Orientation:

A cell is said to have orientation preference if its preferred stimulus passes through the receptive field at a specific angle, regardless of direction.

Motion:

A cell is said to have motion preference if its preferred stimulus moves through the receptive field at any angle and direction.

Direction:

A cell is said to have direction preference if its preferred stimulus moves through the receptive field in one direction, but not the opposite direction.

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Intrinsic optical imaging:

This functional imaging technique collects back-scattered light from the cortex and detects metabolic activity as oxygen-deficient hemoglobin absorbs more light than oxygen-rich hemoglobin.

preference⁶¹. Thus, Area 3b is the first stage of processing in the somatosensory system known to process motion.

Cortical neurons in Areas 3b, 1, and 2 are known to be sensitive to direction (60%), motion (37%), and orientation (3%)⁶². These cells are evenly split between RA type and SA type receptive fields⁶³. Motion cells are mostly located in Area 3b, whereas direction cells are mostly located in Areas 1 and 2. Most of this processing seems to occur in layer 3, as opposed to layer 4 where most thalamic projections terminate⁶⁴. Direction variant cells have also been found⁶⁵⁻⁶⁷. These cells respond to stimulus movement toward or away from a specific spot in the receptive field, typically located over a joint. Although primary somatosensory cortex (SI) has a broad variety of motion sensitivity, the higher-level characteristics of motion, such as velocity, appear to be processed in higher cortical areas.

Motion processing is thought to follow the dorsal pathway, which is used to guide movements. A number of imaging studies have implicated inferior parietal lobe and the human motion complex (hMT+)⁶⁸⁻⁷¹, although it has been proposed that the medial superior temporal area (MST) rather than the middle temporal area (MT) processes tactile motion⁷². Area hMT+ is best known for its role in visual motion processing. However, thanks to functional lesion studies with repetitive Transcranial Magnetic Stimulation (rTMS), it also appears to be necessary for tactile motion speed perception⁷³⁻⁷⁴. In order to deduce whether the area is truly multisensory or not, several studies have looked at combined visual and tactile motion paradigms. Results have included findings of facilitation between modalities⁷⁵ as well as interference between modalities⁷⁶⁻⁷⁷. These studies imply that there are shared resources for visual and tactile motion, lending further support to the idea that hMT+ is a multisensory motion processing area.

Motion Illusions

One of the major goals of cortical studies of motion is to uncover the neural correlates of perception. One way to study this is to use illusions in order to dissociate perception from reality. If the cortical area follows the real stimulus instead of the perceptual experience, then the area is not implicated in the illusory processing. Since many complex cortical processes use population coding, studying such processes at the single neuron level is unfeasible. Imaging methods are much better suited to study population coding, as they can look at processing within or between areas. Intrinsic optical imaging and functional Magnetic Resonance Imaging (fMRI) have been used to study the funneling illusion, finding that Area 3b activation reflects the perceived location rather than the somatotopic location⁷⁸⁻⁸⁰.

Several interesting tactile motion illusions exist and have been psychophysically characterized. One of these is apparent motion. Apparent motion is perceived motion created by sequential discrete tactile stimulations on spatially disparate skin locations. The spatial pattern of stimulation and type of movement (expanding, contracting, etc) have little effect on the saliency of the apparent motion, but the optimal range of inter-stimulus intervals varies with stimulus duration^{81,82}. The cortical mechanisms of apparent motion are not fully understood, but methods are being developed to probe this question with intrinsic optical imaging and fMRI⁸³⁻⁸⁴. These techniques offer the wide field of view necessary to study population coding. However, they sacrifice considerable spatial resolution in doing so. One promising technology that could be applied to tactile motion imaging is voltage sensitive dye imaging⁸⁵. This technique has already been applied to visual motion⁸⁶ and offers temporal resolution on the order of milliseconds. For a rapidly developing, complex stimulus such as motion, such a technique may be necessary to understand the mechanisms of cortical motion processing.

Another promising tactile motion illusion is the motion aftereffect (MAE). By generating a

continuous motion stimulus for a period of time (up to several minutes) and suddenly removing the stimulus, a tactile motion sensation in the other direction is produced⁸⁷⁻⁸⁹. This is very similar to the visual motion aftereffect. In fact, Konkle et al.⁹⁰ found that the aftereffect can transfer between tactile and visual modalities. This further suggests the existence of shared processing for motion in the two modalities. The tactile MAE is probably conferred through RA-1 afferents, as it is much more difficult to evoke in skin locations with a lower innervation density of these fibers⁹¹. There are now tactile MAE paradigms designed specifically to elicit the best responses from RA-1 afferents⁹². Interestingly, the tactile MAE is produced even with apparent motion across crossed digits⁹³. This means that the tactile MAE reflects environmental space as opposed to tactile space. The cortical correlates of this illusion are unknown, but a recent fMRI investigation shows that only SI remains active during the illusion⁹⁴, suggesting a functional role in the illusion.

The biological study of tactile motion seems to be entering a new era. Whereas microneurography and single-unit electrophysiology still dominated the field at the turn of the 21st century, the pendulum is swinging towards studies of population dynamics in relatively large cortical areas. Imaging methods, such as voltage sensitive dye imaging, offer a larger scope on cortical processing. The presence of two robust tactile illusions may prove to be critical for dissociating sensory and perceptual processing.

References

- Lumpkin EA and Caterina MJ (2007). Mechanisms of sensory transduction in the skin. *Nature*. 445: 858–865.
- Delmas P, Hao J and Rodat-Despoix L (2011). Molecular mechanisms of mechanotransduction in mammalian sensory neurons. *Nat Rev Neurosci*. 12: 139–153.
- Gillespie PG (1995). Molecular machinery of auditory and vestibular transduction. *Current Opinion in Neurobiology*. 5: 449–455.
- McCarter GC, Reichling DB and Levine JD (1999). Mechanical transduction by rat dorsal root ganglion neurons in vitro. *Neurosci Lett*. 273: 179–182.
- Paré M, Behets C and Cornu O (2003). Paucity of presumptive ruffini corpuscles in the index finger pad of humans. *J Comp Neurol*. 456: 260–266.
- Merkel F (1875). Tastzellen und Tastkörperchen bei den Hausthieren und beim Menschen. *Archiv f. mikrosk. Anat*. 11: 636–652.
- Meissner G (1853). *Beitrage zur Anatomie und Physiologie der Haut*, Leipzig:Leopold Voss.
- Pacini F (1840). *Nuovi organi scoperti nel corpo umano*, Pistoia:Tipografia Cino.
- Vallbo AB and Hagbarth KE (1967). Impulses recorded with micro-electrodes in human muscle nerves during stimulation of mechanoreceptors and voluntary contractions. *Electroencephalogr Clin Neurophysiol*. 23: 392 (1967).
- Vallbo AB, Hagbarth KE and Wallin BG (2004). Microneurography: how the technique developed and its role in the investigation of the sympathetic nervous system. *J Appl Physiol*. 96: 1262–1269.
- Vallbo AB and Johansson RS (1984). Properties of cutaneous mechanoreceptors in the human hand related to touch sensation. *Human neurobiology*. 3: 3–14.
- Torebjörk HE, Vallbo AB and Ochoa JL (1987). Intraneural microstimulation in man. Its relation to specificity of tactile sensations. *Brain*. 110 (6): 1509–1529.
- Phillips JR, Johansson RS and Johnson KO (1992). Responses of human mechanoreceptive afferents to embossed dot arrays scanned across fingerpad skin. *Journal of Neuroscience*. 12: 827–839.
- Gardner EP and Palmer CI (1989). Simulation of motion on the skin. II. Cutaneous mechanoreceptor coding of the width and texture of bar patterns displaced across the OPTACON. *J Neurophysiol*. 62: 1437–1460.
- Vega-Bermudez F and Johnson KO (1999). SA1 and RA receptive fields, response variability, and population responses mapped with a probe array. *J Neurophysiol*. 81: 2701–2710.
- Vega-Bermudez F and Johnson KO (1999). Surround suppression in the responses of primate SA1 and RA mechanoreceptive afferents mapped with a probe array. *J Neurophysiol*. 81: 2711–2719.
- Phillips JR and Johnson KO (1981). Tactile spatial resolution. III. A continuum mechanics model of skin predicting mechanoreceptor responses to bars, edges, and gratings. *J Neurophysiol*. 46: 1204–1225.
- Kakuda N (1992). Conduction velocity of low-threshold mechanoreceptive afferent fibers in the glabrous and hairy skin of human hands measured with microneurography and spike-triggered averaging. *Neurosci Res*. 15: 179–188.
- Head H and Holmes G (1911). Sensory disturbances from cerebral lesions. *Brain*. 34: 102–254.
- Walker AE and Weaver TA (1942). The topical organization and termination of the fibers of the posterior columns in *Macaca mulatta*. *J Comp Neurol*. 76: 145–158.
- Chang HT and Ruch TC (1947). Organization of the dorsal columns of the spinal cord and their nuclei in the spider monkey. *J Anat*. 81: 140–149.
- Whitsel BL, Petrucelli LM, Sapiro G and Ha H (1970). Fiber sorting in the fasciculus gracilis of squirrel monkeys. *Exp Neurol*. 29: 227–242.
- Florence SL, Wall JT and Kaas JH (1989). Somatotopic organization of inputs from the hand to the spinal gray and

CANDIDATE REVIEWS

- cuneate nucleus of monkeys with observations on the cuneate nucleus of humans. *J Comp Neuro.* 286: 48–70.
24. Ferrington DG, Rowe MJ and Tarvin RP (1987). Actions of single sensory fibres on cat dorsal column nuclei neurones: vibratory signalling in a one-to-one linkage. *The Journal of physiology.* 386: 293–309.
 25. Ferrington DG, Rowe MJ and Tarvin RP (1987). Integrative processing of vibratory information in cat dorsal column nuclei neurones driven by identified sensory fibres. *The Journal of physiology.* 386: 311–331.
 26. Greenstein J, Kavanagh P and Rowe MJ (1987). Phase coherence in vibration-induced responses of tactile fibres associated with Pacinian corpuscle receptors in the cat. *The Journal of physiology.* 386: 263–275.
 27. Douglas PR, Ferrington DG and Rowe M (1978). Coding of information about tactile stimuli by neurones of the cuneate nucleus. *The Journal of physiology.* 285: 493–513.
 28. Mountcastle VB and Henneman E (1952). The representation of tactile sensibility in the thalamus of the monkey. *J Comp Neurol.* 97: 409–439.
 29. Lenz FA, Dostrovsky JO, Tasker RR, Yamashiro K, Kwan HC and Murphy JT (1988). Single-unit analysis of the human ventral thalamic nuclear group: somatosensory responses. *J Neurophysiol.* 59: 299–316.
 30. Jones EG and Friedman DP (1982). Projection pattern of functional components of thalamic ventrobasal complex on monkey somatosensory cortex. *J Neurophysiol.* 48: 521–544.
 31. Jones EG, Friedman DP and Hendry SH (1982). Thalamic basis of place- and modality-specific columns in monkey somatosensory cortex: a correlative anatomical and physiological study. *J Neurophysiol.* 48: 545–568.
 32. **DiCarlo JJ and Johnson KO (2000). Spatial and temporal structure of receptive fields in primate somatosensory area 3b: effects of stimulus scanning direction and orientation. *J Neurosci.* 20: 495–510.**
This paper provides the best description of area 3b receptive fields. It is also the first to explain the lagged inhibitory field that biases in the direction of motion.
 33. Paul RL, Goodman H and Merzenich M (1972). Alterations in mechanoreceptor input to Brodmann's areas 1 and 3 of the postcentral hand area of *Macaca mulatta* after nerve section and regeneration. *Brain Res.* 39: 1–19.
 34. Paul RL, Merzenich M and Goodman H (1972). Representation of slowly and rapidly adapting cutaneous mechanoreceptors of the hand in Brodmann's areas 3 and 1 of *Macaca mulatta*. *Brain Res.* 36: 229–249.
 35. Kaas JH (1987). The Organization of Neocortex in Mammals: Implications for Theories of Brain Function. Annual review of psychology. 38: 129–151.
 36. Jain N, Catania KC and Kaas JH (1998). A histologically visible representation of the fingers and palm in primate area 3b and its immutability following long-term deafferentations. *Cereb Cortex.* 8: 227–236.
 37. Cunningham DJ (1982). Contribution to the surface anatomy of the cerebral hemispheres, Dublin: Academy House.
 38. White LE, Andrews TJ, Hulette C, Richards A, Groelle M, Paydarfar J and Purves D (1997). Structure of the human sensorimotor system. I: Morphology and cytoarchitecture of the central sulcus. *Cereb Cortex.* 7: 18–30.
 39. White LE, Andrews TJ, Hulette C, Richards A, Groelle M, Paydarfar J and Purves D (1997). Structure of the human sensorimotor system II: Lateral symmetry. *Cereb Cortex.* 7: 31–47.
 40. Sastre-Janer FA, Regis J, Belin P, Mangin JF, Dormont D, Masure MC, Remy P, Frouin V and Samson Y (1998). Three-dimensional reconstruction of the human central sulcus reveals a morphological correlate of the hand area. *Cereb Cortex.* 8: 641–647.
 41. Yousry TA, Schmid UD, Alkadhi H, Schmidt D, Peraud A, Buettner A and Winkler P (1997). Localization of the motor hand area to a knob on the precentral gyrus. A new landmark. *Brain.* 120 (Pt 1): 141–157.
 42. Cushing H (1909). A note upon the faradic stimulation of the postcentral gyrus in conscious patients. *Brain.* 32: 44–53.
 43. Foerster O (1936). The motor cortex in man in the light of Hughlings Jackson's doctrines. *Brain.* 59: 135–159.
 44. Penfield W and Jasper HH (1954). *Epilepsy and the functional anatomy of the human brain*, Boston: Little, Brown.
 45. Mountcastle VB (1997). The columnar organization of the neocortex. *Brain.* 120 (Pt 4): 701–722.
 46. Sur M, Wall JT and Kaas JH (1984). Modular distribution of neurons with slowly adapting and rapidly adapting responses in area 3b of somatosensory cortex in monkeys. *J Neurophysiol.* 51: 724–744.
 47. Stevens SS (1970). Neural Events and the Psychophysical Law: Power functions like those that govern subjective magnitude show themselves in neuropsychic effects. *Science.* 170: 1043–1050.
 48. Johansson RS and Vallbo AB (1979). Detection of tactile stimuli. Thresholds of afferent units related to psychophysical thresholds in the human hand. *The Journal of physiology.* 297: 405–422.
 49. Johansson RS and Vallbo AB (1979). Tactile sensibility in the human hand: relative and absolute densities of four types of mechanoreceptive units in glabrous skin. *The Journal of physiology.* 286: 283–300.
 50. Trullsson M, Francis ST, Kelly EF, Westling G, Bowtell R and McGlone F (2001). Cortical responses to single mechanoreceptive afferent microstimulation revealed with fMRI. *Neuroimage.* 13: 613–622.
 51. Kunesch E, Knecht S, Schnitzler A, Tyszer C, Schmitz F and Freund HJ (1995). Somatosensory evoked potentials elicited by intraneural microstimulation of afferent nerve fibers. *Journal of clinical neurophysiology.* 12: 476–487.
 52. LaMotte RH and Whitehouse J (1986). Tactile detection of a dot on a smooth surface: peripheral neural events. *J Neurophysiol.* 56: 1109–1128.
 53. Gardner EP and Palmer CI (1989). Simulation of motion on the skin. I. Receptive fields and temporal frequency coding by cutaneous mechanoreceptors of OPTACON pulses delivered to the hand. *J Neurophysiol.* 62: 1410–1436.

CANDIDATE REVIEWS

54. Srinivasan MA, Whitehouse JM and LaMotte RH (1990). Tactile detection of slip: surface microgeometry and peripheral neural codes. *J Neurophysiol.* 63: 1323–1332.
55. Johansson RS and LaMotte RH (1983). Tactile detection thresholds for a single asperity on an otherwise smooth surface. *Somatosens Res.* 1: 21–31.
56. Essick GK, Bredehoeft KR, McLaughlin DF and Szaniszlo JA (1991). Directional sensitivity along the upper limb in humans. *Somatosens Mot Res.* 8: 13–22.
57. Edin B, Essick G, Trulsson M and Olsson K (1995). Receptor encoding of moving tactile stimuli in humans. I. Temporal pattern of discharge of individual low-threshold mechanoreceptors. *J Neurosci.* 15: 830–847.
58. Wheat HE, Goodwin AW and Browning AS (1995). Tactile resolution: peripheral neural mechanisms underlying the human capacity to determine positions of objects contacting the fingerpad. *Journal of Neuroscience.* 15: 5582–5595.
59. DiCarlo JJ, Johnson KO and Hsiao SS (1998). Structure of receptive fields in area 3b of primary somatosensory cortex in the alert monkey. *Journal of Neuroscience.* 18: 2626–2645.
60. DiCarlo JJ and Johnson KO (1999). Velocity invariance of receptive field structure in somatosensory cortical area 3b of the alert monkey. *Journal of Neuroscience.* 19: 401–419.
61. DiCarlo JJ and Johnson KO (2002). Receptive field structure in cortical area 3b of the alert monkey. *Behav Brain Res.* 135: 167–178.
62. **Warren S, Hämäläinen HA and Gardner EP (1986). Objective classification of motion- and direction-sensitive neurons in primary somatosensory cortex of awake monkeys. *J Neurophysiol.* 56 (3): 598–622.**
- This paper was the first to describe orientation cells in SI. It also contains great characterizations of movement and direction cells.*
63. Ruiz S, Crespo P and Romo R (1995). Representation of moving tactile stimuli in the somatic sensory cortex of awake monkeys. *J Neurophysiol.* 73 (2): 525–537.
64. Whitsel BL, Roppolo JR and Werner G (1972). Cortical information processing of stimulus motion on primate skin. *J Neurophysiol.* 35: 691–717.
65. Essick GK and Whitsel BL (1985). Factors influencing cutaneous directional sensitivity: a correlative psychophysical and neurophysiological investigation. *Brain Res.* 357: 213–230.
66. Pei YC, Hsiao SS, Craig JC and Bensmaïa SJ (2010). Shape invariant coding of motion direction in somatosensory cortex. *PLoS Biol.* 8: e1000305.
67. Pei YC, Hsiao SS and Bensmaïa SJ (2008). The tactile integration of local motion cues is analogous to its visual counterpart. *Proceedings of the National Academy of Sciences.* 105: 8130–8135.
68. Francis S, Summers I, Clemence M, McGlone F, Chanter C and Bowtell R (2001). An fMRI study of brain activation due to tactile motion. *Neuroimage.* 13: S1166.
69. Hagen MC, Franzén O, McGlone F, Essick G, Dancer C and Pardo JV (2002). Tactile motion activates the human middle temporal/V5 (MT/V5) complex. *Eur J Neurosci.* 16: 957–964.
70. Summers IR, Francis ST, Bowtell RW, McGlone FP and Clemence MA (2009). Functional-magnetic-resonance-imaging investigation of cortical activation from moving vibrotactile stimuli on the fingertip. *J Acoust Soc Am.* 125: 1033–1039.
71. Wacker E, Spitzer B, Lützkendorf R, Bernarding J and Blankenburg F (2011). Tactile motion and pattern processing assessed with high-field fMRI. *PLoS ONE.* 6: e24860.
72. Beauchamp MS, Yasar NE, Kishan N and Ro T (2007). Human MST but not MT responds to tactile stimulation. *J Neurosci.* 27: 8261–8267.
73. Ricciardi E, Basso D, Sani L, Bonino D, Vecchi T, Pietrini P and Miniussi C (2011). Functional inhibition of the human middle temporal cortex affects non-visual motion perception: a repetitive transcranial magnetic stimulation study during tactile speed discrimination. *Exp. Biol. Med.* (Maywood). 236: 138–144.
74. Basso D, Pavan A, Ricciardi E, Fagioli S, Vecchi T, Miniussi C and Pietrini P (2012). Touching Motion: rTMS on the Human Middle Temporal Complex Interferes with Tactile Speed Perception. *Brain topography* (epub ahead of print).
75. Gori M, Mazzilli G, Sandini G and Burr D (2011) Cross-Sensory Facilitation Reveals Neural Interactions between Visual and Tactile Motion in Humans. *Front Psychol* 2: 55.
76. Craig JC (2006). Visual motion interferes with tactile motion perception. *Perception.* 35: 351–367.
77. Bensmaïa SJ, Killebrew JH and Craig JC (2006). Influence of visual motion on tactile motion perception. *J Neurophysiol.* 96: 1625–1637.
78. Chen LM, Friedman RM and Roe AW (2003). Optical imaging of a tactile illusion in area 3b of the primary somatosensory cortex. *Science.* 302: 881–885.
79. Chen LM, Turner GH, Friedman RM, Zhang N, Gore JC, Roe AW and Avison MJ (2007). High-resolution maps of real and illusory tactile activation in primary somatosensory cortex in individual monkeys with functional magnetic resonance imaging and optical imaging. *J Neurosci.* 27: 9181–9191.
80. Roe AW and Chen LM (2008). High-resolution fMRI maps of cortical activation in nonhuman primates: correlation with intrinsic signal optical images. *ILAR J.* 49: 116–123.
81. Kirman J (1974). Tactile apparent movement: The effects of interstimulus onset interval and stimulus duration. *Percept Psychophys.* 1–6.
82. Kirman JH (1983). Tactile apparent movement: the effects of shape and type of motion. *Percept Psychophys.* 34: 96–102.
83. Peelen MV, Rogers J, Wing AM, Downing PE and Bracewell RM (2010). Unitary haptic perception: integrating moving tactile inputs from anatomically adjacent and non-adjacent digits. *Experimental Brain Research.* 204: 457–464.
84. **Friedman RM, Dillenburger BC, Wang F, Avison MJ, Gore JC, Roe AW and Chen LM (2011). Methods for fine scale functional imaging of tactile motion in human and nonhuman primates. *Open Neuroimag J.* 5: 160–171.**
- This paper lays the groundwork for studying tactile motion perception with functional imaging methods, including fMRI and intrinsic optical imaging. These methods are promising because they can be used to study population responses in cortex.*

CANDIDATE REVIEWS

85. Grinvald A and Hildesheim R (2004). VSDI: a new era in functional imaging of cortical dynamics. *Nat Rev Neurosci.* 5: 874–885.
86. Onat S, Nortmann N, Rekauzke S, Konig P and Jancke D (2011). Independent encoding of grating motion across stationary feature maps in primary visual cortex visualized with voltage-sensitive dye imaging. *Neuroimage* 55: 1763–1770.
87. **Hollins M and Favorov O (1994). The Tactile Movement Aftereffect. *Somatosens Mot Res.* 11: 153–162.**
This paper describes the first successful method for presenting a tactile motion aftereffect. This aftereffect may prove useful in studying perceptual experience in primate cortex.
88. Lerner EA and Craig JC (2002). The prevalence of tactile motion aftereffects. *Somatosens Mot Res.* 19: 24–29.
89. Planetta PJ and Servos P (2008). The tactile motion aftereffect revisited. *Somatosens Mot Res.* 25: 93–99.
90. Konkle T, Wang Q, Hayward V and Moore CI (2009). Motion aftereffects transfer between touch and vision. *Curr Biol.* 19: 745–750.
91. Planetta PJ and Servos P (2010). Site of stimulation effects on the prevalence of the tactile motion aftereffect. *Experimental Brain Research.* 202: 377–383.
92. Watanabe J, Hayashi S, Kajimoto H, Tachi S and Nishida S (2007). Tactile motion aftereffects produced by appropriate presentation for mechanoreceptors. *Experimental Brain Research.* 180: 577–582.
93. Kuroki S, Watanabe J, Mabuchi K, Tachi S and Nishida S (2011). Directional remapping in tactile inter-finger apparent motion: a motion aftereffect study. *Experimental Brain Research.* 216: 311–320.
94. Planetta PJ and Servos P (2011). The postcentral gyrus shows sustained fMRI activation during the tactile motion aftereffect. *Experimental Brain Research.* 216: 535–544.

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