

Laser Speckle Contrast System for Determination of Parathyroid Perfusion

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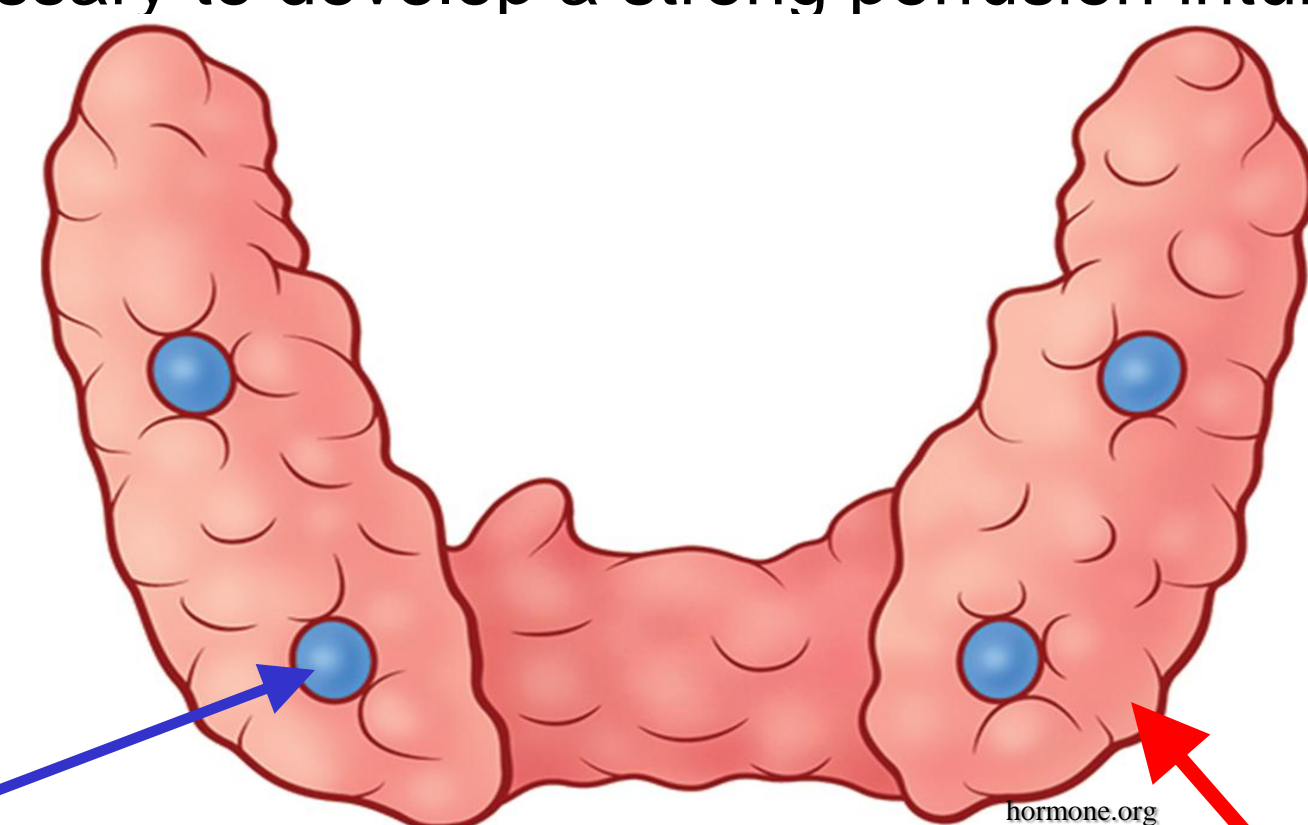
PROBLEM STATEMENT

Surgical operations to remove the thyroid gland may inadvertently result in a loss of perfusion (blood flow) to the surrounding parathyroid glands. Other than visual observation, endocrine surgeons have no way of knowing the perfusion state of the parathyroid glands.

BACKGROUND

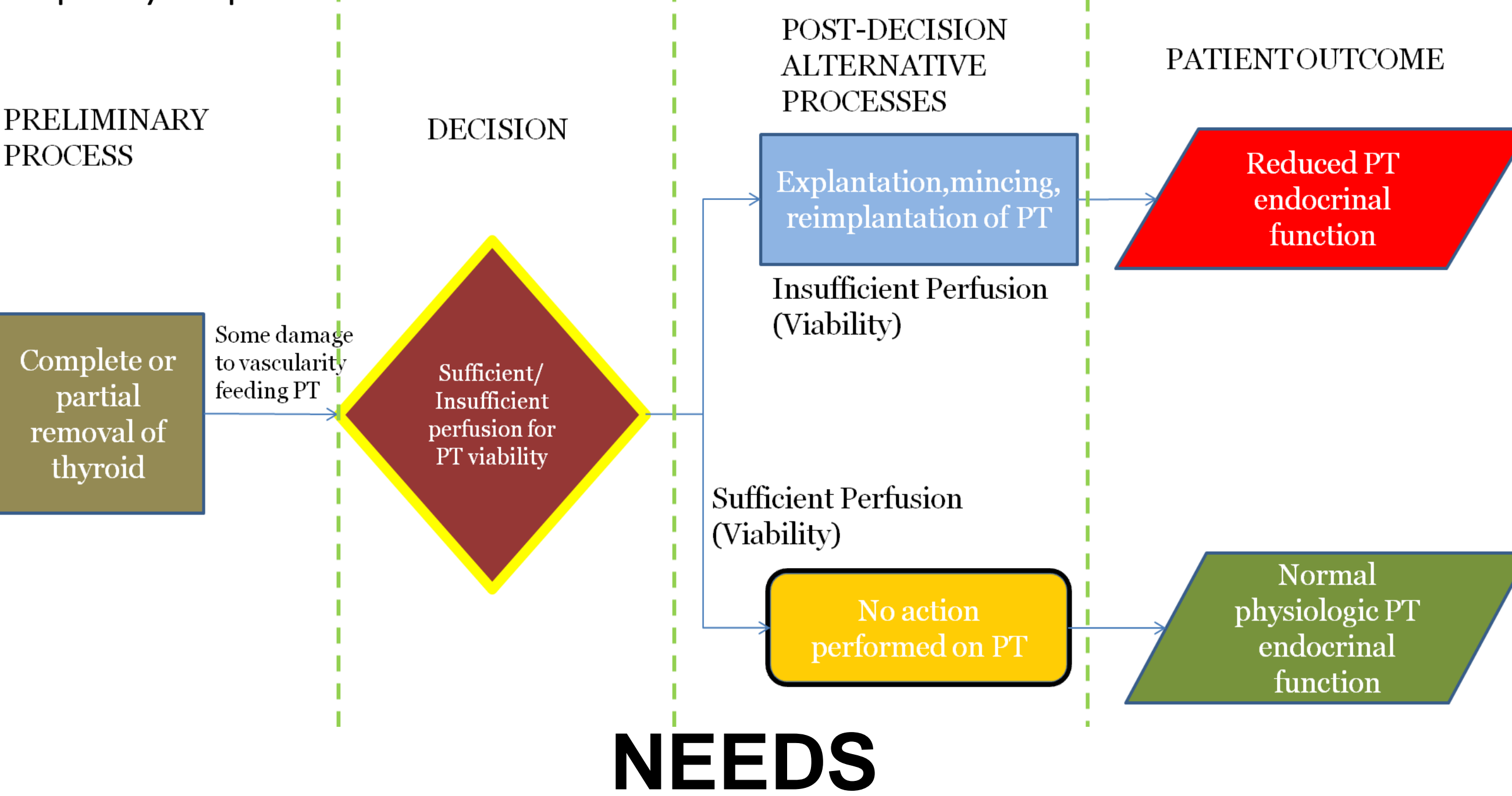
The thyroid gland, an important part of the endocrine system, sometimes displays hypertrophy, becomes cancerous, or experiences other pathophysiological conditions. The parathyroid glands, four of which are situated posterior to the thyroid, are critical to endocrine system regulation of calcium homeostasis.

In an operation to remove the thyroid, surgeons may inadvertently lacerate or otherwise damage vascular beds feeding the parathyroid; consequently parathyroid glands lose perfusion and will eventually necrose. Surgeons currently inspect the parathyroid glands visually to best determine the perfusion state; however, many surgeons do not have the high patient volumes necessary to develop a strong perfusion intuition.



In blue are the parathyroid glands nestled among the red thyroid.

If the parathyroid is adequately perfused, no further action is needed to ensure optimal parathyroid gland function; however, in a situation of deperfusion, the parathyroid is explanted, minced, and reimplanted into the sternocleidomastoid muscle to maintain optimal parathyroid output by simple osmotic diffusion of hormones.



NEEDS

Device must determine perfusion state.

Must have clinical utility.

Device should use maximally low cost components.



DESIGN THEORY/COMPONENTS

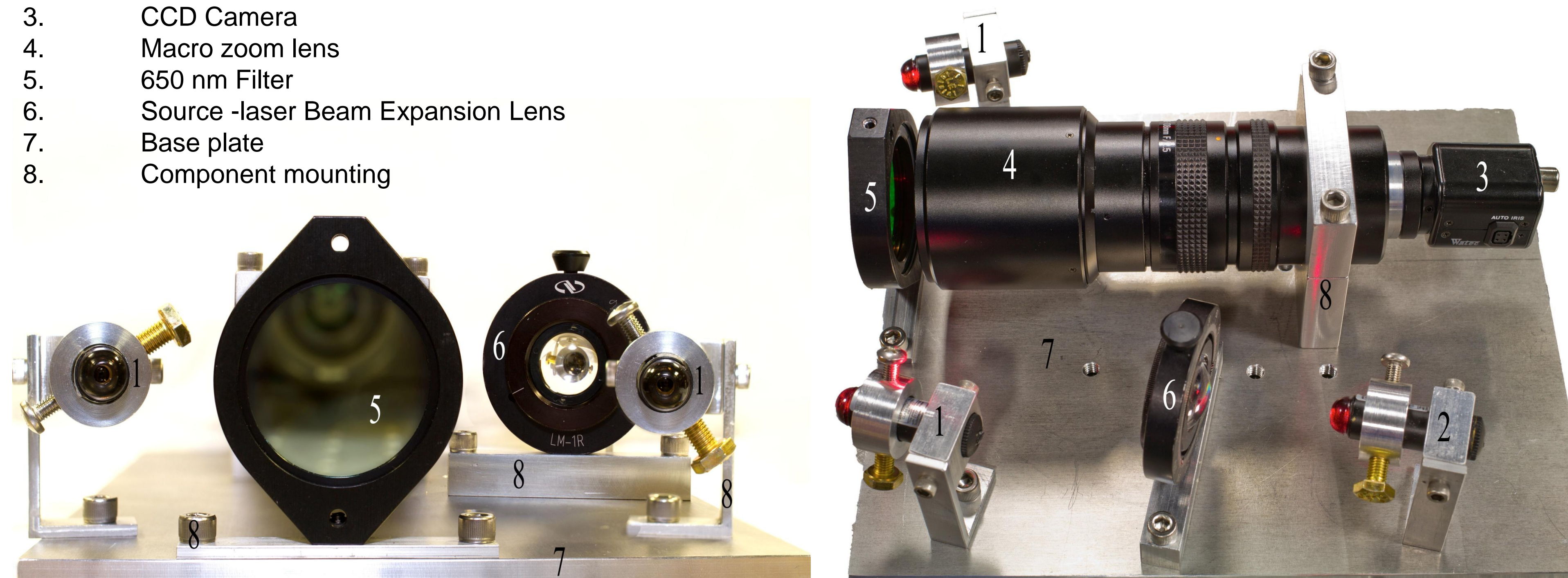
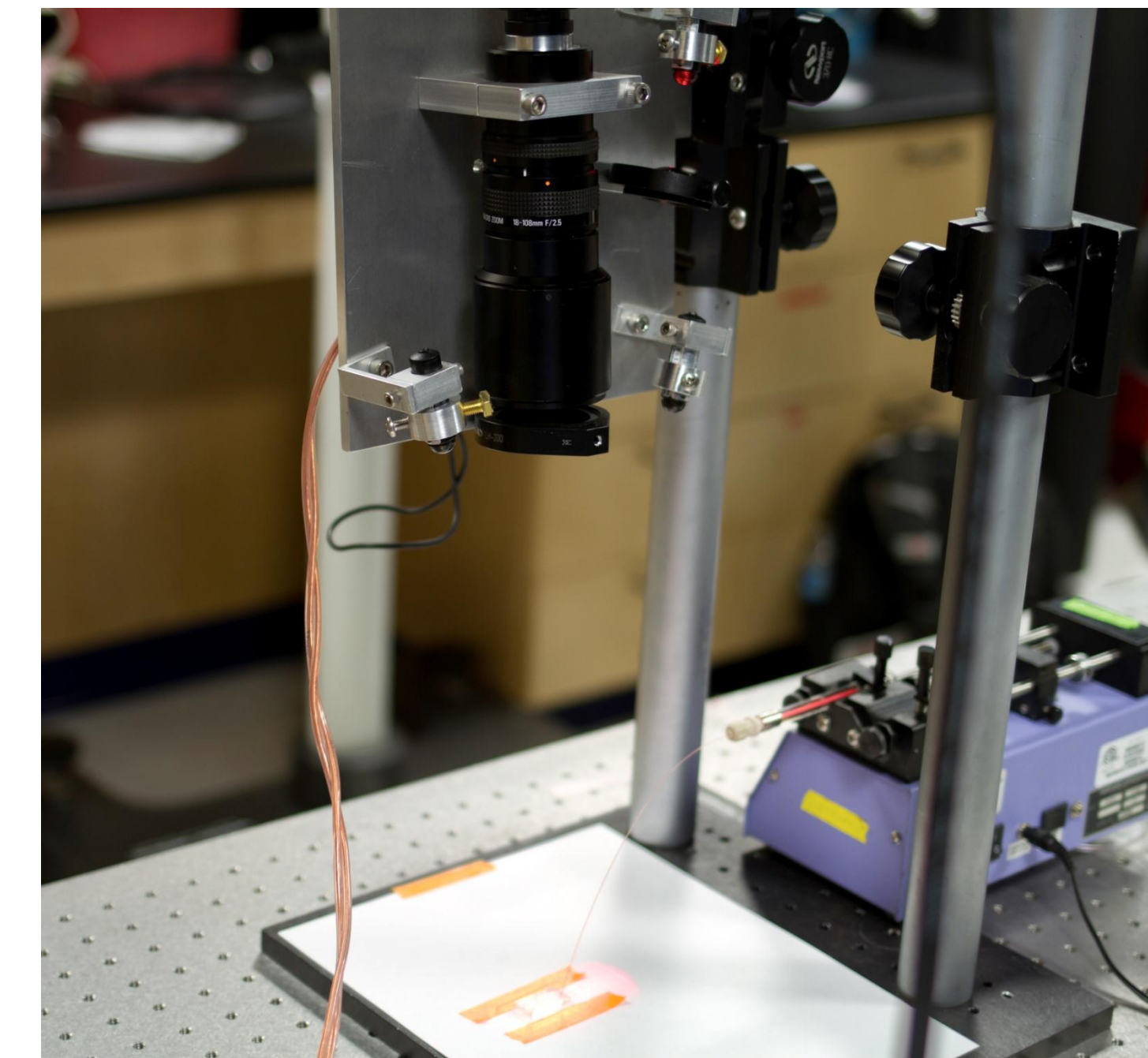
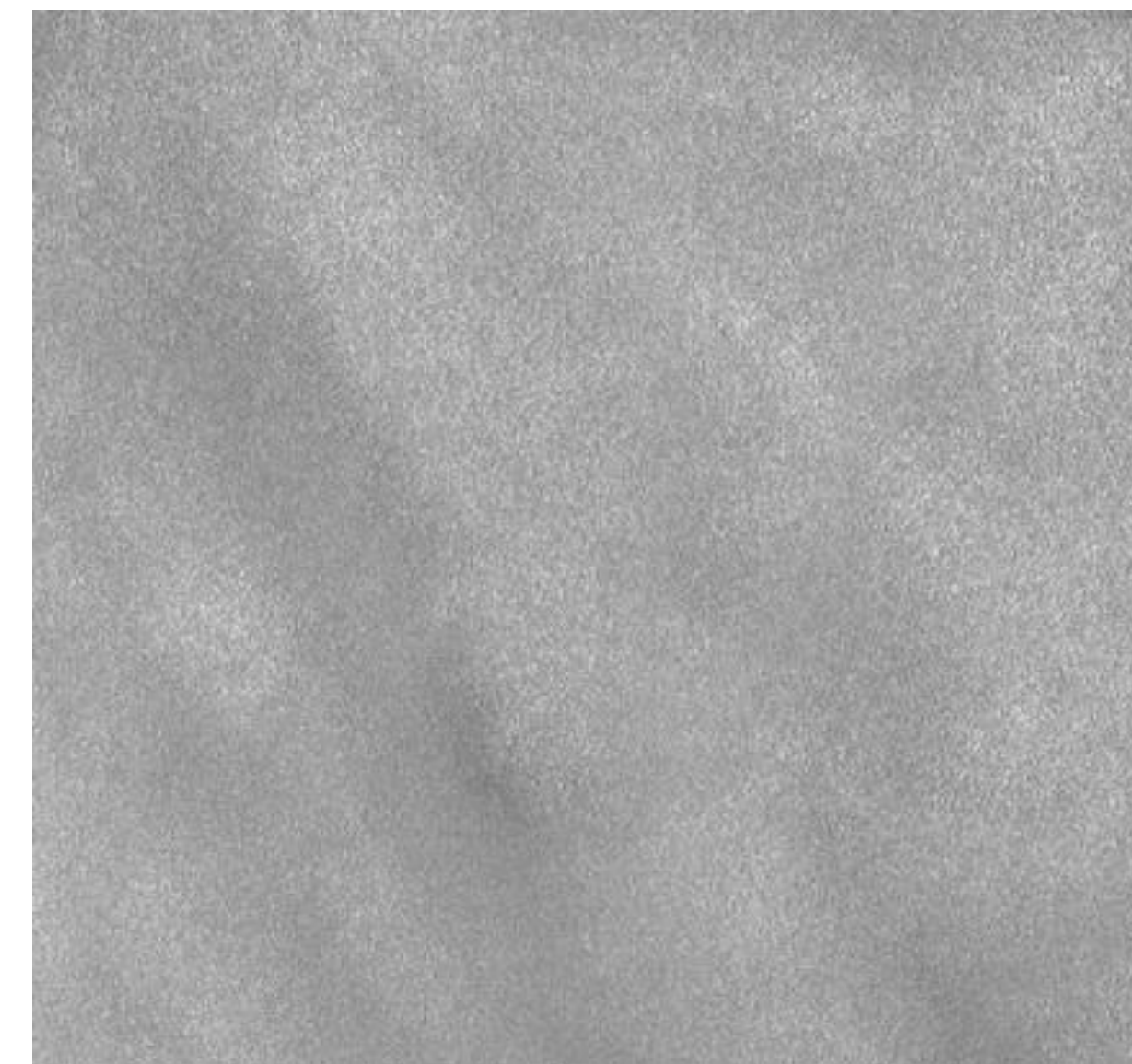
Laser Speckle Theory

- Background
1. Diffuse Reflectance
 2. Speckle Pattern
 3. Contrast Equation

$$K_s = \frac{\sigma_s}{\langle I \rangle}$$

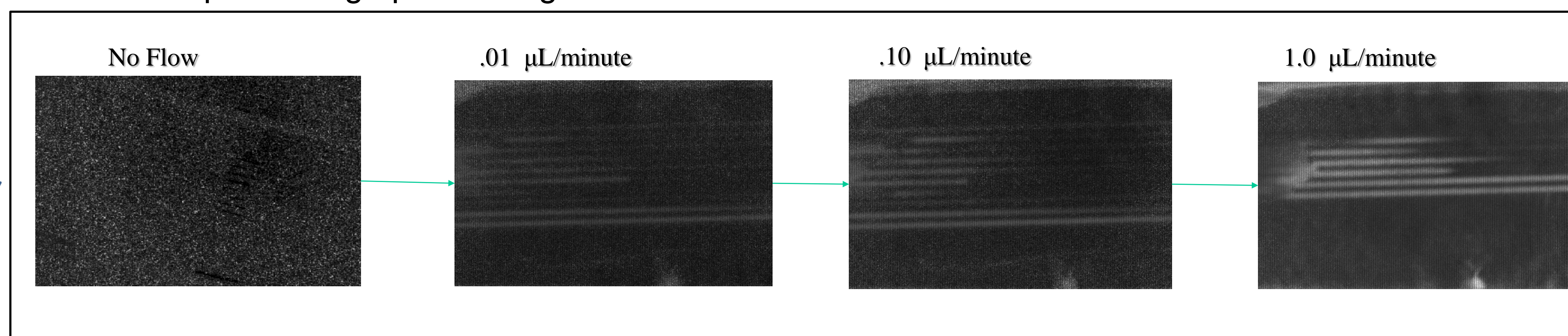
Design Components

1. 2 Guide Laser Pointers
2. 1 Source Laser Pointer
3. CCD Camera
4. Macro zoom lens
5. 650 nm Filter
6. Source -laser Beam Expansion Lens
7. Base plate
8. Component mounting



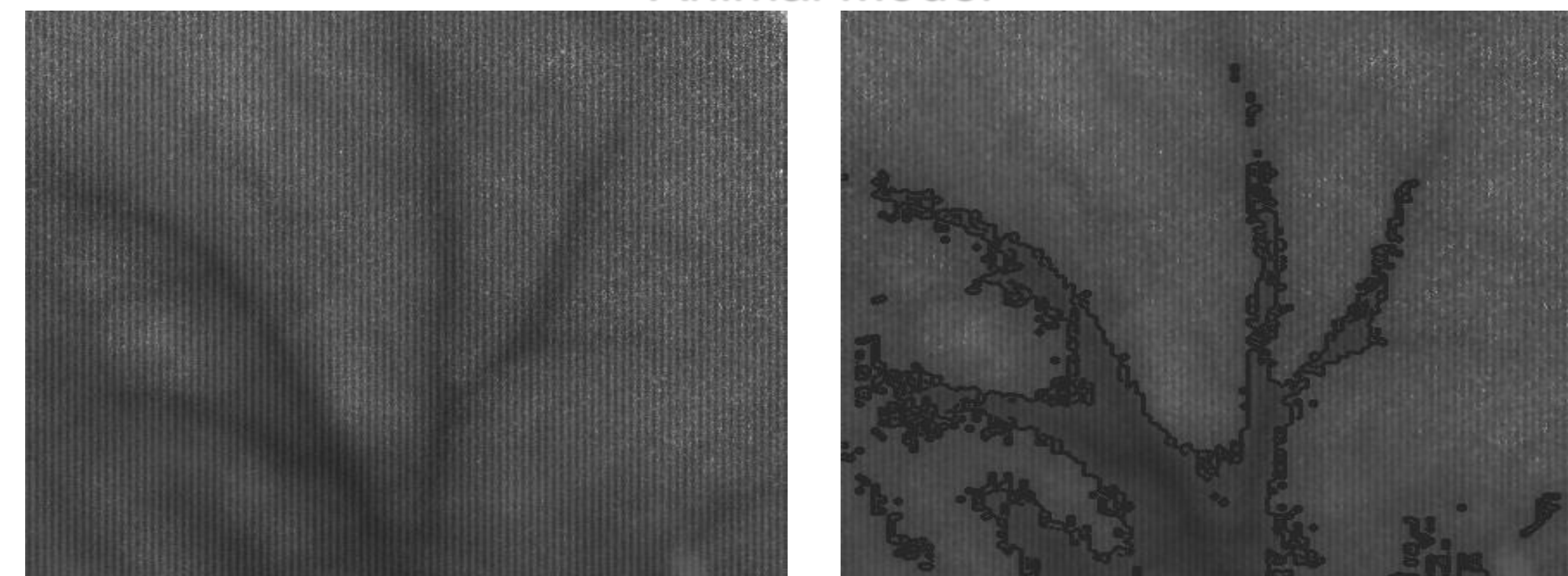
VALIDATION

- Using a PDMS microfluidics phantom pumped with a solution of polystyrene beads, we were able to validate that our device is able to detect the presence of flow.
- Capture 150 images through LabView at 29.7 fps
- Use temporal image processing in MatLab



- Speckle Ratio: a means to give a quantitative value to our images
- Speckle ratio = mean intensity vessel / mean intensity background
- Early tests indicate a positive correlation, although more tests are needed

Animal Model



	Test 5	Test 6	Test 7	Test 8	Test 9
Ratio	0.7205	0.7164	0.7132	0.7277	0.7161

FUTURE CONSIDERATIONS

DEVICE IMPROVEMENTS

Reduce size/ weight of overall device

- Cut out parts of base not supporting the components
- Use a smaller and more compact macro lens
 - The current macro lens is the largest and the most expensive part of our design

Quantifying flow relationship

Current progress has shown that in no flow conditions of the phantom, device does not register perfusion. The processed image shows a stronger contrast between areas of flow and no flow. We are currently investigating this relationship to set a standard for flow detection. Some additional investigation needs to be conducted regarding the device's variability, detection range and sensitivity.

Preparations for clinical testing

In order to improve the ease of our device, several developments could be made:

- Improve compatibility with current parathyroid detection device
- Better integrate device with surgical equipment: surgical arm or IV pole
- Develop components that allows for easier sterilization
- Streamline data acquisition process

CONCLUSION

We have:

- Identified Laser Speckle as an imaging technique that will detect perfusion,
- Designed and created a device that meets the needs
- Validated our device through both animal and phantom testing. Future work needs to be done to improve the device for a clinical setting.

ACKNOWLEDGEMENTS

We would like to thank our advisors **Dr. Anita Mahadevan-Jansen** and **Dr. Matthew Walker III** for their insight and guidance throughout this process, **Dr. James Broome** for his expertise on the clinical considerations, **Melanie McWade** for her weekly help and meetings, **John Fellenstein** for his tireless and high quality machining, **Kristin Poole** for her help with animal testing, and **Isaac Pence** and the other graduate students at the BME Biophotonics Lab for their help.

REFERENCES

1. Briers, David J.. "Laser speckle contrast imaging for measuring blood flow ." *Optica Applicata* 37 (2007): 139-152. Print.
2. Richards, Lisa M., S. M. Shams Kazmi, Janel L. Davis, Katherine E. Olin, and Andrew K. Dunn. "Low-cost laser speckle contrast imaging of blood flow using a webcam." *Biomedical Optics Express* 4.10 (2013): 2269. Print.
3. Tom, W.j., A. Ponticorvo, and A.k. Dunn. "Efficient Processing of Laser Speckle Contrast Images." *IEEE Transactions on Medical Imaging* 27.12 (2008): 1728-1738. Print.