

DynaStar NanoPro DLS User Guide

The DynaPro NanoStar is a dynamic light scattering (DLS) instrument that is used for the analysis of protein solutions, promiscuous inhibitors, micelles, buffers or other products in solution.

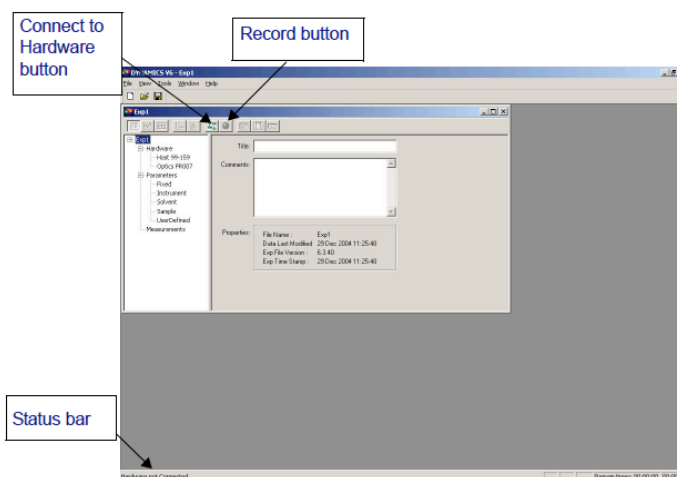
Sample Preparation:

1. Concentration of 1 mg/mL is an ideal starting point
 - a. Bring extra buffer to dilute
 - b. A lower concentration may work, but it would just have to be tested
2. Samples can be run as-is, centrifuged, or filtered.
 - a. Unspun – 5uL
 - b. Centrifuged – 10uL
 - c. Filtered – 100uL (accounts for dead volume)
3. Run a blank if using surfactant or glycerol
4. Cuvette types
 - a. Disposable:
 - Minimum volume =5uL
 - For dynamic light scattering only
 - Tolerates temperatures up to 80°C
 - b. Quartz:
 - Minimum volume =1.25uL
 - For static light scattering or dynamic light scattering
 - Used when you have precious protein and/or organic solvents
 - Tolerates temperatures >80°C
 - Replacement cost is \$2,500
5. Temperature control
 - a. System can be heated to 150°C. If heating a sample add ~100uL of paraffin or silicon oil around the chimney to prevent evaporation
 - b. Our system is not currently set up for runs below 20°C. We would need a dry gas source (i.e. nitrogen)

Operation:

1. Power on instrument, laser, and computer
 - Username: wyattlab
 - Password: dynapro
 - Wait 30 minutes before running any experiments
2. Launch “Dynamics” software
 - a. File -> open -> preset (configuration/method file)
 - b. File -> open -> previous experiment (for comparative reasons i.e. buffer conditions)
 - c. File -> open -> new (NOT recommended per technician, saves settings from previous user)

3. Using Preset:
 - Auto attenuation— system focusing is like a pupil and will adjust based on light
 - Automatic 5 minute delay for temperature stability
 - Standard data acquisition is 3 measurements, but can take 5+ if better statistics are needed.
4. Click “connect” icon in toolbar
 - Record button will activate (green circle)
 - Trouble? – Disconnect/connect Ethernet cable then retry or contact Heather Darling (h.darling@vanderbilt.edu room 5128C)

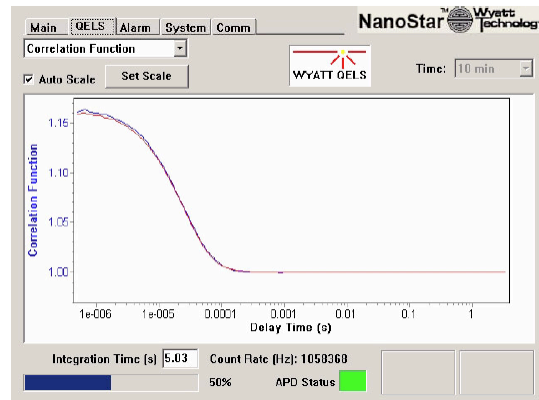


5. Double click on event schedule
 - Under commands and values -> label sample -> enter
 - Save data as -> click edit to open directory
 - Verifies all data is going to this file
6. Load sample in cuvette
 - Insert pipette tip all the way to the bottom of cuvette then slowly pull up while volume is dispensed. This helps prevent air bubble formation
 - For disposable cuvette the sample goes in the center “chimney” indicated below by the dark purple color



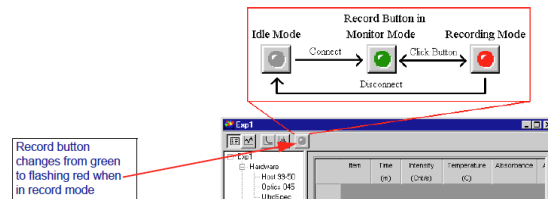
7. Place cuvette in cell chamber and close lid
 - Disposable -> make sure “front” faces forward
 - Quartz -> flattened corner positioned on the left front side
8. On the Dynapro instrument highlight the “QUELS” tab
 - Quels = quasi electric light scattering
 - No correlation/flat line indicates no protein in sample (i.e. buffer), air bubble, or cuvette is not pushed all the way in

- Correlation (like below) indicates protein



9. Acquire data -> push green icon on computer

- Arrow in left column will point to which step the system is on

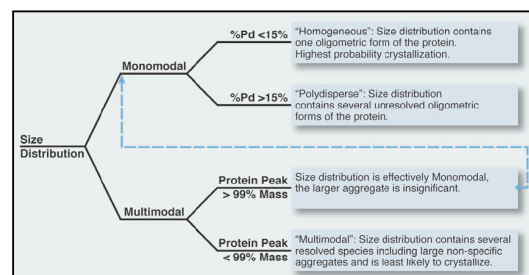


10. Analysis can begin once the green icon comes back -> see next section

Analysis:

Dynamics software can be installed in any computer on campus. Return the disc and license key once this is complete!

- Tools -> calculations -> optimization
 - Insert molar mass
 - Helps understand expected size
 - Can be used to figure out minimum concentration needed
- Fit models/data correlation
 - Postulates size distribution as Gaussian (radius, polydispersity)
 - Multixponential analysis (regularization)
- If all red data (thrown out data):
 - Right click -> select data filter
 - Lower SOS = better Gaussian -> this can be disabled
 - Tweak baseline to 0.05



Finishing and clean up:

1. Remove cuvette
 - Disposable – discard
 - Quartz – clean using cuvette washer in Megalab
2. Turn off the laser and Dynapro instrument
3. Export data via internet and/or flash drive
4. Log off computer
5. Have a nice day ☺