

**Amanda Linkous, Ph.D.**  
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**Current Position:**           **Research Associate Professor, Scientific Center Manager**  
NCI Center for Cancer Systems Biology of Small Cell Lung Cancer  
Department of Pharmacology, Vanderbilt University  
Nashville, TN 37232

**Educational Background:** **Doctorate of Philosophy, Cancer Biology**, December 2009  
Vanderbilt University, Nashville, TN

**Bachelor of Science, Biology**, May 2005  
Minor: Microbiology  
East Tennessee State University, Johnson City, TN  
Honors: Summa Cum Laude

**Technical Skills and Managerial Responsibilities:**

- Over 18 years of expertise in cancer biology including 3D culture models, metastasis, and cancer stem cells
- 10+ years of management experience in academic research institutions
- Manages multi-PI Center (>8 laboratories) and activities including scientific research projects, outreach, NCI and external advisory board site visits and Center workshops
- Responsible for creating and submitting Research Performance Progress Reports (RPPRs) for all NIH-related Center funding mechanisms
- Routinely interacts and initiates collaborations with patient foundations and industry partners
- Single-handedly coordinated three new laboratory site transitions from NIH/NCI, NYU, and Weill Cornell Medicine
- Personally trained and mentored 20+ scientists/trainees
- Established novel, *ex vivo* cerebral organoid and lung organoid model systems for the laboratory
- Implemented organoid/drug screening platforms for CLIA-certified core facility
- Routinely derived glioma stem cells from patient brain tumor specimens
- Generated pediatric tumor stem cells and subsequent tumor organoids from wide variety of pediatric brain tumor specimens; key collaborator for Institute of Precision Medicine
- Reprogrammed patient blood into induced pluripotent stem cells for cerebral organoid generation
- Extensively educated and trained new scientists to culture patient-derived glioma stem cells
- Derived unique neural stem cell line from human embryonic stem cells
- Served as Co-Investigator on IRB protocol for Weill Cornell/New York-Presbyterian
- Responsible for the acquisition and cryopreservation of patient-derived tissue and cell lines for multiple investigators
- Routinely interacts with biotech industry leaders and senior management from pharmaceutical companies to facilitate collaborations; serves as consultant to life sciences corporations

- Managed financial accounts in excess of \$18M
- Authored and received IACUC/RARC approval for animal research protocols
- Negotiates with vendors and reduced operating costs by more than \$100K per year
- Prioritizes and delegates tasks to ensure that all time-sensitive projects are completed
- Served as primary author of document for City, State, and Federal Regulatory Compliance
- Responsible for all staff evaluation, training, credentials, and licensure
- Developed and implemented protocols to meet EPA, DEP, FDNY, and EH&S compliance programs
- Responsible for all inspections from New York City, New York State, and Federal Agencies
- Maintains advanced proficiency in REDCap database management, PeopleSoft & SRM Applications

**Professional Activities, Awards, and Service:**

**Awardee**, 2023 Celselect Research Grant Recipient

**Invited Speaker and Panelist**, Corning Life Sciences 3D Cell Culture Summit, New York, NY, September 21-22, 2022

**Invited Session Chair**, Systems Biology of Human Disease Conference, Vanderbilt University, June 21, 2022

**Featured Speaker**, Corning Life Sciences, AACR, New Orleans, LA, April 11, 2022

**Initiative Review Committee Member**, Accelerating Radiotherapeutics through Molecular Constructs (ARM) Initiative, Oak Ridge National Laboratory, 2021-present

**Translational Research Fellowship Mentor for UAMS**, 2021-present

**CCSB of SCLC, Outreach Core Leader (U54)**, 2021-2023

**Grant Referee**, Neurological Foundation, 2021-present

**Invited Speaker and Panelist**, ARIA Workshop, Oak Ridge National Laboratory, September 2020

**Review Editor**, *Nature Protocols*

**Review Editor**, *Frontiers in Neuro-Oncology & Neurosurgical Oncology*

**Review Editor**, *Cell Press: STAR Protocols*

**Review Editor**, *Journal of Cancer Metastasis and Treatment*

**Invited Lecturer**, Quantitative & Chemical Biology Program, Vanderbilt University 2019-Present

**Invited Session Chair**, Single Cell Biology Symposium, Vanderbilt University, February 12, 2019

**Invited Panelist**, Corning Life Sciences 3D Cell Culture Summit, Boston, MA, October 16-17, 2018

**Consultant**, Corning Life Sciences

**Consultant**, University of Arkansas for Medical Sciences

**Research Experience:**

**Research Associate Professor, Scientific Center Manager (January 2019-Present)**

NCI Center for Systems Biology of Small Cell Lung Cancer, Vanderbilt University  
Laboratory of Vito Quaranta, M.D.

**Patient Advocacy Lead (November 2019-September 2020)**

Parthenon Therapeutics, LLC

**Director, Starr Foundation Cerebral Organoid Translational Core (February 2017-December 2018)**

Weill Cornell Medicine, NewYork-Presbyterian Hospital

**Senior Research Associate and Laboratory Manager (February 2015-February 2017)**

Sandra and Edward Meyer Cancer Center, Weill Cornell Medicine  
Laboratory of Howard A. Fine, M.D.

**Associate Research Scientist and Laboratory Manager (March 2013-January 2015)**

Laura & Isaac Perlmutter Cancer Center, New York University Langone Medical Center  
Laboratory of Howard A. Fine, M.D.

**Postdoctoral Training (February 2010-February 2013)**

Neuro-Oncology Branch, National Cancer Institute, National Institutes of Health  
Advisor: Howard A. Fine, M.D.

***Project: Tumor metabolism in glioma stem cells.***

**Predoctoral Training (April 2006-December 2009)**

Department of Cancer Biology, Vanderbilt University School of Medicine.  
Advisor: Dennis E. Hallahan, M.D.

***Dissertation: The role of lipid second messengers in angiogenesis and the vascular endothelium response to radiation.***

**Undergraduate Training (May 1, 2004-May 31, 2005)**

Department of Internal Medicine, James H. Quillen College of Medicine  
East Tennessee State University  
Advisor: David S. Chi, Ph.D.

***Project(s): Effects of Catecholamines on Pro-atherogenic Chemokine Production from Mast Cells and Development of Flow Cytometry for Glucose Transporters***

**Research support** - Student-Faculty Collaborative Research Grant, The University Honors Program, East Tennessee State University. Project Title: "Effects of catecholamines on pro-atherogenic chemokine production from mast cells," December 7, 2004- June 30, 2005.

**Teaching and Mentorship experience:**

Mary Kate Macedonia, M.S., Predoctoral Candidate, Vanderbilt University, 2023

Jason Tran, CSBC Summer Undergraduate Research Student, 2022

Tolu Omokehinde, Ph.D., Postdoctoral Fellow, Vanderbilt University, 2022

Megan Reed, Ph.D., Postdoctoral Fellow, UAMS Health Sciences Innovation and Entrepreneurship Training Program, 2021

Michael Quan, CSBC Summer Undergraduate Research Student, 2021

Scott Kamen, M.S. Research Specialist, 2018

Ying Lin, Ph.D., Postdoctoral Associate, 2018

Stefano Cirigliano, Ph.D., Postdoctoral Associate, 2018

Richa Singhania, Ph.D., Postdoctoral Associate, 2018

Ashley Kilcollins, Ph.D., Postdoctoral Associate, 2017-2018

Emily Schenkein, B.S., Research Technician, 2017-2018

Yasumi Nakayama, B.A., Research Technician, 2017-2018

Tarig Magdeldin, Ph.D., Postdoctoral Associate, 2016-2018  
 Arash Samadi, Medical Student, Weill Cornell Medicine, Summer 2016  
 Ben Jaffe, Undergraduate Volunteer, Summer 2016  
 Jianxue Li, M.D. Ph.D., Research Technician, 2015-2017  
 Ken Miyaguchi, Ph.D., Postdoctoral Associate, 2015-Present  
 Bethany Schaffer, Ph.D., Postdoctoral Associate, 2015-2016  
 Andrew Storaska, Ph.D., Postdoctoral Associate, 2014-2016  
 Jayson Bastien, Ph.D., Postdoctoral Associate, 2013-2016  
 Demis Balamatsias, Ph.D., Postdoctoral Associate, 2013-2015  
 Tara Jennings, NYU Post-Baccalaureate Program, 2013-2014  
 Jie Sun, Ph.D. Candidate, New York University School of Medicine, 2013  
 Rachael Chase, Vanderbilt University School of Medicine Emphasis Program, 2009  
 Stephen Schleicher, HHMI Medical Research Fellows Program,  
 Vanderbilt University School of Medicine, 2009

### Research Support:

1. TIPs Novel Ideas in Neuroscience (Vanderbilt Brain Institute), (PIs: Linkous and Kavalali). “Brain Organoid Platform for Synaptogenesis in Metastatic Brain Tumors.” February 9, 2023-June 30, 2023.
2. 3U54 CA217450-03S1 (NIH/NCI) Linkous (Investigator). “Brain Metastasis and Cerebral Organoids: Identifying SCLC Tumor Subtypes in Metastatic Disease.” April 1, 2020-March 31, 2021.
3. U54 CA217450 (NIH/NCI), Quaranta (PI). “Phenotype Heterogeneity and Dynamics in SCLC.” Role: Scientific Center Manager. April 13, 2018-March 31, 2023.
4. U01 CA215845 (NIH/NCI), Quaranta (PI), MPI-Lopez. “Phenotype Transitions in Small Cell Lung Cancer.” June 9, 2017-May 31, 2022.
5. Vanderbilt University Graduate Student Grant. “Cytosolic Phospholipase A2: A novel molecular target for tumor sensitization to radiation therapy.” April 18-22, 2009.
6. Scholar-in-Training Travel Award, Radiation Research Society. “Cytosolic phospholipase A2-dependent lysophosphatidylcholine (LPC) production and signaling mediates immediate response to 3 Gy in endothelial cells.” September 21-24, 2008.
7. Vanderbilt University Graduate Student Grant. “Cytosolic Phospholipase A2- Dependent Signaling Protects Normal Cells from Radiation-Induced Cytotoxicity.” October 28-November 1, 2007.
8. Scholar-in-Training Travel Award, International Congress of Radiation Research. “Inhibition of cytosolic phospholipase A2 (cPLA<sub>2</sub>) leads to decreased function in irradiated vascular endothelium.” July 8-12, 2007.
9. Scholar-in-Training Travel Award, Radiation Research Society. “Cytosolic phospholipase A2-dependent lysophosphatidylcholine (LPC) production and signaling mediates immediate response to 3 Gy in endothelial cells.” November 5-8, 2006.

10. Student-Faculty Collaborative Research Grant, The University Honors Program, East Tennessee State University. “Effects of catecholamines on pro-atherogenic chemokine production from mast cells,” December 7, 2004-June 30, 2005. (Faculty Advisor: David S. Chi, Ph.D.)

#### **Media Appearances and Invited Speaking Engagements:**

1. “Organoids as a Model for Small Cell Lung Cancer Brain Metastasis.” Oral Presentation at the NCI’s CSBC Annual Investigators Meeting. Denver, Colorado, March 15-17, 2023.
2. “Precision Medicine: How Cutting Edge Research Unlocks the Promise of More Effective Cancer Treatments.” Corning Life Sciences, Video, December 14, 2022.
3. “Newer 3D Lung Models Starting to Remake Research.” *CHEST Physician*, September 28, 2022.
4. “Q&A: Creating Opportunities for Women in STEM.” Corning Life Sciences *Nucleus* Blog. July 15, 2021.
5. ARIA Series: Evolving Targeted Therapies for Cancer Workshop, Oak Ridge National Laboratory, September 17, 2020. “Using Mini-brains to Improve Outcomes for Brain Tumor Patients.”
6. Biochemistry and Molecular Biology Faculty Seminar Series, University of Arkansas for Medical Sciences, Little Rock, AR, October 2, 2019. “3D Brain Tumor Modeling using Cerebral Organoids.”
7. **Linkous, Amanda.** “Cerebral Organoids as a 3D Model for Glioma.” September 26, 2019. Corning Life Sciences Webinar.
8. Corning Life Sciences Presentation at the Annual Meeting of the Society for Laboratory Automation and Screening (SLAS) Conference, Washington D.C., February 4, 2019. “Spheroids and Mini Brains for Studying Glioblastoma.”
9. Corning Life Sciences 3D Cell Culture Summit, Boston, MA, October 16-17, 2018. “Cerebral Organoids as a 3D Model for Glioma.” Served as Invited Speaker and Panelist.
10. **Linkous, Amanda.** “Editorial Article: Glioma Stem Cells Enable the Production of 3D Human “Mini Brain” to Improve Glioblastoma Treatment.” *SelectScience*, July 25, 2018.
11. **Linkous, Amanda** and Howard Fine. “Brain Organoids Get Cancer, Too, Opening a New Frontier in Personalized Medicine.” *STAT*, December 1, 2017.
12. **Linkous, Amanda** and Howard Fine. “Research Trial in NYC May Change Course of Brain Tumor Treatment.” *NYI News*, August 23, 2017.
13. **Linkous, Amanda** and Howard Fine. “Human ‘Mini Brains’ Grown in Labs May Help Solve Cancer, Autism, Alzheimer’s.” *CNN, Pioneers*, October 22, 2015.
14. Meyer Cancer Center Group Meeting, Weill Cornell Medical College, New York, New York,

September 16, 2015. “Cerebral Organoids and Glioma Stem Cells: A Better Model for Brain Tumor Biology.”

15. Internal Medicine Research Seminar Series, ETSU Quillen College of Medicine, Johnson City, Tennessee, November 26, 2013. “Implications for Metabolic Signaling in Glioma Stem Cell Therapy.”
16. St. Jude National Graduate Student Symposium, Memphis, Tennessee, March 31-April 4, 2009. “Cytosolic Phospholipase A2: A novel molecular target for tumor sensitization to radiation therapy.”
17. Vanderbilt University Annual Host-Tumor Interactions Program and Department of Cancer Biology Retreat, Lake Barkley, Kentucky, November 21-22, 2008. “Cytosolic Phospholipase A2: Targeting cancer through the ablation of tumor vasculature.”
18. Annual Meeting of the Radiation Research Society, Boston, Massachusetts, September 21-24, 2008. “Cytosolic Phospholipase A2: Targeting cancer through the ablation of tumor vasculature.”
19. Annual Meeting of the American Society for Therapeutic Radiology and Oncology, Los Angeles, California, October 28-November 1, 2007. “Cytosolic Phospholipase A2- Dependent Signaling Protects Normal Cells from Radiation-Induced Cytotoxicity.”
20. Annual Meeting of the Radiation Research Society, Philadelphia, Pennsylvania, November 5-8, 2006. “Cytosolic phospholipase A2-dependent lysophosphatidylcholine (LPC) production and signaling mediates immediate response to 3 Gy in endothelial cells.”

## Bibliography:

### Abstracts:

1. **Linkous AG.** “Organoids as a Model for Small Cell Lung Cancer Brain Metastasis.” Presented at the NCI’s CSBC Annual Investigators Meeting. Denver, Colorado, March 15-17, 2023.
2. **Linkous AG.** “Cerebral Organoids as a Systems Platform for SCLC Brain Metastasis and Plasticity.” Presented at the IASLC SCLC Hot Topic Meeting. New York, NY, April 5-7, 2023.
3. **Linkous, AG.** “Using Mini-brains to Improve Outcomes for Brain Tumor Patients.” Presented at the ORNL ARIA Series: Evolving Targeted Therapies for Cancer Workshop. Oak Ridge, Tennessee, September 17-18, 2020.
2. Schleicher SM, **Linkous AG**, Hallahan DE, Yazlovitskaya EM. “LysoPLD plays a role in the radioresistance of malignant glioma through effects on tumor vasculature.” Presented at the American Association for Cancer Research Genetics and Biology of Brain Cancers Conference. San Diego, California, December 13-15, 2009.
3. Chase R, **Linkous AG**, Hallahan DE, Yazlovitskaya EM. “Cytosolic Phospholipase A2 as a novel molecular target for the radiosensitization of ovarian cancer.” Presented at the 55<sup>th</sup> Annual Meeting of the Radiation Research Society, Savannah, Georgia, October 4-7, 2009.

4. **Linkous AG**, Hallahan DE, Yazlovitskaya EM. "Cytosolic Phospholipase A2: A novel molecular target for tumor sensitization to radiation therapy." Presented at the American Association for Cancer Research 2009 Annual Meeting. Denver, Colorado, April 18-22, 2009.
5. **Linkous AG**, Hallahan DE, Yazlovitskaya EM. "Cytosolic Phospholipase A2: A novel molecular target for tumor sensitization to radiation therapy." Presented at the St. Jude National Graduate Student Symposium, Memphis, Tennessee, March 31-April 4, 2009.
6. **Linkous AG**, Geng Ling, Hallahan DE, Yazlovitskaya EM. "Cytosolic Phospholipase A2: Targeting cancer through the ablation of tumor vasculature." Presented at the 54<sup>th</sup> Annual Meeting of the Radiation Research Society, Boston, Massachusetts, September 21-24, 2008.
7. **Linkous AG**, Hallahan DE, Yazlovitskaya EM. "Cytosolic Phospholipase A2 is a Novel Molecular Target for Anti-Angiogenesis Therapy." Presented at the American Association for Cancer Research 2008 Annual Meeting. San Diego, California, April 12-16, 2008.
8. **Linkous AG**, Cuneo KC, Hallahan DE, Yazlovitskaya EM. "Cytosolic Phospholipase A2-Dependent Signaling Protects Normal Cells from Radiation-Induced Cytotoxicity." Presented at the 49<sup>th</sup> Annual Meeting of the American Society for Therapeutic Radiology and Oncology, Los Angeles, California, October 28-November 1, 2007.
9. **Linkous AG**, Cuneo KC, Lyshchik A, Hallahan DE, Yazlovitskaya EM. "Inhibition of cytosolic phospholipase A2 (cPLA<sub>2</sub>) leads to decreased function in irradiated vascular endothelium." Presented at the 13<sup>th</sup> International Congress of Radiation Research. San Francisco, California, July 8-12, 2007.
10. **Linkous AG**, Cuneo KC, Thotala DK, Hallahan DE, Yazlovitskaya EM. "Cytosolic phospholipase A2 regulates viability and function of irradiated vascular endothelial cells." Presented at the Vanderbilt-Ingram Cancer Center Retreat. Nashville, Tennessee, May 4, 2007.
11. **Linkous AG**, Cuneo KC, Thotala DK, Hallahan DE, Yazlovitskaya EM. "Cytosolic phospholipase A2 regulates viability and function of irradiated vascular endothelial cells." Presented at the American Association for Cancer Research 2007 Annual Meeting. Los Angeles, California, April 14-18, 2007.
12. Yazlovitskaya EM, Cuneo KC, **Linkous AG**, Hallahan DE. "Low Dose of Radiation (3 Gy) Initiates an Immediate Signal Transduction through Lysophosphatidylcholine (LPC), a Novel Second Messenger Generated by Cytosolic Phospholipase A2, Which Regulates Response in Vascular Endothelium." Presented at the 48<sup>th</sup> Annual Meeting of the American Society for Therapeutic Radiology and Oncology, Philadelphia, PA November 5-9, 2006.
13. **Linkous AG**, Cuneo KC, Milne SB, Hallahan DE, Yazlovitskaya EM. "Cytosolic phospholipase A2-dependent lysophosphatidylcholine (LPC) production and signaling mediates immediate response to 3 Gy in endothelial cells." Presented at the 53<sup>rd</sup> Annual Meeting of the Radiation Research Society, Philadelphia, PA, November 5-8, 2006.
14. Chi DS, **Mullins AG**, Cantor K, Milhorn D, Krishnaswamy G. "Effects of catecholamines on pro-atherogenic chemokine production from mast cells." Presented at Experimental Biology 2005 meeting.

San Diego, California, April 2-6, 2005.

15. **Mullins AG**, Cantor K, Hall K, Krishnaswamy G, Chi DS. “Enhancing effects of epinephrine on pro-atherogenic chemokine production in IL-1 activated mast cells.” Presented at the 2005 Appalachian Student Research Forum, Johnson City, TN, March 31, 2005. (Division I Undergraduates, 2<sup>nd</sup> Place Award.

## Publications:

### Peer-reviewed journal articles:

1. Quaranta V and **Linkous A**. Organoids as a Systems Platform for SCLC Brain Metastasis. *Frontiers in Oncology*, 2022; Apr 28.
2. Pine A, Cirigliano S, Nicholson J, Hu Y, **Linkous A**, Miyaguchi K, Edwards L, Singhanian R, Schwartz T, Ramakrishna R, Pisapia D, Snuderl M, Elemento O, and Fine H. Tumor Microenvironment Is Critical for the Maintenance of Cellular States Found in Primary Glioblastomas. *Cancer Discovery*, 2020; July.
3. **Linkous A** and Fine H. Generating Patient-Derived Gliomas within Cerebral Organoids. *STAR Protocols* 2020; June 19.
4. **Linkous A**, Sherman H, Li I and Eglen R. Three-Dimensional Organoid Cell Culture: Optimising Disease Models for Research and Drug Discovery. *Drug Discovery World Spring* 2019.
5. **Linkous A**, Balamatsias D, Snuderl M, Edwards L, Miyaguchi K, Milner T, Reich B, Cohen-Gould L, Storaska A, Nakayama Y, Schenkein E, Singhanian R, Cirigliano S, Magdeldin T, Lin Y, Nanjangud G, Chadavalavada K, Pisapia D, Liston C, Fine HA. Modeling Patient-Derived Glioblastoma with Cerebral Organoids. *Cell Reports* 2019; Mar 19;26(12):3203-3211.
6. Baysan M, Woolard K, Cam MC, Zhang W, Song H, Kotliarova S, Balamatsias D, **Linkous A**, Ahn S, Walling J, Belova G, Fine HA. Detailed longitudinal sampling of glioma stem cells in situ reveals Chr7 gain and Chr10 loss as repeated events in primary tumor formation and recurrence. *International Journal of Cancer* 2017; Nov 15; 141(10):2002-2013.
7. Riddick G, Kotliarova S, Rodriguez V, Kim HS, **Linkous A**, Storaska AJ, Ahn S, Walling J, Belova G, Fine HA. A Core Regulatory Circuit in Glioblastoma Stem Cells Links MAPK Activation to a Transcriptional Program of Neural Stem Cell Identity. *Scientific Reports* 2017; Mar 3;7:43605.
8. Bhave SR, Dadey DY, Karvas RM, Ferraro DJ, Kotipatruni RP, Jaboin JJ, Hallahan AN, Dewees TA, **Linkous AG**, Hallahan DE, Thotala D. Autotaxin Inhibition with PF-8380 Enhances the Radiosensitivity of Human and Murine Glioblastoma Cell Lines. *Frontiers in Oncology* 2013; 3:236.
9. **Linkous AG**, Yazlovitskaya EM. Novel radiosensitizing anticancer therapeutics. *Anticancer*

*Research* 2012; 32(7):2487-99.

10. **Linkous AG**, Yazlovitskaya EM. Novel therapeutic approaches for targeting tumor angiogenesis. *Anticancer Research* 2012; 32(1):1-12.
11. Schleicher SM, Thotala DK, **Linkous AG**, Hu R, Leahy KM, Yazlovitskaya EM, Hallahan DE. Autotaxin and LPA receptors represent potential molecular targets for the radiosensitization of murine glioma through effects on tumor vasculature. *PLoS One* 2011; 6(7):e22182.
12. **Linkous AG**, Yazlovitskaya EM. Angiogenesis in glioblastoma multiforme: navigating the maze. *Anticancer Agents Med Chem.* 2011; 11(8):712-8
13. Schulte RR\*, **Linkous AG\***, Hallahan DE, Yazlovitskaya EM. Cytosolic phospholipase A2 as a molecular target for the radiosensitization of ovarian cancer. *Cancer Letters* 2011; 304(2):137-43;\* denotes co-first authorship.
14. **Linkous AG**, Yazlovitskaya EM, Hallahan DE. Cytosolic Phospholipase A2 and Lysophospholipids in Tumor Angiogenesis. *Journal of the National Cancer Institute* 2010; 102(18):1398-412.
15. **Linkous AG** and Yazlovitskaya EM. Cytosolic Phospholipase A2 as a mediator of disease pathogenesis. *Cellular Microbiology* 2010; 12(10):1369-77.
16. **Linkous AG**. PLA2G4A (phospholipase A2, group IVA (cytosolic, calcium-dependent)). *Atlas Genet Cytogenet Oncol Haematol.* December 2009.
17. **Linkous AG**, Geng L, Lyshchik A, Hallahan DE, Yazlovitskaya EM. Cytosolic Phospholipase A2: Targeting Cancer through the Tumor Vasculature. *Clin Cancer Research* 2009; 15:1635-44.
18. Yazlovitskaya EM, **Linkous AG**, Thotala DK, Cuneo KC, Hallahan DE. Cytosolic Phospholipase A2 Regulates Viability of Irradiated Vascular Endothelium. *Cell Death and Differentiation* 2008; 15:1641-53.

#### **Book Chapters:**

1. **Linkous AG** and Yazlovitskaya EM. (2012). New Molecular Targets for Anti-Angiogenic Therapeutic Strategies, *Tumor Angiogenesis*, Sophia Ran (Ed.), ISBN: 978-953-51-0009-6, InTech, Available from: <http://www.intechopen.com/books/tumor-angiogenesis/new-molecular-targets-for-anti-angiogenic-therapeutic-strategies>.